

Review

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# DNA damage and metabolic mechanisms of cancer drug resistance

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## Abstract

Cancer drug resistance is one of the main barriers to overcome to ensure durable treatment responses. While many pivotal advances have been made in first combination therapies, then targeted therapies, and now broadening out to immunomodulatory drugs or metabolic targeting compounds, drug resistance is still ultimately universally fatal. In this brief review, we will discuss different strategies that have been used to fight drug resistance from synthetic lethality to tumor microenvironment modulation, focusing on the DNA damage response and tumor metabolism both within tumor cells and their surrounding microenvironment. In this way, with a better understanding of both targetable mutations in combination with the metabolism, smarter drugs may be designed to combat cancer drug resistance.

**Keywords:** Cancer drug resistance, drug resistance, metabolism, DNA damage, DNA repair, hypoxia, synthetic lethality, overcoming resistance



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## INTRODUCTION

Over the past decades, our understanding of cancer as a disease has increased immensely. The realization of using DNA damaging agents to inhibit the growth of fast-dividing cells with chemotherapy was a game-changing step in treating many types of cancers<sup>[1]</sup>. However, as cancer cells are in a more plastic state with increased genomic instability, resistance to single-agent chemotherapy became prevalent. Therefore, following the steps of infectious disease protocols, combination therapies have evolved to combine multiple chemotherapeutic agents to elicit a longer-lasting effect<sup>[2]</sup>. While this is more beneficial than single-agent treatments, drug/therapy resistance in cancer is still inevitable and universally fatal<sup>[3]</sup>.

Targeted therapies have emerged to combat the rapid drug resistance of broad DNA damaging chemotherapy compounds, which use our increased knowledge of specific vulnerabilities in different types of cancers<sup>[4]</sup>. Success has been seen targeting specific proteins, such as BCR-ABL, the estrogen receptor<sup>[5]</sup>, the androgen receptor (AR)<sup>[6]</sup>, HER2<sup>[7]</sup>, the epidermal growth factor (EGFR)<sup>[8]</sup>, and others. More recently, targeting the immune system via checkpoint inhibitors, like PD1/PD-L1<sup>[9]</sup> and CTLA4<sup>[10]</sup>, have produced cures in a subset of patients. Nevertheless, in patients where a cure cannot be achieved with targeted or conventional chemotherapy, cancers will recur and become drug-resistant which ultimately leads to patients' death. Therefore, drug resistance is the principal limiting factor in patient overall survival.

In this review, we discuss relevant resistance mechanisms tumor cells use to adapt to both chemotherapy and targeted therapies. Furthermore, we summarize some of the promising avenues that are currently being investigated to target the tumor resistance pathways and mutations that arise from the treatments. Overall, a better mechanistic insight into drug-resistant cells will hopefully allow for smarter drug design to help combat the major problem of drug resistance and extend patient survival.

## MECHANISMS OF RESISTANCE

### Changes in the drug-induced DNA damage response

DNA damage continues to be an effective target for cancer therapy as the definition of cancer is uncontrolled cell growth<sup>[4]</sup>. In this way, increased cell cycling can lead to more error-prone DNA replication, which relies on DNA damage repair pathways to ensure cell fitness. Therefore, targeting DNA replication via chemo- and radio-therapy to induce DNA damage and ultimately cell death is still the most common - and sometimes the most effective - in cancer treatment<sup>[11]</sup>. However, while chemo- and radio-therapy can be initially successful, therapy resistance in cancer is common and ultimately fatal. For this reason, extensive efforts have been focused on both determining and targeting the protein or pathways involved in chemo- and radio-resistance.

In glioblastoma (GBM) - an extremely deadly brain cancer with a ~14-16-month median survival rate - the standard of care includes radiation therapy (RT), maximal surgical resection, and the chemotherapeutic agent temozolomide (TMZ)<sup>[12]</sup>. The mechanism of action of TMZ was later shown to create O<sup>6</sup>-methylguanine adducts, which would create double-strand breaks (DSBs) post replication. However, TMZ-resistance is rapid and was found to be partly due to the DNA damage repair protein O<sup>6</sup>-methylguanine-DNA methyltransferase (MGMT)<sup>[13]</sup>. MGMT is the suicide DNA repair protein responsible for removing the O<sup>6</sup>-methylguanine adducts and allowing for its damage repair over DSB formation and cell death<sup>[14]</sup>. In this way, MGMT inhibitors have been reported that inhibit the function of MGMT. MGMT inhibition has also been shown, to reverse pancreatic tumor gemcitabine resistance via suppressing the expression of survivin in animal models<sup>[15]</sup>. However, little clinical success has been realized with MGMT inhibitors with or without TMZ treatment<sup>[16]</sup>.

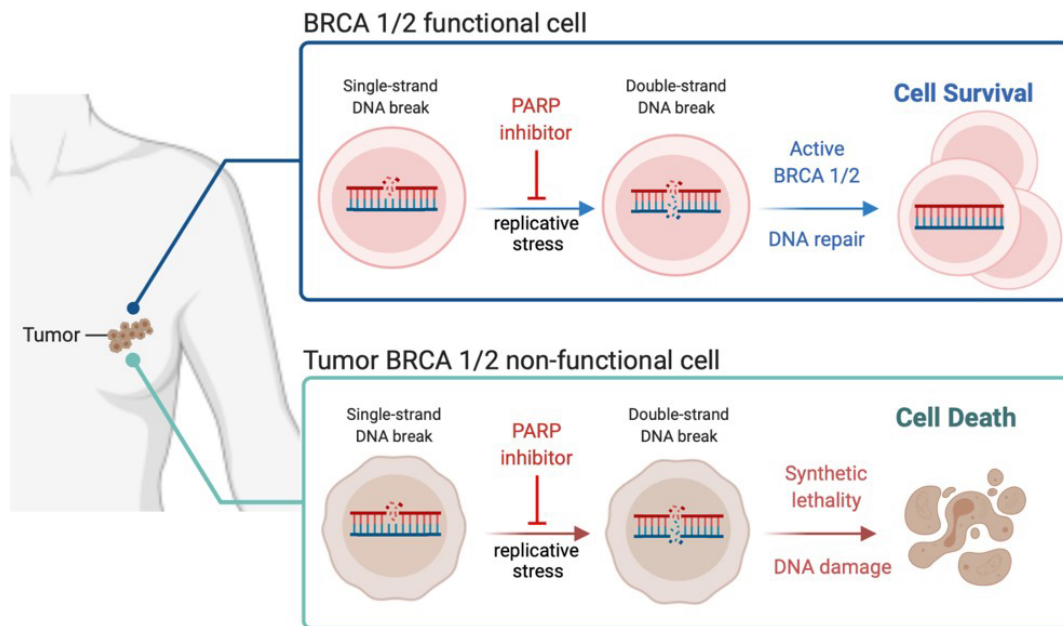
DNA-protein kinase (DNA-PK) is a DSB DNA damage sensing complex composed of Ku70, Ku80, and the DNA protein kinase catalytic subunit (DNA-PKcs)<sup>[17]</sup>. After a DSB has occurred, DNA-PK binds to the broken end of the DSB to protect it from nuclease degradation and recruits the other DNA damage repair proteins to initiate DSB repair via non-homologous end joining (NHEJ)<sup>[18]</sup>. RT has been shown to induce DSBs of which DNA-PK can identify and repair with NHEJ, thereby preventing radiation-induced cell death<sup>[19]</sup>. Cells which have a decreased expression of DNA-PKcs have also been shown to be more sensitive to RT<sup>[20]</sup>. In this way, many DNA-PKcs inhibitors have been developed to overcome radiation resistance and/or enhance radiation-induced cell death. Radio-sensitization via DNA-PKcs inhibition has been observed with DNA-PK inhibitor VX-984 in GBM<sup>[21]</sup>, NU7441 in cervical and breast cancers<sup>[22]</sup>, and NU7026 in non-small cell lung cancer<sup>[23]</sup>. Another DNA-PK inhibitor, NU5455, has also been shown to enhance the killing of doxorubicin in lung cancer models<sup>[24]</sup>.

Lastly, it has been shown that chemotherapy can modulate pro-survival pathways like increasing the expression of drug efflux pumps, apoptosis defects, DNA adduct tolerance, cellular detoxification, and inducing a hypoxic environment<sup>[25]</sup>. While the metabolic effects of hypoxia will be discussed in more detail below, Chen *et al.*<sup>[26]</sup> have taken advantage of intracellular hypoxia in cisplatin-resistant cells and created a hypoxia-amplifying DNA repair-inhibiting (HYDRI) nanomedicine. HYDRI specifically targeted cancer cells because of their drug-induced hypoxic environment, and then released its payload of hypoxia-activatable chemotherapeutic tirapazamine. In this way, previous studies which determined upregulated pathways in cisplatin resistance could be used to create a smarter therapy that bypassed drug efflux pumps, induced a unique DNA damage profile, and relied on the inevitable hypoxic environment created by cisplatin resistance to target these drug-refractory models<sup>[26]</sup>.

### Synthetic lethality

In 2005, the idea of synthetic lethality in the DNA damage repair pathway, via BRCA1 mutation, was published by two groups<sup>[27,28]</sup>. Poly ADP ribose polymerase (PARP) 1 is a key activating protein in the single-strand break (SSB) or base excision repair DNA damage repair pathway<sup>[29]</sup>. In parallel, a double-strand break (DSB) can be fixed by two major DNA damage repair pathways - homologous recombination (HR) or NHEJ. HR utilizes a sister chromatid and has a lower error rate, with BRCA1/2 playing a major activating role for proper DNA damage repair, whereas NHEJ is more error-prone, but quicker in repairing a DSB in interphase<sup>[30]</sup>. Upon the advent of genetic profiling of tumors, it was discovered that many breast and ovarian cancers had either germline or tumor-specific mutations in the *BRCA1* or *BRCA2* gene<sup>[27]</sup>. With the knowledge that SSBs that were unresolved by PARP1 became DSBs with replicative stress that requires BRCA1/2 for repair, the idea of cancer DNA damage response (DDR) synthetic lethality was tested<sup>[31]</sup> [Figure 1]. PARP1 inhibition (PARPi) in BRCA1/2 mutated tumors has been successful in many types of cancers, including breast, ovarian, prostate, pancreatic, colon, and lung<sup>[32]</sup>. One of the main advantages of tumor specific BRCA mutations is the decreased toxicity of single-agent PARPi treatment. Combination therapies to mimic this synthetic lethality by combining both a BRCA inhibitor with PARPi have proven to be toxic<sup>[33]</sup>.

Currently, there are at least five PARPi - veliparib (Abbvie), rucaparib (Pfizer/Clovis), Olaparib (KuDOS/AstraZeneca), niraparib (Merck/Tesaro), and talazoparib (Lead/Biomarin/Medivation/Pfizer)<sup>[32]</sup> - where the most common mechanism of action is the “trapping” of PARP on the DNA to induce DSBs<sup>[34]</sup>. Talazoparib is the newest PARPi and has the highest ability to “trap” PARP on the DNA with ~100 times greater efficacy than the next best PARPi<sup>[35]</sup>. Nevertheless, this increased trapping of PARP increases the toxicity of talazoparib - compared to other PARPi<sup>[36]</sup>. In clinical results, the phase 3 trial EMBARCA had 431 BRCA1/2 mutant patients with advanced breast cancer, where the talazoparib group had a 62.6% response rate compared to the 27.2% of the standard chemotherapy group<sup>[37]</sup>.



**Figure 1.** PARPi synthetic lethality in BRCA 1/2 non-functional tumors. BRCA 1/2 functional tumor cells will repair the double-strand break (DSB) induced by PARP inhibition and sequential replicative stress allowing cell survival and growth, whereas BRCA 1/2 non-functional cells cannot repair the DSB and therefore succumb to DNA damage-induced cell death. Figure created with BioRender.

The synthetic lethality success story has other groups actively looking for other synthetic lethal interactions with the many abundant cancer-associated mutations. While not dependent on DDR mutations, AR signaling and PARPi have also been shown to give rise to a synthetic lethal phenotype in preclinical models. However, it is important to note that 19% of primary prostate cancers and 23% of metastatic castration-resistant prostate cancer (mCRPC) have DNA damage repair gene mutations. BRCA2 mutations also have increased levels of prostate specific antigen, a larger percentage of high Gleason scored tumors, and elevated rates of distant and nodal metastases<sup>[38]</sup>. Clinically, prostate cancer patients treated with abiraterone plus olaparib showed improved radiographic progression free survival over abiraterone alone in a phase 2 trials which was independent of DDR mutations<sup>[39]</sup>. This has also led to an interest in combining PARPi with androgen deprivation therapy alone or in combination with AR signaling inhibitors, which is currently ongoing in mCRPC, and should be considered even in a non-DDR altered state<sup>[38]</sup>.

### DNA damage response with immunotherapy

Immune checkpoint inhibitors (ICI) had great initial promise, with early clinical trial results showing obvious tumor shrinkage, initially. However, after further evaluation, ICI can have about a ~10%-20% durable response rate, depending on the types of cancer<sup>[40]</sup>. Therefore, like what we previously described with chemotherapy and targeted therapy, combinatorial studies have been designed to determine whether ICI efficacy can be improved when combined with other conventional therapies<sup>[41,42]</sup>.

Melanoma was the first cancer to show preliminary success with ICI, and it is well-known that melanoma has one of the highest rates of tumor mutation burden (TMB)<sup>[43]</sup>. This brought about the hypothesis that higher rates of TMB in cancer would increase the number of neo-antigens which were predicted to produce a stronger immune response and increase sensitivity to ICI. Radiation and ICI have been tested in combination as radiation treatment for cancer will induce DNA damage, neo-antigens, and immune response<sup>[44]</sup>. Furthermore, it was shown in 1979 that the effect of radiation is linked to the immune system when twice the dose of radiation was needed to control tumor growth in thymectomized mice compared to

mice with an intact immune system<sup>[45]</sup>. Within the clinic, overall survival of concurrent radiation and ICI - compared to radiation before or after ICI - was shown to improve overall survival (OS) in a retrospective review of lung cancer patients with distant brain metastases<sup>[46]</sup>.

Other chemotherapy agents are currently being combined with ICI to determine combinatorial efficacy, where one combination - PARPi + ICI - is the most developed. PARPi have shown great promise in many avenues, as shown above with synthetic lethality strategies<sup>[47]</sup>. With the increasing characterization of PARPi pathway changes, it was noted in breast cancer that PARPi induced PD-L1 expression<sup>[48]</sup>. Not only has PD-L1 been shown to increase with PARPi, but also an increase in cytoplasmic DNA, which activates the cyclic guanosine monophosphate-AMP synthase (cGAS)/stimulator of interferon genes (STING) pathway<sup>[47]</sup>. PARPi has also been shown to inactivate the glycogen synthase kinase 2 beta (GSK3)<sup>[36]</sup>. An *in vivo* model of BRCA-deficient triple-negative breast cancer also demonstrated that PARPi activated the cGAS/STING pathway and increased CD8<sup>+</sup> T cell infiltration<sup>[48]</sup>, as well as decreasing T-cell activation resulting in enhanced cancer cell apoptosis<sup>[36]</sup>. Currently, biomarkers are being investigated to select non-BRCA patients that would respond to PARPi + ICI, where the mutational signature 3 - associated with HR deficiency - positively predicted patient responses<sup>[49]</sup>. Both the TOPACIO trial and MEDIOLA study are investigating feasibility of immune checkpoint blockade with PARPi<sup>[36]</sup>.

### Reactive oxygen species in drug resistance

Reactive oxygen species (ROS) play a well-known role in cell growth and proliferation in cancer cells, where an increase in ROS can enhance cell growth and post-treatment survival<sup>[50]</sup>. One way in which ROS can increase cancer cell survival is genetically by oxidizing nucleic acids, which will cause random mutations and increase genomic instability<sup>[51,52]</sup>. ROS can also affect the normal redox balance within the cell. Cysteine is a readily oxidizable amino acid containing a thiol (-SH) group<sup>[53]</sup>. Many enzymes have active sites that contain necessary cysteine residues to assist in biochemical reactions<sup>[54]</sup>. In Fms-related receptor tyrosine kinase (FLT)3-ITD (a mutation in the tyrosine kinase domain) expressing acute myeloid leukemia, NADPH oxidase 4 generated ROS will inactivate the protein-tyrosine phosphatase (PTP) DEP-1/PTPRJ, which negatively regulates FLT3-ITD transformation<sup>[55]</sup>. PTP phosphatase and tensin homolog (PTEN), a PTP family member and potent tumor suppressor, has also been shown to be susceptible to H<sub>2</sub>O<sub>2</sub>-mediated oxidation and inactivation<sup>[56]</sup>. As PTEN is a negative regulator of PI3K and Akt pathways, oxidation and inactivation of PTEN augments downstream signaling and cell growth<sup>[57]</sup>.

While ROS have been used to create DNA damage via chemotherapy and are necessary byproducts of radiation, drug-resistant cells have been shown to increase their intracellular ROS levels, and thereby adapt to this intracellular hypoxic environment<sup>[58]</sup>. Initially, when naïve cells are exposed to chemotherapy, an increase in ROS is noted, as well as a concomitant increase in the antioxidant systems to combat this onslaught of oxidants<sup>[58]</sup>. However, this seems to differ in some drug-resistant, or persister, cells where an increase in ROS is still true, but antioxidant genes like glutathione peroxidase (GPX) 4 are now downregulated<sup>[59]</sup>. One hypothesis is that these cells use this ROS to their advantage as the hypoxia response element (HRE) shows a higher binding of HIF1 when the G's of the HRE are modified via ROS<sup>[60]</sup>. Accordingly, in a new study which dives into the metabolic and transcriptional changes between untreated and persister cells post drug treatment, two of the main upregulated pathways in their model, osimertinib-treated Trp53-knockout with a lung-specific EGFR (L858R) mutation, were ROS and fatty acid metabolism (FAM)<sup>[61]</sup>. Furthermore, the cells that had the increased ROS/FAM gene signature were also the cycling persister cells, compared with the non-cycling persister cells. In post-treatment patient samples, 8 out of 11 melanoma samples had an increase in either ROS or FAM signatures, as well as 50% of HER2<sup>+</sup> breast cancer samples, but the increase in HER2<sup>+</sup> samples was only in the post-treatment samples<sup>[61]</sup>.



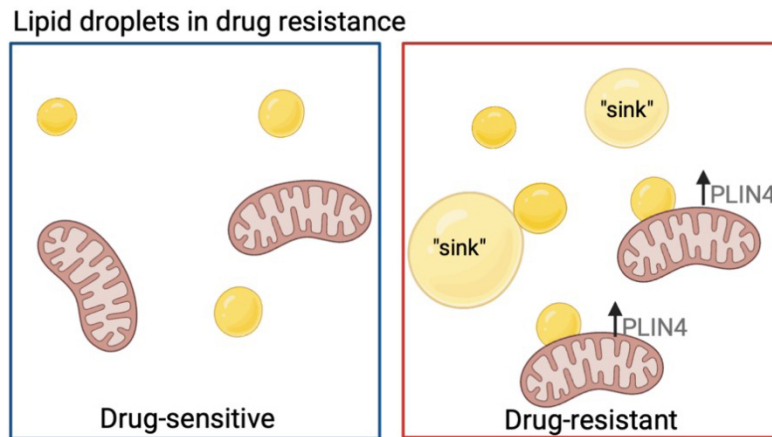
In these ways, ROS-induced metabolic reprogramming has been an active non-mutational target. The most promising area of research is ferroptosis - or iron-dependent cell death - induction<sup>[62]</sup>. As drug-resistant cells have been shown to increase ROS and decrease their antioxidant gene expression, this leaves the cells exquisitely sensitive to ferroptosis via GPX4 or xCT inhibition<sup>[59]</sup>. GPX4 is the main regulator to decrease lipid oxidation, and xCT is a cystine/glutamate antiporter, where cysteine is a necessary component for the reducing agent glutathione<sup>[63]</sup>. While ferroptosis induction has shown promising initial results in drug-resistant cell and animal models, most drugs never reach the target in cancers such as pancreatic cancer. In this largely drug-refractory cancer, Badgley *et al.*<sup>[64]</sup> showed that a cyst(e)ine degrading enzyme - cyst(e)inase - was able to deplete cyst(e)ine from the extracellular environment, thereby decreasing intracellular cyst(e)ine and preventing GSH production. In both *in vitro* and *in vivo* models, cyst(e)ine deprivation led to robust induction of ferroptosis, cell death, and longer animal survivals<sup>[64]</sup>.

### Drug resistance via lipid metabolism and import

Fatty acid (FA) metabolism plays a host of roles within the cell. FAs may be most well-known for being membrane building blocks with the synthesis of glycerophospholipids<sup>[65]</sup>. Interestingly, the lipid composition of membranes has garnered recent interest as chemotherapy-resistant cancer cells, in preclinical models, show reduced fluidity of their membranes. These membranes have an increase in saturated fatty acyl chains and are especially enriched for monounsaturated fatty acyl chains in glycerophospholipids<sup>[66]</sup>. While this may seem inconsequential, one of the most promising therapeutic targets of drug-resistant cells - ferroptosis - can depend on poly-unsaturated fatty acyl chains to induce toxic lipid peroxidation and cell death<sup>[67]</sup>. Furthermore, in chemotherapy-resistant leukemia or ovarian cancer cell lines, the reduced membrane fluidity can come from an increase of cholesterol and/or sphingomyelin within the membrane<sup>[68,69]</sup>. This stiffened membrane has been shown to decrease passive diffusion of drug uptake and enhance detergent-resistant membrane domains, which can activate the family of ATP-binding cassette multidrug efflux transporters - including p-glycoprotein - potentiating the multidrug-resistant (MDR) phenotype. However, modulation of membrane fluidity was able to alter the drug efflux transporters, suggesting a potential for diet interventions<sup>[70]</sup>.

Targeting lipid synthesis itself may also have a benefit in re-sensitizing cells to chemotherapy. Fatty acid synthase (FAS) inhibitors have re-sensitized ovarian cells *in vitro*<sup>[71,72]</sup>, *ex vivo*<sup>[71]</sup>, and *in vivo* for T cell lymphoma and ovarian cancer models<sup>[73,74]</sup>. In breast cancer cells, overexpression of FAS was able to confer chemoresistance *in vitro*<sup>[72]</sup>. While the mechanism by which FAS inhibition can alter chemotherapy sensitivity is unknown, a decrease in MDR proteins has been observed, suggesting membrane composition may be important<sup>[75]</sup>. Fatty acid oxidation has also been shown to increase with chemotherapy resistance. GBM cellular and patient-derived xenograft models showed an increase in fatty acid beta-oxidation post-TMZ treatment<sup>[76]</sup>. In breast cancer patient samples, the necessary beta-oxidation enzyme carnitine palmitoyltransferase I (CPT1) was increased in tumors that recurred and was also higher in chemo-resistant tumors<sup>[77]</sup>. CPT1 inhibitors have also been shown to re-sensitize tumor cells to chemotherapeutic agents<sup>[78]</sup>.

Lastly, lipid droplet (LD) number and function play an important role in chemotherapy resistance. LDs may directly assist in cell survival by providing an energy reserve of lipids to be oxidized in case of nutrient deprivation<sup>[79]</sup>. Hydrophobic drugs can also be sequestered within lipid droplets, creating a drug “sink” for detoxifying chemotherapeutic agents<sup>[79]</sup>. Interestingly, LDs were found to co-localize with the mitochondria more frequently in chemo-resistant breast cancer cells, where the LD protein perilipin 4 (PLIN4) was increased. PLIN4 assists in mobilizing lipids for oxidation from LDs, where silencing of PLIN4 decreased the growth of the chemotherapy-resistant, but not the parental, breast cancer cells, suggesting that lipid beta-oxidation is necessary for the sustained growth of chemotherapy-resistant cells<sup>[80]</sup> [Figure 2]. Inhibition of long-chain fatty acyl-CoA synthetase via triacsin C blocked fatty acid activation and LD biogenesis which



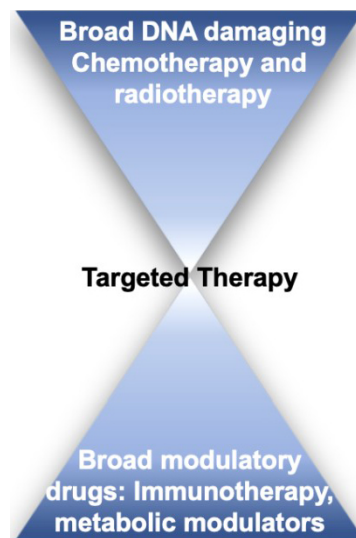
**Figure 2.** Lipid droplet usage in drug-resistant cells. Drug-resistant cells have been shown to increase lipid droplet accumulation (yellow circles) and have a higher percentage co-localized to the mitochondria, where an increase in PLIN4 helps to better utilize lipids for fatty acid beta-oxidation. Lipids can also be used as drug “sinks” for hydrophobic drugs. Figure created with BioRender.

rendered drug-resistant colorectal cancer cells sensitive to chemotherapy treatment both *in vitro* and *in vivo*<sup>[81]</sup>.

### Hypoxia and drug resistance

The 2019 Nobel Prize was awarded to Kaelin, Ratcliffe, and Semenza for their seminal work on discovering how oxygen is sensed in the cell and the way in which cells are able to adapt to changing oxygen concentrations<sup>[82,83]</sup>. Hypoxia-inducible factor (HIF) and HIF signaling are now realized as both a hallmark of cancer, and also affect the surrounding microenvironment<sup>[4]</sup>. Through large data analyses, Bhandari *et al.*<sup>[84]</sup> used the Buffa-defined hypoxia signature to determine the breadth of hypoxia amongst 1188 samples from 27 types of cancer. They found that hypoxia was varied both between cancers and even within a single patient. The most hypoxic tumors were cervical squamous cell carcinoma and lung cancer, with thyroid adenocarcinoma and chronic lymphocytic leukemia being the least hypoxic<sup>[84]</sup>. This study corroborated the findings of The Cancer Genome Atlas. Furthermore, higher hypoxia scores also correlate to both lower overall survival and progression-free survival in multiple cancer types<sup>[85]</sup>. A forced hypoxic environment in GBM models has also been shown to decrease the sensitivity of cells to the standard of care chemotherapeutic agent TMZ<sup>[86]</sup>.

As cancer cells grow faster than normal cells, they quickly outgrow their nutrient supply, which creates a lower level of oxygen in the tumor microenvironment (TME). This low oxygen, or hypoxic TME, affects gene signatures and pathways activated within the tumor cells<sup>[87]</sup>. Carbonic anhydrases and CO<sub>2</sub> levels are increased, which leads to increased cellular acidification<sup>[88]</sup>. However, cancer cells depend on a higher intracellular pH (~7.4 *vs.* ~7.2) and acidify their TME, decreasing the extracellular pH (~6.0~7.1 *vs.* ~7.4)<sup>[89]</sup>. This acidic niche has been shown to increase the expression of MDR genes and decrease drug import into cancer cells<sup>[90]</sup>. A highly acidic TME can also prevent proper immune profusion and cause resistance to ICI<sup>[91]</sup>. Acidity can increase the immune checkpoint protein expression, CTLA-4 on T cells, raise the threshold for T cell activation<sup>[92]</sup>, and decrease CD8<sup>+</sup> memory T cell lifespan<sup>[93]</sup>. Long-term exposure to an acidic environment can decrease natural killer cell function, activation, and survival<sup>[92]</sup>. Therefore, modulation of TME pH has been an active area of research.



**Figure 3.** Therapy schematic. Broad-reaching drugs may lend to more durable responses as resistance can arise more rapidly to targeted therapy. Modulating more broad cancer hallmarks - like immune and metabolic targets - may offer smarter drug targets.

To neutralize the acidic TME, oral bicarbonate was given and artificially increased the pH of the TME, which allowed for a better response to ICI in multiple cancer models<sup>[94]</sup>. In an effort to maintain normal levels of pH,  $\text{Na}^+/\text{H}^+$  exchanges, like sodium-hydrogen exchanger isoform (NHE), are upregulated to uptake sodium and pump out protons<sup>[95]</sup>. The  $\text{Na}^+/\text{H}^+$  exchangers are the most common membrane proteins and attempt to regulate the hypoxia-induced pH changes within the cell<sup>[96]</sup>. As intracellular acidosis can induce necroptosis and apoptosis, NHE1 inhibitors have been found to modulate intracellular pH and lead to cell death<sup>[97]</sup>. Cariporide, an NHE1 inhibitor, can induce apoptosis in breast cancer, reduce MDR1 expression, and decrease tumor volume<sup>[98]</sup>. Another NHE inhibitor, amiloride, increased ROS abundance, thereby stimulating PAR synthesis and inducing the PAR-dependent cell death termed parthanatos<sup>[99]</sup>. In GBM,  $\text{Na}^+$  was shown to be increased almost 3-fold between cancer and normal cells, while NHE1 overexpression is noted and increased NHE1 correlated to worse overall survival<sup>[97]</sup>.

## CONCLUSIONS AND FUTURE PERSPECTIVES

Cancer drug resistance remains the biggest challenge in successfully treating cancer patients today. Here we have outlined some of the current strategies to target the DNA damage repair proteins via chemo-induced dependencies, synthetic lethality, and combination with immunotherapy. More research insight into cancer-specific deficiencies can only lead to better responses, or third-line therapeutic options. Tumor metabolism and the microenvironment may also prove to be promising drug targets as uncontrolled cell growth will always be a hallmark of cancer. Therefore, if we are truly able to understand the metabolic changes and vulnerabilities of cancer, we may be able to develop biomarkers to help dictate metabolic status and treatment plans.

As a whole, we may be better off looking at broad regulatory pathways, as we seem to be at the far end of an hourglass curve [Figure 3]. In the beginning, broad DNA damaging agents were game-changing as they targeted the quintessential cancer dependency - cell growth. Then we narrowed the focus to specific proteins with either activating mutations or cancer-driving functions. While some success has been achieved with targeted therapy, new avenues like modulating the immune system, or using HYDRI-like methods to target broad cancer hallmarks may lead to more durable and smart therapeutic designs.



## DECLARATIONS

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### Authors' contributions

Accepted invitation and developed the manuscript: Tiek D, Cheng SY

Wrote the manuscript: Tiek D

Edited, provided feedback, and administrative support: Cheng SY

### Availability of data and materials

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Both authors declared that there are no conflicts of interest.

### Ethical approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

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## REFERENCES

1. Farber S, Diamond LK. Temporary remissions in acute leukemia in children produced by folic acid antagonist, 4-aminopteroyl-glutamic acid. *N Engl J Med* 1948;238:787-93. [DOI PubMed](#)
2. Crofton J. Chemotherapy of pulmonary tuberculosis. *Br Med J* 1959;1:1610-4. [DOI PubMed PMC](#)
3. Vasan N, Baselga J, Hyman DM. A view on drug resistance in cancer. *Nature* 2019;575:299-309. [DOI PubMed PMC](#)
4. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011;144:646-74. [DOI PubMed](#)
5. Schafer J, Liu H, Levenson AS, Horiguchi J, Chen Z, Jordan V. Estrogen receptor  $\alpha$  mediated induction of the transforming growth factor  $\alpha$  gene by estradiol and 4-hydroxytamoxifen in MDA-MB-231 breast cancer cells. *J Steroid Biochem Mol Biol* 2001;78:41-50. [DOI](#)
6. Huggins C, Hodges CV. Studies on prostatic cancer. I. The effect of castration, of estrogen and androgen injection on serum phosphatases in metastatic carcinoma of the prostate. *CA Cancer J Clin* 1972;22:232-40. [DOI PubMed](#)
7. Hicks DG, Kulkarni S. HER2+ breast cancer: review of biologic relevance and optimal use of diagnostic tools. *Am J Clin Pathol* 2008;129:263-73. [DOI PubMed](#)
8. Fukuoka M, Yano S, Giaccone G, et al. Multi-institutional randomized phase II trial of gefitinib for previously treated patients with advanced non-small-cell lung cancer (The IDEAL 1 Trial) [corrected]. *J Clin Oncol* 2003;21:2237-46. [DOI PubMed](#)
9. Iwai Y, Ishida M, Tanaka Y, Okazaki T, Honjo T, Minato N. Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor immunotherapy by PD-L1 blockade. *Proc Natl Acad Sci U S A* 2002;99:12293-7. [DOI PubMed PMC](#)
10. Leach DR, Krummel MF, Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. *Science* 1996;271:1734-6. [DOI PubMed](#)
11. Shaked Y. The pro-tumorigenic host response to cancer therapies. *Nat Rev Cancer* 2019;19:667-85. [DOI PubMed](#)
12. Killock D. Brain cancer: chemoradiotherapy for low-grade glioma: battle won, but the war goes on. *Nat Rev Clin Oncol* 2016;13:328-9. [DOI PubMed](#)
13. Hegi ME, Diserens AC, Gorlia T, et al. MGMT gene silencing and benefit from temozolomide in glioblastoma. *N Engl J Med* 2005;352:997-1003. [DOI PubMed](#)
14. Yu W, Zhang L, Wei Q, Shao A. O<sup>6</sup>-Methylguanine-DNA methyltransferase (MGMT): challenges and new opportunities in glioma

- chemotherapy. *Front Oncol* 2019;9:1547. DOI PubMed PMC
15. Bobustuc GC, Patel A, Thompson M, et al. MGMT inhibition suppresses survivin expression in pancreatic cancer. *Pancreas* 2015;44:626-35. DOI PubMed
  16. Quinn JA, Jiang SX, Reardon DA, et al. Phase II trial of temozolomide plus o6-benzylguanine in adults with recurrent, temozolomide-resistant malignant glioma. *J Clin Oncol* 2009;27:1262-7. DOI PubMed PMC
  17. Huang RX, Zhou PK. DNA damage response signaling pathways and targets for radiotherapy sensitization in cancer. *Signal Transduct Target Ther* 2020;5:60. DOI PubMed PMC
  18. Blackford AN, Jackson SP. ATM, ATR, and DNA-PK: the trinity at the heart of the DNA damage response. *Mol Cell* 2017;66:801-17. DOI PubMed
  19. Lu H, Saha J, Beckmann PJ, Hendrickson EA, Davis AJ. DNA-PKcs promotes chromatin decondensation to facilitate initiation of the DNA damage response. *Nucleic Acids Res* 2019;47:9467-79. DOI PubMed PMC
  20. Douglas P, Gupta S, Morrice N, Meek K, Lees-Miller SP. DNA-PK-dependent phosphorylation of Ku70/80 is not required for non-homologous end joining. *DNA Repair (Amst)* 2005;4:1006-18. DOI
  21. Timme CR, Rath BH, O'Neill JW, Camphausen K, Tofilon PJ. The DNA-PK inhibitor VX-984 enhances the radiosensitivity of glioblastoma cells grown in vitro and as orthotopic xenografts. *Mol Cancer Ther* 2018;17:1207-16. DOI PubMed PMC
  22. van Oorschot B, Granata G, Di Franco S, et al. Targeting DNA double strand break repair with hyperthermia and DNA-PKcs inhibition to enhance the effect of radiation treatment. *Oncotarget* 2016;7:65504-13. DOI PubMed PMC
  23. Yang L, Liu Y, Sun C, et al. Inhibition of DNA-PKcs enhances radiosensitivity and increases the levels of ATM and ATR in NSCLC cells exposed to carbon ion irradiation. *Oncol Lett* 2015;10:2856-64. DOI PubMed PMC
  24. Willoughby CE, Jiang Y, Thomas HD, et al. Selective DNA-PKcs inhibition extends the therapeutic index of localized radiotherapy and chemotherapy. *J Clin Invest* 2020;130:258-71. DOI PubMed PMC
  25. Drayton RM, Catto JW. Molecular mechanisms of cisplatin resistance in bladder cancer. *Expert Rev Anticancer Ther* 2012;12:271-81. DOI PubMed
  26. Chen J, Wang X, Yuan Y, et al. Exploiting the acquired vulnerability of cisplatin-resistant tumors with a hypoxia-amplifying DNA repair-inhibiting (HYDRI) nanomedicine. *Sci Adv* 2021;7:eabc5267. DOI PubMed PMC
  27. Farmer H, McCabe N, Lord CJ, et al. Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nature* 2005;434:917-21. DOI PubMed
  28. Bryant HE, Schultz N, Thomas HD, et al. Specific killing of BRCA2-deficient tumours with inhibitors of poly(ADP-ribose) polymerase. *Nature* 2005;434:913-7. DOI PubMed
  29. Satoh MS, Lindahl T. Role of poly(ADP-ribose) formation in DNA repair. *Nature* 1992;356:356-8. DOI PubMed
  30. Pilié PG, Tang C, Mills GB, Yap TA. State-of-the-art strategies for targeting the DNA damage response in cancer. *Nat Rev Clin Oncol* 2019;16:81-104. DOI PubMed PMC
  31. Ashworth A, Lord CJ, Reis-Filho JS. Genetic interactions in cancer progression and treatment. *Cell* 2011;145:30-8. DOI PubMed
  32. Lord CJ, Ashworth A. PARP inhibitors: synthetic lethality in the clinic. *Science* 2017;355:1152-8. DOI PubMed PMC
  33. Dréan A, Lord CJ, Ashworth A. PARP inhibitor combination therapy. *Crit Rev Oncol Hematol* 2016;108:73-85. DOI PubMed
  34. Purnell MR, Whish WJ. Novel inhibitors of poly(ADP-ribose) synthetase. *Biochem J* 1980;185:775-7. DOI PubMed PMC
  35. Shen Y, Rehman FL, Feng Y, et al. BMN 673, a novel and highly potent PARP1/2 inhibitor for the treatment of human cancers with DNA repair deficiency. *Clin Cancer Res* 2013;19:5003-15. DOI PubMed PMC
  36. Boussios S, Abson C, Moschetta M, et al. Poly (ADP-Ribose) polymerase inhibitors: talazoparib in ovarian cancer and beyond. *Drugs R D* 2020;20:55-73. DOI PubMed PMC
  37. Litton JK, Rugo HS, Ettl J, et al. Talazoparib in patients with advanced breast cancer and a germline BRCA mutation. *N Engl J Med* 2018;379:753-63. DOI PubMed
  38. Boussios S, Rassy E, Shah S, et al. Aberrations of DNA repair pathways in prostate cancer: a cornerstone of precision oncology. *Expert Opin Ther Targets* 2021;25:329-33. DOI PubMed
  39. Clarke N, Wiechno P, Alekseev B, et al. Olaparib combined with abiraterone in patients with metastatic castration-resistant prostate cancer: a randomised, double-blind, placebo-controlled, phase 2 trial. *Lancet Oncol* 2018;19:975-86. DOI PubMed
  40. Dempke WCM, Fenchel K, Dale SP. Programmed cell death ligand-1 (PD-L1) as a biomarker for non-small cell lung cancer (NSCLC) treatment-are we barking up the wrong tree? *Transl Lung Cancer Res* 2018;7:S275-9. DOI PubMed PMC
  41. Gandhi L, Rodríguez-Abreu D, Gadgeel S, et al; KEYNOTE-189 Investigators. Pembrolizumab plus chemotherapy in metastatic non-small-cell lung cancer. *N Engl J Med* 2018;378:2078-92. DOI PubMed
  42. Antonia SJ, Villegas A, Daniel D, et al; PACIFIC Investigators. Durvalumab after chemoradiotherapy in stage III non-small-cell lung cancer. *N Engl J Med* 2017;377:1919-29. DOI PubMed
  43. McGrail DJ, Pilié PG, Rashid NU, et al. High tumor mutation burden fails to predict immune checkpoint blockade response across all cancer types. *Ann Oncol* 2021;32:661-72. DOI PubMed PMC
  44. Le DT, Durham JN, Smith KN, et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. *Science* 2017;357:409-13. DOI PubMed PMC
  45. Stone HB, Peters LJ, Milas L. Effect of host immune capability on radiocurability and subsequent transplantability of a murine fibrosarcoma. *J Natl Cancer Inst* 1979;63:1229-35. PubMed
  46. Schapira E, Hubbeling H, Yeap BY, et al. Improved overall survival and locoregional disease control with concurrent PD-1 pathway

- inhibitors and stereotactic radiosurgery for lung cancer patients with brain metastases. *Int J Radiat Oncol Biol Phys* 2018;101:624-9. DOI PubMed
47. Sun W, Zhang Q, Wang R, Li Y, Sun Y, Yang L. Targeting DNA damage repair for immune checkpoint inhibition: mechanisms and potential clinical applications. *Front Oncol* 2021;11:648687. DOI PubMed PMC
  48. Pantelidou C, Sonzogni O, De Oliveria Taveira M, et al. PARP inhibitor efficacy depends on CD8<sup>+</sup> T-cell recruitment via intratumoral STING pathway activation in BRCA-deficient models of triple-negative breast cancer. *Cancer Discov* 2019;9:722-37. DOI PubMed PMC
  49. Färkkilä A, Gulhan DC, Casado J, et al. Immunogenomic profiling determines responses to combined PARP and PD-1 inhibition in ovarian cancer. *Nat Commun* 2020;11:1459. DOI PubMed PMC
  50. Panieri E, Santoro MM. ROS homeostasis and metabolism: a dangerous liason in cancer cells. *Cell Death Dis* 2016;7:e2253. DOI PubMed PMC
  51. Pelicano H, Carney D, Huang P. ROS stress in cancer cells and therapeutic implications. *Drug Resist Updat* 2004;7:97-110. DOI PubMed
  52. Szatrowski TP, Nathan CF. Production of large amounts of hydrogen peroxide by human tumor cells. *Cancer Res* 1991;51:794-8. PubMed
  53. Moloney JN, Cotter TG. ROS signalling in the biology of cancer. *Semin Cell Dev Biol* 2018;80:50-64. DOI PubMed
  54. Veal EA, Day AM, Morgan BA. Hydrogen peroxide sensing and signaling. *Mol Cell* 2007;26:1-14. DOI PubMed
  55. Jayavelu AK, Müller JP, Bauer R, et al. NOX4-driven ROS formation mediates PTP inactivation and cell transformation in FLT3ITD-positive AML cells. *Leukemia* 2016;30:473-83. DOI PubMed
  56. Lee SR, Yang KS, Kwon J, Lee C, Jeong W, Rhee SG. Reversible inactivation of the tumor suppressor PTEN by H<sub>2</sub>O<sub>2</sub>. *J Biol Chem* 2002;277:20336-42. DOI PubMed
  57. Leslie NR, Bennett D, Lindsay YE, Stewart H, Gray A, Downes CP. Redox regulation of PI 3-kinase signalling via inactivation of PTEN. *EMBO J* 2003;22:5501-10. DOI PubMed PMC
  58. Cui Q, Wang JQ, Assaraf YG, et al. Modulating ROS to overcome multidrug resistance in cancer. *Drug Resist Updat* 2018;41:1-25. DOI PubMed
  59. Hangauer MJ, Viswanathan VS, Ryan MJ, et al. Drug-tolerant persister cancer cells are vulnerable to GPX4 inhibition. *Nature* 2017;551:247-50. DOI PubMed PMC
  60. Al-Mehdi AB, Pastukh VM, Swiger BM, et al. Perinuclear mitochondrial clustering creates an oxidant-rich nuclear domain required for hypoxia-induced transcription. *Sci Signal* 2012;5:ra47. DOI PubMed PMC
  61. Oren Y, Tsabar M, Cuoco MS, et al. Cycling cancer persister cells arise from lineages with distinct programs. *Nature* 2021;596:576-82. DOI PubMed
  62. Hassannia B, Vandenabeele P, Vanden Berghe T. Targeting ferroptosis to iron out cancer. *Cancer Cell* 2019;35:830-49. DOI PubMed
  63. Dixon SJ, Lemberg KM, Lamprecht MR, et al. Ferroptosis: an iron-dependent form of nonapoptotic cell death. *Cell* 2012;149:1060-72. DOI PubMed PMC
  64. Badgley MA, Kremer DM, Maurer HC, et al. Cysteine depletion induces pancreatic tumor ferroptosis in mice. *Science* 2020;368:85-9. DOI PubMed PMC
  65. Zhu J, Thompson CB. Metabolic regulation of cell growth and proliferation. *Nat Rev Mol Cell Biol* 2019;20:436-50. DOI PubMed PMC
  66. Butler LM, Perone Y, Dehairs J, et al. Lipids and cancer: emerging roles in pathogenesis, diagnosis and therapeutic intervention. *Adv Drug Deliv Rev* 2020;159:245-93. DOI PubMed PMC
  67. Stockwell BR, Friedmann Angeli JP, Bayir H, et al. Ferroptosis: a regulated cell death nexus linking metabolism, redox biology, and disease. *Cell* 2017;171:273-85. DOI PubMed PMC
  68. Veldman RJ, Klappe K, Hinrichs J, et al. Altered sphingolipid metabolism in multidrug-resistant ovarian cancer cells is due to uncoupling of glycolipid biosynthesis in the Golgi apparatus. *FASEB J* 2002;16:1111-3. DOI PubMed
  69. Mannechez A, Reungpatthanaphong P, de Certaines JD, Leray G, Le Moyec L. Proton NMR visible mobile lipid signals in sensitive and multidrug-resistant K562 cells are modulated by rafts. *Cancer Cell Int* 2005;5:2. DOI PubMed PMC
  70. Hajjaji N, Bougnoux P. Selective sensitization of tumors to chemotherapy by marine-derived lipids: a review. *Cancer Treat Rev* 2013;39:473-88. DOI PubMed
  71. Bauerschlag DO, Maass N, Leonhardt P, et al. Fatty acid synthase overexpression: target for therapy and reversal of chemoresistance in ovarian cancer. *J Transl Med* 2015;13:146. DOI PubMed PMC
  72. Wu X, Dong Z, Wang CJ, et al. FASN regulates cellular response to genotoxic treatments by increasing PARP-1 expression and DNA repair activity via NF-κB and SP1. *Proc Natl Acad Sci U S A* 2016;113:E6965-73. DOI PubMed PMC
  73. Kant S, Kumar A, Singh SM. Tumor growth retardation and chemosensitizing action of fatty acid synthase inhibitor orlistat on T cell lymphoma: implication of reconstituted tumor microenvironment and multidrug resistance phenotype. *Biochim Biophys Acta* 2014;1840:294-302. DOI PubMed
  74. Papaevangelou E, Almeida GS, Box C, deSouza NM, Chung YL. The effect of FASN inhibition on the growth and metabolism of a cisplatin-resistant ovarian carcinoma model. *Int J Cancer* 2018;143:992-1002. DOI PubMed PMC
  75. Hoy AJ, Nagarajan SR, Butler LM. Tumour fatty acid metabolism in the context of therapy resistance and obesity. *Nat Rev Cancer* 2021;21:753-66. DOI PubMed

76. Caragher S, Miska J, Shireman J, et al. Temozolomide treatment increases fatty acid uptake in glioblastoma stem cells. *Cancers (Basel)* 2020;12:3126. DOI PubMed PMC
77. Wang T, Fahrman JF, Lee H, et al. JAK/STAT3-regulated fatty acid  $\beta$ -oxidation is critical for breast cancer stem cell self-renewal and chemoresistance. *Cell Metab* 2018;27:136-50.e5. DOI PubMed PMC
78. Luo J, Hong Y, Tao X, Wei X, Zhang L, Li Q. An indispensable role of CPT-1a to survive cancer cells during energy stress through rewiring cancer metabolism. *Tumour Biol* 2016. DOI PubMed
79. Dubey R, Stivala CE, Nguyen HQ, et al. Lipid droplets can promote drug accumulation and activation. *Nat Chem Biol* 2020;16:206-13. DOI PubMed PMC
80. Sirois I, Aguilar-Mahecha A, Lafleur J, et al. A unique morphological phenotype in chemoresistant triple-negative breast cancer reveals metabolic reprogramming and PLIN4 expression as a molecular vulnerability. *Mol Cancer Res* 2019;17:2492-507. DOI PubMed
81. Cotte AK, Aires V, Fredon M, et al. Lysophosphatidylcholine acyltransferase 2-mediated lipid droplet production supports colorectal cancer chemoresistance. *Nat Commun* 2018;9:322. DOI PubMed PMC
82. Schito L, Semenza GL. Hypoxia-inducible factors: master regulators of cancer progression. *Trends Cancer* 2016;2:758-70. DOI PubMed
83. Semenza GL, Wang GL. A nuclear factor induced by hypoxia via de novo protein synthesis binds to the human erythropoietin gene enhancer at a site required for transcriptional activation. *Mol Cell Biol* 1992;12:5447-54. DOI PubMed PMC
84. Bhandari V, Li CH, Bristow RG, Boutros PC; PCAWG Consortium. Divergent mutational processes distinguish hypoxic and normoxic tumours. *Nat Commun* 2020;11:737. DOI PubMed PMC
85. Jin MZ, Jin WL. The updated landscape of tumor microenvironment and drug repurposing. *Signal Transduct Target Ther* 2020;5:166. DOI PubMed PMC
86. Dico A, Martelli C, Diceglie C, Lucignani G, Ottobrini L. Hypoxia-inducible factor-1 $\alpha$  activity as a switch for glioblastoma responsiveness to temozolomide. *Front Oncol* 2018;8:249. DOI PubMed PMC
87. Qiu GZ, Jin MZ, Dai JX, Sun W, Feng JH, Jin WL. Reprogramming of the tumor in the hypoxic niche: the emerging concept and associated therapeutic strategies. *Trends Pharmacol Sci* 2017;38:669-86. DOI PubMed
88. Becker HM. Carbonic anhydrase IX and acid transport in cancer. *Br J Cancer* 2020;122:157-67. DOI PubMed PMC
89. Webb BA, Chimenti M, Jacobson MP, Barber DL. Dysregulated pH: a perfect storm for cancer progression. *Nat Rev Cancer* 2011;11:671-7. DOI PubMed
90. Schöning JP, Monteiro M, Gu W. Drug resistance and cancer stem cells: the shared but distinct roles of hypoxia-inducible factors HIF1 $\alpha$  and HIF2 $\alpha$ . *Clin Exp Pharmacol Physiol* 2017;44:153-61. DOI PubMed
91. Lacroix R, Rozeman EA, Kreutz M, Renner K, Blank CU. Targeting tumor-associated acidity in cancer immunotherapy. *Cancer Immunol Immunother* 2018;67:1331-48. DOI PubMed
92. Bosticardo M, Ariotti S, Losana G, Bernabei P, Forni G, Novelli F. Biased activation of human T lymphocytes due to low extracellular pH is antagonized by B7/CD28 costimulation. *Eur J Immunol* 2001;31:2829-38. DOI PubMed
93. Sukumar M, Liu J, Ji Y, et al. Inhibiting glycolytic metabolism enhances CD8<sup>+</sup> T cell memory and antitumor function. *J Clin Invest* 2013;123:4479-88. DOI PubMed PMC
94. Pilon-Thomas S, Kodumudi KN, El-Kenawi AE, et al. Neutralization of tumor acidity improves antitumor responses to immunotherapy. *Cancer Res* 2016;76:1381-90. DOI PubMed PMC
95. Reshkin SJ, Cardone RA, Harguindey S. Na<sup>+</sup>-H<sup>+</sup> exchanger, pH regulation and cancer. *Recent Pat Anticancer Drug Discov* 2013;8:85-99. DOI PubMed
96. Orłowski J, Grinstein S. Emerging roles of alkali cation/proton exchangers in organellar homeostasis. *Curr Opin Cell Biol* 2007;19:483-92. DOI PubMed PMC
97. Tamtaji OR, Mirzaei H, Shamsirian A, Shamsirian D, Behnam M, Asemi Z. New trends in glioma cancer therapy: targeting Na<sup>+</sup>/H<sup>+</sup> exchangers. *J Cell Physiol* 2020;235:658-65. DOI PubMed
98. Chen Q, Liu Y, Zhu XL, Feng F, Yang H, Xu W. Increased NHE1 expression is targeted by specific inhibitor cariporide to sensitize resistant breast cancer cells to doxorubicin in vitro and in vivo. *BMC Cancer* 2019;19:211. DOI PubMed PMC
99. Aredia F, Czaplinski S, Fulda S, Scovassi AI. Molecular features of the cytotoxicity of an NHE inhibitor: evidence of mitochondrial alterations, ROS overproduction and DNA damage. *BMC Cancer* 2016;16:851. DOI PubMed PMC