

Review

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Epigenetic dysregulation in cardiovascular aging and disease

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Abstract

Cardiovascular disease (CVD) is the leading cause of mortality and morbidity for all sexes, racial and ethnic groups. Age, and its associated physiological and pathological consequences, exacerbate CVD incidence and progression, while modulation of biological age with interventions track with cardiovascular health. Despite the strong link between aging and CVD, surprisingly few studies have directly investigated heart failure and vascular dysfunction in aged models and subjects. Nevertheless, strong correlations have been found between heart disease, atherosclerosis, hypertension, fibrosis, and regeneration efficiency with senescent cell burden and its proinflammatory sequelae. In agreement, senotherapeutics have had success in reducing the detrimental effects in experimental models of cardiovascular aging and disease. Aside from senotherapeutics, cellular reprogramming strategies targeting epigenetic enzymes remain an unexplored yet viable option for reversing or delaying CVD. Epigenetic alterations comprising local and global changes in DNA and histone modifications, transcription factor binding, disorganization of the nuclear lamina, and misfolding of the genome are hallmarks of aging. Limited studies in the aging cardiovascular system of murine models or human patient samples have identified strong correlations between the epigenome, age, and senescence. Here, we compile the findings in published studies linking epigenetic changes to CVD and identify clear themes of epigenetic deregulation during aging. Pending direct investigation of these general mechanisms in aged tissues, this review predicts that future work will establish epigenetic rejuvenation as a potent method to delay CVD.



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INTRODUCTION

Cardiovascular disease (CVD), including heart failure, hypertension, atherosclerosis, and cardiomyopathy, remains the leading cause of death worldwide and carries a severe socioeconomic burden. While many factors contribute to CVD development, including diet, genetics, and the environment, one core, independent risk factor amongst almost everyone with a CVD is aging. It was once estimated that by 2030, about 20% of the United States population would be aged 65 or older and that CVD will account for 40% of the deaths of that group, making CVD the leading cause of death^[1,2]. According to the United States Census Bureau, we have observed an increase in the elderly population from 40 million in 2007 to 51 million in 2017, with the projected number of people over 65 to leap to 95 million in 2060. CVD prevalence continues to increase as human life expectancy also continues to rise, likely due to greater exposure to the traditional external risk factors and intrinsic pathways of aging^[3]. After adjusting for the other major risk factors for CVD, one study found the odds of vascular diseases increased with every decade of life, demonstrating a strong increase in peripheral arterial disease (PAD), carotid artery stenosis, and abdominal aortic aneurysm (AAA) with advanced age^[4].

The pathological consequences associated with normal cardiovascular aging include hypertrophy, altered left ventricular (LV) diastolic and systolic function, heart failure, enhanced arterial stiffness, and endothelial dysfunction, all of which can alter the structure and function of the heart and arterial system^[5,6]. In the vasculature specifically, aging contributes to decreasing vascular compliance. Furthermore, it promotes vascular remodeling, including calcification and fibrosis, which in turn precedes the development of hypertension and accelerates the progression of other vascular-related diseases such as atherosclerosis or heart failure^[7]. In addition, the incidence of metabolic diseases such as diabetes also increases significantly with age and contributes greatly to CVD morbidities and mortalities^[8]. Interestingly, many metabolic disorders are associated with premature aging, suggesting that there are mechanisms we can unravel to potentially intervene and prevent the deterioration of the cardiovascular system independent of natural aging.

Until recently, aging has widely been considered an unmodifiable risk factor for many chronic diseases (cancer and neurodegenerative diseases) and very prominently CVDs^[9-11]. Aging interventions have become a rising area of interest, where molecular and clinical dissection of aging processes have begun to show promising therapeutic targets. The monumental finding in 1939 that caloric restriction (CR) in mice and rats, and most recently in primates, extended lifespan led to the important hypothesis that lifespan extension with delayed aging improved healthspan^[12,13]. Since then, an examination of healthy aging and processes that promote age-related deterioration across species and organs has increased our understanding of the involvement of aging in chronic diseases^[14].

THE ROLE OF SENESCENT CELLS IN CARDIOVASCULAR PATHOLOGIES AND AGE-RELATED PATHWAYS

CVD and cellular senescence

A major contributor to age-related cellular dysfunction was found to be the accumulation of senescent cells in tissues^[15]. Senescent cells were discovered in 1965 by Hayflick^[16] as cells with a limited proliferative capacity; however, we now define senescence as those cells with indefinite cell cycle arrest, resistance to apoptosis, and expression of a senescence-associated secretory phenotype (SASP)^[17]. Thus, senescent cells provide a new avenue for therapeutic interventions, known as senotherapies^[18,19]. Specifically, a class of

drugs known as “senolytics” is designed to take advantage of the senescent cell’s resistance to apoptosis by targeting cell survival pathways to eliminate senescent cells from tissue selectively, thereby removing their detrimental effects^[20-22]. Alternatively, another class of drugs known as “senostatics” is designed to modulate the proinflammatory SASP; however, the complex composition of the SASP varies widely among different cell types, different stages of senescence (early, middle, or late), and various senescence inducers, providing many obstacles to a successful therapeutic intervention^[23]. While the components of the SASP may vary, the beneficial effects of senotherapies (both senolytics and senostatics) are mostly attributed to blunting the secretion of proinflammatory cytokines, chemokines, growth factors, and extracellular matrix (ECM) remodeling proteins, among others secreted by senescent cells^[24-26]. For a more detailed discussion of these senotherapeutic agents, please consult the reviews by Kirkland and Tchkonina^[24], 2020 and Robbins *et al.*^[26], 2021.

There is a rapidly growing body of evidence supporting the deleterious role of senescent cells in several CVDs. During embryonic development, tissue regeneration, and wound healing, vascular senescent cells have a beneficial presence to maintain homeostasis^[27]; however, we have learned that impaired removal and accumulation of senescent cells in cardiovascular tissue fomented impaired function and disease development. Senescent cells have been implicated in several CVD pathologies, most notably, atherosclerosis^[28], AAA^[29], cardiac fibrosis^[30], heart failure^[31], and hypertension^[32]. Further incriminating senescent cells as causative agents of CVD, Childs *et al.*^[33] demonstrated that senescent cells are critical drivers of atherosclerosis and selective removal of these cells has therapeutic potential to improve disease outcomes. In the same year, Roos *et al.*^[34] found that pharmacological clearance of senescent cells can lessen the vasomotor dysfunction that occurs in murine aging and atherosclerosis.

Senescent cardiomyocytes contribute to cardiac pathologies

Heart failure is an age-related cardiac pathology that is a major source of mortality, affecting approximately 1% of all people over 50 years and doubling its prevalence with each decade of life^[35,36]. Cardiomyocyte senescence is common in cardiac aging and related diseases, although senescent cardiomyocytes are more difficult to identify due to their terminally differentiated state^[37]. Senescent cardiomyocytes display contractile dysfunction, endoplasmic reticulum (ER) stress, DNA damage, genomic instability, declining mitochondrial function, SASP, and hypertrophic growth^[38]. Further, the exact triggers and effects of cardiomyocyte senescence *in vivo* have not been well described. However, studies in mice and rats have identified many of the signatures of cellular senescence, such as increased cardiomyocyte size, telomere attrition, ROS production, and elevated senescence markers p16 (CDKN2A) and p53 (TP53)^[39,40]. In hypertrophic cardiomyopathy patients, cardiomyocytes with DNA damage also had the shortest telomeres, and patients with ischemic cardiomyopathy also displayed shortened telomere length^[41]. While senescence is often associated with telomere shortening, cardiomyocytes are post-mitotic cells that do not experience replicative exhaustion; therefore, senescent cardiomyocytes demonstrate length-independent telomere damage caused by mitochondrial dysfunction and ROS^[42]. As mentioned above, hypertrophy is a hallmark of age-associated heart dysfunction, and although cardiomyocyte hypertrophic growth is commonly associated with senescence, it is unclear whether senescent myocyte growth directly contributes to cardiac hypertrophy^[43]. A few studies have found that ER stress appears to promote a hypertrophic cardiomyocyte phenotype *in vitro*, hypertrophy was detected in hearts post-infarction, and aged rat hearts demonstrated cardiomyocyte hypertrophy and increased LV fibrosis; however, none of these studies directly measured senescence^[43-45]. Interestingly, treatment of aged mice with the senolytic drug navitoclax selectively removed senescent cardiomyocytes, which improved myocardial remodeling and increased survival following myocardial infarction^[46]. While studies have outlined that accumulated damage to mitochondria, proteins, and DNA with age contributes to cardiomyocyte malfunction, telomere damage and cellular senescence are also critical to heart failure in humans, and more efforts will be needed to fully elucidate the contribution of

senescent cardiomyocytes to age-related cardiac pathologies^[47].

Further, cardiomyocyte senescence and the downstream pathologies are also the results of stress-induced premature senescence. Cardiomyocytes treated with doxorubicin demonstrated similar characteristics to those of aged rats, including increased senescence-associated beta-galactosidase positive cells, reduced telomerase activity, and increased expression of cell cycle regulatory proteins such as p16 and p21 (CDKN1A)^[48]. Recently, Mitry *et al.*^[49], further described the mechanism by which doxorubicin accelerates cardiomyocyte senescence and cardiotoxicity. In the study, Mitry *et al.*^[49] found that doxorubicin caused early and persistent topoisomerase-induced mtDNA damage that enhanced cardiomyocyte senescence, in turn straining the heart's aerobic metabolism over time and promoting late-onset heart failure often observed in survivors of childhood cancers.

Aside from cardiomyocytes, other cardiac cells promote senescence and aging and the downstream age-related diseases. For example, cardiac fibroblasts secrete many paracrine factors such as matrix metalloproteinases and express integrins to promote signaling and ECM interactions that regulate cardiomyocyte senescence^[50,51]. Endothelial cell senescence has also been implicated in heart failure with preserved ejection fraction, in which the activation of p53 signaling generates cardiac inflammation and left ventricular pressure overload in mice^[52]. Interestingly, cardiomyocyte dysfunction can also promote changes in neighboring cell types, such as fibroblasts, and impair the reparative function of cardiac fibrosis after cardiomyocyte injury^[53,54].

An important area of study for cardiac aging is impaired cardiomyocyte regeneration. We have discussed the key hallmark of cardiac aging; the increased size of cardiomyocytes, but another critical change is the loss of cardiomyocytes with age^[55]. While the neonatal heart demonstrates regenerative capacity, it was long thought the adult heart lacked the ability to renew cardiomyocytes^[56]. Recent observations that adult cardiomyocytes renew at a rate of 0.5% to 2% per year demonstrating a limited, innate regenerative ability that has dismantled those previous theories; however, the capacity of the heart to regenerate declines with age^[57-59]. Increased cardiomyocyte death, even in the very small numbers, was shown experimentally to promote heart failure, and that inhibiting the loss of cardiomyocytes, potentially through regeneration, could be an ideal therapeutic avenue^[60]. The heart regeneration field suffers from a lack of consistent and reproducible data on the subject; however, a consensus has developed that stem cells are not the source of cardiomyogenesis, but rather preexisting cardiomyocytes divide to give rise to new cells^[57,61-63]. A deeper understanding of the mechanisms that drive cardiomyocyte death with age may yield therapeutic potential for promoting regeneration in aged and damaged hearts.

Senescent vascular cells contribute to vascular diseases of aging

Among the many changes observed with aging, arterial remodeling and dysfunction are critical to the development of CVD, even in individuals who may be deemed healthy by all other standards. For example, aged arteries are defined by an increased ratio of intima-to-media thickness, and multiple reports have determined a 2- to 3-fold increase in intima thickness between 20- and 90-year-old people^[5]. In addition, changes in the arterial wall feature increased collagen synthesis and elastin degradation with age, promoting arterial stiffness and reduced elasticity^[64]. The consequence of such vascular remodeling manifests as increased blood pressure and lower diastolic pressure generating a predisposition to developing hypertension and atherosclerosis, among other vascular diseases^[65-67].

Both of the primary cell types of the artery, vascular smooth muscle cells (VSMCs) and endothelial cells (ECs), become senescent with age, regardless of the presence of a vascular-related disorder^[68-70]. The human

VSMCs from aged vessels and advanced-stage atherosclerotic plaques displayed senescence indicators with prolonged population doubling times and reduced cell proliferation^[71,72]. These findings were corroborated by associating the growth arrest of VSMCs with increased expression of p16 and p21, cyclin-dependent kinase inhibitors, and RB1 phosphorylation, all of which are observed during replicative VSMC senescence and are widely considered hallmarks of senescence^[28,73,74]. The VSMCs from the fibrous cap region of the atherosclerotic plaque compared to the vascular media demonstrated telomere shortening caused by oxidative stress-induced DNA damage. The resulting VSMC senescence accelerates vascular disorders such as atherosclerosis^[28]. Angiotensin II is another well-described driver of VSMC senescence, and recently, smooth muscle 22 α , an actin-binding protein, has been shown to prevent p53 degradation via MDM2 suppression to promote angiotensin II-induced VSMC senescence^[75]. Importantly, senescent VSMCs in the plaque of carotid arteries express enhanced levels of interleukin-6 (IL-6), signifying VSMCs as a SASP producer and source of inflammation during vascular disease^[76]. Most recently, Uryga *et al.*^[77] suggested persistent telomere damage in VSMCs causes senescence and inflammation via immune cell recruitment and retention. Overall, senescent VSMCs have been recognized in atherosclerotic lesions, AAA, and PAD, suggesting that VSMCs have a critical role in age-related vascular pathologies^[70].

Aside from VSMCs, ECs play an influential role in vascular disorders with age. While ECs typically maintain vascular homeostasis, senescent or dysfunctional ECs establish proinflammatory, prothrombotic, and vasoconstrictor characteristics in addition to reduced proliferation and migration. Replicative senescent ECs express increased cell adhesion molecules such as ICAM-1 and decreased endothelial nitric oxide synthase and activity, caused by telomere shortening^[70]. Another cause of EC senescence may be disturbed flow during atherosclerosis. In both mice and *in vitro*, the aberrant flow was a driver of EC senescence by activating the p21-p53 pathway^[78]. Aged and senescent ECs are also producers of inflammatory cytokines, namely IL-6, tumor necrosis factor alpha (TNF α), and monocyte chemoattractant protein-1 (MCP-1), which also suggests that the accumulation of senescent ECs in the artery with age causes chronic sterile inflammation and vascular changes that predispose one to vascular diseases^[79,80].

Although VSMCs and ECs compose most of the artery, immune cell aging and senescence may also contribute greatly to vascular pathologies associated with aging. Individuals 60 years or older with shortened telomeres in leukocytes experience a higher mortality rate that has been linked to increased death from CVD^[81]. In an interesting and clinically relevant study, the analysis of leukocyte populations led to the finding that telomere length was strongly associated with the development of atherosclerosis and CVD^[82,83]. Furthermore, senescent leukocytes and senescent effector memory T cells were found preferentially in unstable atherosclerotic plaques^[84]. Additionally, enhanced cytokine expression (TNF, MCP-1/CCL2, IL6) and ROS production have been observed in monocytes from atherosclerosis patients^[85]. Importantly, the proinflammatory phenotype of aged and senescent monocytes is driven by senescence^[86]. **Figure 1** summarizes the known consequences of cardiovascular aging and the molecular mechanisms, cell types, environmental factors involved, and potential therapeutics and interventions.

Overall, the evidence overwhelmingly points to the need to continue to study aging and senescence in CVD. Here, we present the body of work thus far that has uncovered numerous important pathways and mechanisms by which aged and senescent cells contribute to the development of different cardiovascular pathologies. Recurring thematic features of cardiovascular aging and disease suggest an unstable genome with shortened telomeres and a deregulated transcriptome that is pro-fibrotic, proinflammatory, but anti-proliferative with reduced regenerative capacity. In sum, these cellular phenotypes suggest an altered epigenome that has emerged as one of the hallmarks of aging in recent years. By focusing on mechanisms with druggable targets such as epigenetic alterations, we can develop therapies to modulate aging and

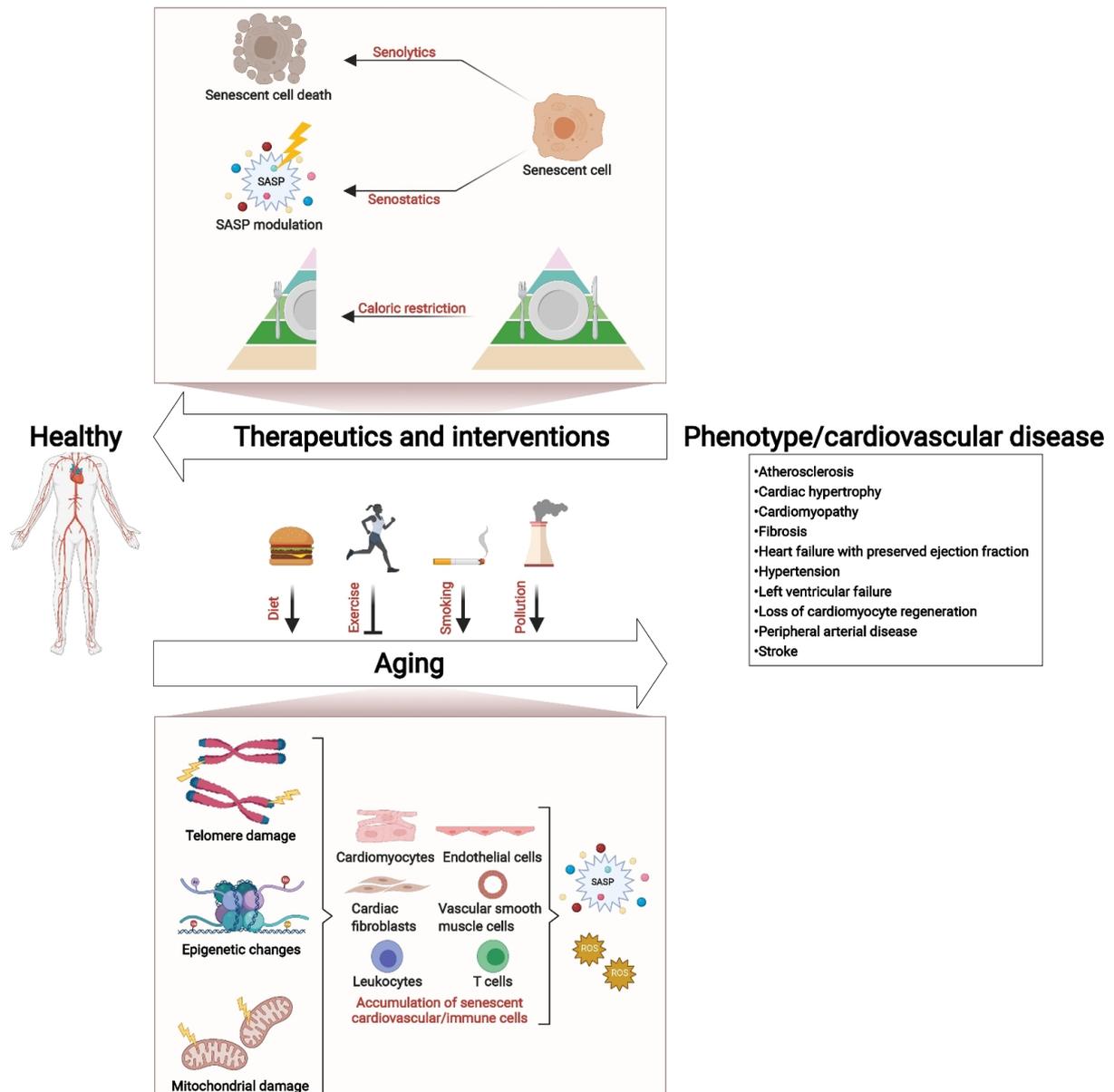


Figure 1. Molecular mechanisms involved in cardiovascular aging: consequences and potential therapeutics and interventions. Telomere damage, epigenetic changes, and mitochondrial damage are associated with the accumulation of senescent cardiovascular/immune cells, cardiovascular aging, and disease. Diet, smoking, and air pollution can also negatively contribute to aging, while physical exercise may improve cardiovascular health. Potential therapeutics and interventions include targeted elimination of senescent cells (senolytics), modulation of the proinflammatory SASP (senescence-associated secretory phenotype; senostatics), and dietary interventions (caloric restriction).

senescence in CVD. Outlined below are the central findings from studies that have investigated epigenetic changes in CVD, although as discussed in **FUTURE PERSPECTIVES**, these studies are limited and mostly out of context with aging. Nevertheless, these studies have revealed important insights that can be validated and developed into targeted therapeutics in the future.

EPIGENETIC CHANGES IN THE AGING CARDIOVASCULAR SYSTEM

Epigenetic alterations are one of the key features of aging and age-related disease, including CVD. These alterations include changes in DNA modifications, histone modifications, histone composition, transcription factor (TF) binding, non-coding RNA-mediated regulation, chromatin remodeling, nucleosome positioning, and 3D genome folding^[87,88]. Whether epigenetic changes drive aging or are a consequence of activated stress signaling pathways remains to be dissected but likely are part of a vicious cycle that ultimately leads to tissue damage, inflammation, and disease. In the following sections, we first discuss the probable impact of diet, exercise, and other environmental factors on cardiovascular health and then elaborate on the role of specific epigenetic regulators studied in the context of cardiovascular aging and disease.

Effect of diet, exercise, and other environmental factors in cardiovascular health

Aside from age, obesity and metabolic syndrome (characterized by sarcopenic obesity, insulin resistance, inflammation, *etc.*) are also major risk factors for CVD, partly due to their systemic proinflammatory effects, much like in aging^[89]. With increased body mass, there is an increase in the overall size of the heart, concentric hypertrophy, increased left ventricular mass, hypertension, and diastolic dysfunction, partially overlapping age-related cardiac symptoms. CR (i.e., a reduction in daily calorie intake without malnutrition) is one of the most reproducible lifestyle interventions that improve cardiovascular health and increase lifespan in multiple models^[90]. Studies in non-human primates show that rhesus monkeys on long-term, moderate CR show improvements in metabolic syndrome, including decreased body weight primarily due to loss of fat, decreased visceral fat mass, improved insulin sensitivity, and an altered lipid profile with more cardioprotective high-density lipoprotein compared to ad libitum fed controls^[13,91]. Interestingly, an inadvertent CR in humans participating in the Biosphere 2 experiment showed tremendous cardiovascular health benefits^[92].

Much like CR, exercise has demonstrated effects on cardiovascular health. Physical inactivity is a major contributing factor to age-related disabilities, declining heart health, stroke, cognitive impairment, and frailty primarily due to progressive arterial stiffness^[93]. Older individuals undergoing regular exercise show increased maximal oxygen consumption rate (VO_{2max})^[94]. Endurance exercise improves not only VO_{2max} but also early diastolic left ventricular filling and relaxation, peak ejection fraction, and cardiac output. There are also general improvements in vascular physiology and endothelial function^[95,96].

Unlike CR and exercise that have health benefits, smoking is a serious risk factor for cardiovascular disease. Smoking is often quantified in “pack-years”, i.e., the number of packs of cigarettes smoked per day multiplied by the number of years an individual has smoked ([cancer.gov](https://www.cancer.gov/)). In a study of > 13000 participants, Ding *et al.*^[97] determined a strong correlation between pack-years, duration, intensity, and cessation time of smoking to deleterious cardiovascular outcomes, with the strongest risk being PAD. In addition, smoking has been shown to directly target the epigenome, altering DNA methylation profiles, specifically 187 CpG sites independently validated in a separate cohort^[98]. In fact, some mortality predictive DNA methylation clocks, such as GrimAge (discussed below), directly incorporate smoking-related changes through an estimate of pack-years of smoking^[99].

Similar to smoking, air pollution is a major contributing factor that accelerates the decline of cardiopulmonary health. A growing body of epidemiological and clinical evidence indicates that ambient particulate matter may directly impact the cardiovascular system, although exact biological mechanisms are unknown^[100]. In part, the deleterious effects of particulate pollutants may be mediated by oxidative stress and systemic inflammation^[101]. Therefore, long-term studies focused on elucidating the molecular pathways

involved will be critical for designing mitigative approaches.

Given that environmental factors can impact multiple aspects of cardiovascular health and modulate lifespan, we discuss below some of the key molecular mechanisms that might be involved in this process. The epigenome is the interface between the environment and phenotype and, consequently, plays an important role in regulating health and disease.

DNA modifications in cardiac pathology

Methylation of cytosine (5-methylcytosine or 5mC) is the best-studied and most abundant modification on DNA. The 5mC status of groups of CpGs is associated with disease onset and mortality and therefore serves as the basis for several pan-tissue clocks that have been designed to act as “biological age” estimators. For example, GrimAge^[99] and PhenoAge^[102], two epigenetic clocks trained on chronological age and blood-based biomarkers, are associated with time to the incidence of CVD events^[103]. Although a clear mechanistic basis for these clocks is still obscure, the primary genomic regions affected seem to be polycomb targets and those near developmental genes^[104]. In concordance, a DNA methylome profiling in purified cardiomyocytes of mice undergoing heart failure showed methylation patterns that resembled those in neonates^[105]. Another independent epigenome-wide association study examining relationships between DNA methylation and incident CVD discovered two CpG modules in human cohorts: one associated with developmental genes and the other with immune functions^[106]. In keeping with the developmental gene activation theme in diseased hearts, the landscape of 5-hydroxymethylcytosine (5hmC, an oxidative product of 5mC) in cardiomyocytes derived from developing and hypertrophic hearts resemble, in part, a neonate-like signature. It was shown that 5hmC, which is positively correlated with gene transcription, was reduced over mitochondrial genes and increased over enhancers and gene bodies of fetal genes such as *Myh7*, thereby reactivating them^[107].

Investigation of DNA methylation in healthy and atherosclerotic lesions from donor-matched aorta samples interestingly revealed focal hypermethylation in the diseased tissue over repeat and non-repeat regions of the genome and in both a CpG and non-CpG context^[108]. Furthermore, the differentially methylated regions were associated with endothelial and smooth muscle function. A related study in swine, investigating differential methylation in ECs from an athero-susceptible location (inner curvature of the aortic arch) and an athero-protected region (descending thoracic aorta), also identified many hypermethylated sites that were linked to genes related to transcriptional regulation, pattern-specification HOX loci, oxidative stress, and ER stress adaptive pathway^[109]. Furthermore, 5'UTR hypermethylation exhibited an inverse relationship with gene expression at the HOX loci primarily. These observations contrast with DNA methylation changes in aging^[110] or cancer^[111], where global hypomethylation over megabase-sized blocks of the genome is the primary feature despite aging being a risk factor for atherosclerosis.

The derepression of repeat elements with retrotransposon activation is a known molecular event in senescence and aging. Evidence in senescent cells and mouse tissues indicates that these non-coding transcripts, generated from repeat elements, in turn, are reverse transcribed and activate an interferon response contributing to a systemic proinflammatory status in aging^[112]. The overall coverage of 5hmC over repeat elements was shown to decrease during cardiac development but increase in the hypertrophied heart, particularly at long interspersed nuclear elements. This was accompanied by reduced CG methylation and other repressive histone modifications (discussed below), suggesting a consequential activation of these regions in disease^[107].

Most genome-wide methylation studies have been done using bead-based arrays, whole-genome bisulfite sequencing, or reduced representation bisulfite sequencing post-bisulfite treatment of DNA. However, these methods fail to distinguish between 5mC or 5hmC and thereby may complicate mechanistic inferences on gene regulation. The recent development of the oxidative bisulfite sequencing (oxBS-seq) method allows for the simultaneous measurement of 5mC and 5hmC at single-nucleotide resolution^[113]. We propose that investigation of these distinct DNA modifications in cardiac aging and disease is an understudied but important future research direction.

Altered balance of activating and repressive histone modifications

Histone post-translational modifications (PTMs) represent another epigenetic mechanism to control gene expression. A core octamer comprising two copies each of H2A, H2B, H3, and H4 histones wraps 147 bp of DNA to form the basic unit of chromatin, the nucleosome. Linker histone H1 binds to linker DNA at the entry and exit sites of DNA on nucleosomes to form the next level of recurring chromatin structural unit, the chromatosome^[114]. Core and linker histones are modified by diverse PTMs such as acetylation, methylation, ubiquitylation, phosphorylation, *etc.* primarily on the unstructured tail regions (although there are many core modifications) and regulate activation or repression of gene expression via opening and closing of chromatin structure in a heritable fashion^[115]. [Table 1](#) provides an overview of known functions of specific histone modifications from the literature. Active or open chromatin is referred to as euchromatin, and inactive, closed chromatin is called heterochromatin. An existing notion in senescence studies points towards the progressive euchromatinization of the genome with concomitant loss of repressive modifications.

A genome-wide investigation of 7 histone PTMs, lysine 9 acetylation on histone H3 (H3K9ac), H3K27ac, H3K79me2, H3K4me3, H3K9me2, H3K9me3, and H3K27me3 (“me” indicating methylation) in cardiomyocytes isolated from normal and pressure-overloaded hearts revealed a subset of hypertrophy-associated genes that follow the conventional histone code, i.e., a mutually exclusive enrichment of activating (H3K9ac, H3K27ac, H3K79me2, and H3K4me3) and repressive (H3K9me2, H3K9me3, and H3K27me3) modifications. Additionally, this study identified a network of ~9000 putative active enhancers in the hypertrophic heart that might correlate to disease pathology, suggesting that histone PTMs regulate the gene network involved in this process^[116].

A cross-tissue analysis of chromatin marks (H3K4me3 and H3K27ac) revealed a clear separation in the RNA and chromatin profiles of young, middle-aged, and old hearts. Importantly, these age-related chromatin features included H3K4me3 and H3K27ac intensity and H3K4me3 breadth, which was previously shown to be linked to transcriptional consistency and high expression output required for maintenance of cell identity. Both dynamic features (such as enhancer score and H3K4me3 breadth) and static features (such as H3K4me3 promoter intensity and H3K4me3 domain breadth in young tissue) were key predictors of age^[117].

Studies focusing on the repressive H3K9 methylation, specifically H3K9me2, revealed that it promotes the reexpression of fetal genes during pathological cardiac hypertrophy. Downregulation of the H3K9 dimethyltransferases EHMT1/2 by miR-217 leads to loss of H3K9me2 over the promoters of fetal heart genes such as atrial natriuretic peptide (*Nppa*), brain natriuretic peptide (*Nppb*), and *Myh7* in cardiomyocytes^[118]. Knockout or overexpression of the H3K9 trimethyl demethylase JMJD2A (or KDM4A) had no overt cardiac phenotype but exhibited an altered response to stress. For example, overexpression of JMJD2A resulted in exacerbated cardiac hypertrophy while its inactivation was protective after aortic constriction^[119]. These results suggest that histone H3K9 repressive modifications play a critical role in

Table 1. Histone modifications involved in cardiovascular aging and disease

	Function	Enzymes(s)	Involvement in cardiovascular aging/disease	Model	Ref.
H3K9ac	Active promoter	KAT2A/2B, KDM4A-D, KDM3A-B	Positively associated with transcription in hypertrophic cardiomyocytes	<i>Homo sapiens, Mus musculus</i>	[116,174]
H3K27ac	Active promoter and enhancer	KAT3A-B, various HDACs	Increased at promoters of age-related genes	<i>Mus musculus</i>	[127,175]
H3K79me2	Body of active genes	KMT4	Associated with cardiac hypertrophy	<i>Mus musculus</i>	[116,176]
H3K4me3	Active promoters	KMT2F, KMT2G, KMT2A, KMT2D, KMT8B, KDM5A-D	Domain breadth increases in the aging heart	<i>Homo sapiens, Mus musculus, Drosophila melanogaster, Caenorhabditis elegans, Arabidopsis thaliana, and Saccharomyces cerevisiae</i>	[115,117]
H3K9me2	Nuclear lamina-associated heterochromatin	KMT1C, KMT1D, KDM3A-B, KDM4A-E, KDM7B	Reduced in pathological cardiac hypertrophy	<i>Mus musculus</i>	[118,177]
H3K9me3	Constitutive heterochromatin, repeat elements	KMT1A-B, KDM4A-E	JMJD2A-mediated demethylation is associated with cardiac hypertrophy and heart failure. Reduced in HGPS patients	<i>Homo sapiens, Mus musculus</i>	[119,156, 171,178]
H3K27me3	Facultative heterochromatin	KMT6A-B, KDM6A-B	Reduced in HGPS patients	<i>Homo sapiens</i>	[157,179]
H4K20me3	Heterochromatin	KMT5B-C, KDM7C	Increased in aging mice fibroblasts	<i>Homo sapiens</i>	[171,180]

suppressing a cardiac stress response that may also be occurring during aging, although it remains to be explicitly tested.

Histone acetylation is a very dynamic histone PTM and is regulated by histone acetyltransferases and histone deacetylases (HDACs). Loss of HDAC1, 2, 3, 5, and 9 results in exacerbated cardiac hypertrophy and, in some cases, neonatal lethality or a shortened lifespan^[120-122]. Sirtuins (SIRT1-7) are a family of nicotinamide adenine dinucleotide (NAD⁺)-dependent class III HDACs that have established protective roles in lifespan regulation in multiple species. However, both SIRT1 and NAD⁺ levels decline during aging^[123]. Furthermore, loss of SIRT1 interferes with angiogenesis and neovascularization after ischemia due to aberrant acetylation of FOXO1, potentiating its anti-angiogenic function^[124]. Conversely, overexpression of SIRT1 has many beneficial effects on endothelial cell function, including increased migration^[124], decreased endothelial progenitor cell senescence^[125], and reduced vascular oxidative stress and inflammation via inhibition of NFκB and PARP^[126]. While SIRT1 also acts on histones, this aspect of regulation remains unexplored.

Available studies taken together confirm that histone modifications contribute to CVD, with the direction of changes similar to that observed during senescence. There is a pronounced shift in the balance characterized by reduced repressive marks, especially over repeat elements and increased active modifications. However, due to the paucity of direct work in aged tissue and the lack of integrative analysis, the exact mechanisms remain to be elucidated.

A core TF network in aging

A multi-omic (DNA methylome, transcriptome, and epigenome) profiling and integrative analyses of the aging murine heart, liver, and quadriceps muscle identified some common and unique aging footprints across tissues. As a note, this study primarily performed a gene-centric analysis, and therefore changes over features such as enhancers or repeat regions were not analyzed. In the heart, the DNA methylome was the primary epigenetic signature that changed around transcription start sites (TSSs) with approximately an equal number of hypo- and hyper-methylated CpGs. Although more subtle, H3K27ac enrichment increased in the ~5 Kb region around the TSSs, while the H3K27me3 signal decreased. A TF motif enrichment analysis around promoters of genes up- or down-regulated in the heart during aging revealed that TF motifs enriched in genes that increase expression are also enriched in genes that have an increase in H3K27ac and a decrease in H3K27me3. Interestingly, a few TFs common to all three tissues were enriched in upregulated genes and genes with increases in H3K27ac and decreases in H3K27me3 in the heart. These TFs belong to the zinc finger of the cerebellum (Zic) family of factors. Conversely, HMGA1 binds to genes that are downregulated with age. Importantly, the expression of these TFs in humans is altered during aging, and epidemiological studies suggest a link between the altered expression of some of these TFs and the mother's age^[127]. These results suggest that a common set of epigenetic “master” regulators may be responsible for driving some of the key transcriptomic changes in aging.

Increased transcriptional noise in the aging heart

Single-cell studies have emphasized the presence of heterogeneity and variability within tumors, complex tissues, and surprisingly even overtly pure cell populations. For example, an early study with purified cardiomyocytes isolated from fresh young and old mice hearts revealed increased gene expression variability in old cells^[128]. The authors of the study attributed this increased variability to the stochastic nature of the aging process contributed by DNA damage and accumulating somatic mutations. Indeed, mouse embryonic fibroblasts treated with hydrogen peroxide showed a similar increase in expression variability. A more recent comprehensive single-cell atlas (Tabula Muris Senis^[129]) of multiple mouse tissues, including heart and aorta, is available but begs for a deeper dive into the dataset to enable the discovery of age-related changes specific to the cardiovascular system.

Non-coding RNA in cardiovascular aging

The vast majority of the genome is not translated into proteins but rather serves either as cis-regulatory elements or mediates post-transcriptional gene regulation^[130]. These non-coding areas of the genome encode small non-coding RNAs (< 200 nucleotides) or long non-coding RNAs (> 200 nucleotides). Small non-coding RNAs mainly comprise micro-RNAs (miRNAs), piwi-interacting RNAs (piRNAs), transfer RNAs (tRNAs), small nuclear RNAs (snRNAs), small nucleolar RNA (snoRNAs), *etc.* Long non-coding RNAs can be either linear (lncRNAs) or circular (circRNAs). Non-coding RNAs have long been implicated in senescence and aging, with several studies conducted in the context of cardiovascular aging^[131,132].

miRNAs and circRNAs present reliable biomarkers of aging due to their stability in circulation and conservation across species. miR-21 is a particularly well-characterized miRNA targeting SPRY1, a potent inhibitor of the ERK-MAPK pathway. miR-21 increases in cardiofibroblasts of the failing heart, augmenting ERK-MAP kinase activity impacting interstitial fibrosis and cardiac hypertrophy^[133]. In a study profiling miRNAs in the heart of neonatal, 1 month, 6 months and 19 months old mice, miR-22, which targets osteoglycin, was found to be robustly upregulated. miR-22 overexpression induced senescence and promoted the migration of cardiac fibroblasts^[134]. miR-34a is induced in aging cardiomyocytes where it targets PNUMS, a cardioprotective protein that otherwise reduces age-associated cardiomyocyte cell death^[135]. Interestingly, miR-34a, through the targeting of a different protein, SIRT1 (discussed above), induces endothelial and VSMC senescence and proinflammatory SASP expression^[136,137]. SIRT1 is also

targeted by miR-217 in ECs, where it induces premature senescence and leads to an impairment in angiogenesis via modulation of FOXO1 and nitric oxide synthase acetylation^[138]. Transcriptomic analysis of aortic tissue in old mice revealed miR-29 upregulation and the concomitant downregulation of many ECM components that in turn sensitizes the aorta to aneurysm formation^[139]. In contrast, a number of other miRNAs (miR-18, miR-19, miR-17-3p, miR-92, reviewed in^[140]) are reduced in expression during aging, specifically elevating their targets to affect cardiovascular aging and disease.

CircRNAs impact transcription by acting as sponges of miRNA and RNA binding proteins (RBPs) or serving as scaffolds for assembly of larger complexes^[141]. Many circRNAs are altered in expression upon hypoxic injury or myocardial infarction (reviewed extensively in^[142]); a few relevant to aging are discussed here. circFoxo3 is generated from the *Foxo3* transcript and was shown to be overexpressed in the aged hearts of mice and humans and correlated to senescence markers. CircFoxo3 is localized to the cytoplasm where it retains several anti-senescence proteins such as ID1, E2F1, FAK, and HIF1 α , thus siphoning their activity away from the nucleus^[143]. In addition, a circRNA produced from the senescence/aging relevant *Cdkn2b* locus called Antisense non-coding RNA in the INK4 locus (circANRIL) correlates with the expression of its linear RNA and confers atheroprotection. circANRIL binds to PES1, a 60S-preribosomal assembly factor, impairs ribosome biogenesis and thereby induces nucleolar stress and apoptosis in atherogenic VSMCs and macrophages^[144,145]. Unlike miRNAs that are well studied in aged hearts and vasculature, key age-related circRNAs remain to be profiled in detail.

lncRNAs, unlike miRNAs, are not well conserved across species, and therefore their targets and functions should be interpreted with caution. Nevertheless, numerous studies have evaluated the role of lncRNAs in cardiovascular aging and disease (reviewed in^[146]). lncRNAs are highly versatile, serving as expression signals to trigger a response, competitive endogenous RNAs, guides to direct factors to specific genomic locations, scaffolds for RBPs, or mediators of chromatin looping^[146]. For example, *Mhrt*, an antisense lncRNA produced from the region between *Myh6* and *Myh7*, interferes with the switch to fetal *Myh7* expression in hypertrophic hearts. *Mhrt* antagonizes BRG1 function by interacting with its helicase domain and inhibiting chromatin targeting (see next section)^[147]. *Chaer*, another lncRNA mediating cardiac hypertrophy, interacts with PRC2 subunits and thereby inhibits the repression of cardiac hypertrophy-related genes^[148]. *Meg3* expression is upregulated in senescent HUVEC (endothelial) cells and in the aging cardiovascular system, where it also targets PRC2 components to assert a pro-aging function in aging vasculature^[149]. An RNA-seq study in porcine cardiac muscle revealed 4 lncRNAs that were consistently expressed during aging. Ontology analysis of the target genes of these lncRNAs was significantly enriched for negative regulation of myotube differentiation and muscle contraction, suggesting that the lncRNAs likely interfere with the normal muscle physiology^[150].

There are numerous other examples of non-coding RNA functions in cardiovascular aging that are beyond the scope of this review. However, it is interesting to note that many of them target epigenetic enzymes or TFs and therefore may directly and pervasively impact the epigenome.

ATP-dependent chromatin remodeling in the diseased heart

ATP-dependent chromatin remodeling complexes are large multi-subunit molecular machines that utilize ATP to reposition or evict nucleosomes or exchange histones to alter chromatin structure. The BRG1/BRM-associated factor (BAF) chromatin remodeling complexes are comprised of either brahma or brahma-related gene 1 (BRG1) catalytic subunits along with several other accessory proteins. BAF complexes are critical for heart development and disease pathogenesis. For example, BRG1 plays opposing roles at the *Myh6* and *Myh7* loci: in embryos, it interacts with HDACs and poly (ADP ribose) polymerase 1 (PARP1) to

repress the adult-specific *Myh6* while activating fetal *Myh7*^[151]. *Brg1* expression is lost in cardiomyocytes but reactivated in hypertrophic hearts. It sequentially recruits G9a and then DNMT3 to deposit H3K9me2 and 5mC at the *Myh6* promoter impairing cardiac contraction^[152]. Thus, a complex interplay of repressors and co-repressors recruited by BRG1 in injured hearts activates fetal *Myh7* while interfering with adult *Myh6* expression. Cardiac regeneration (as may be promoted by injury) requires BRG1 not only to suppress *Myh6* but also to increase the expression of pro-proliferative *Bmp10* and *Cdkn1c* genes. Although not directly tested in aging hearts, reactivation of *Brg1* is a plausible mechanism to promote repair. Finally, mutations in genes encoding BAF complex subunits have been associated with various cancers and congenital heart diseases^[153,154].

Laminopathy and loss of heterochromatin lead to premature aging

Hutchinson Gilford Progeria Syndrome (HGPS) is a premature aging disorder attributed to a mutation in the lamin A (*LMNA*) gene that results in the production and incorporation of a truncated version of lamin A called progerin in the nuclear membrane. This misincorporation grossly disrupts the nuclear lamina and lamina-associated heterochromatin and pro-senescence/pro-aging gene expression changes. Surprisingly, HGPS patients usually die in their teens from atherosclerosis and CVD complications suggesting strong links between chromatin dysregulation and CVD events in these patients. Interestingly, vascular progerin production and its progressive increase with age have also been noted in normal individuals, and there are many common histological features in the vasculature of HGPS and geriatric subjects^[155]. Additionally, fibroblasts isolated from HGPS patients undergo premature senescence, and in iPSC models of HGPS, many epigenetic changes noted mimic those found in *in vitro* models of cellular senescence. For example, there is a reduction in repressive histone modifications, H3K27me3 and H3K9me3, loss of EZH2, and derepression of LINE elements^[156,157]. This evidence suggests that epigenetic alterations, particularly those found in senescent cells, may drive some of the key CVD pathologies in HGPS. Indeed, selective clearance of naturally occurring p16 positive senescent cells in the heart and senescent foam macrophages at atherosclerotic lesions by senolytics can ameliorate disease symptoms^[33,158].

Figure 2 summarizes the key concepts of epigenetic regulation impacting CVD: DNA modifications [Figure 2A], histone modifications [Figure 2B], TF binding [Figure 2C], altered gene expression [Figure 2D], non-coding RNAs [Figure 2E], chromatin remodeling [Figure 2F] and lamina disorganization [Figure 2G] as derived from models of heart failure, and limitedly, aged tissues.

FUTURE PERSPECTIVES AND PROVOCATIVE THERAPIES FOR AGE-RELATED CVD

With the advent of better health monitoring, chemotherapies, vaccines, and rehabilitation programs, human life expectancy has increased and will continue to increase in the next few decades. This means that the number of people > 65 years of age will comprise 20% or more of the population by the next decade. Unfortunately, CVD will remain one of the top causes of death among older individuals, surpassing neurodegenerative diseases and cancers, suggesting that the cardiovascular system is especially prone to the chronic deleterious changes that come with age. Until recently, age was thought to be a largely unmodifiable feature of life, but the innovations of the longevity biotechnology field are on a trajectory to change this outcome. Thus, now is the time to identify key mechanisms contributing to heart disease in the elderly to design and rapid testing of breakthrough therapeutics.

Several notable interventions which directly or indirectly remodel the epigenome hold promise in ameliorating CVD. Preclinical studies in mouse models have already shown the efficacy of senolytics in countering the deleterious effects of cardiac dysfunction, vascular dysfunction, and calcification^[33,34,159,160]. Senostatics that reduce the SASP without eliminating senescent cells, which carry the risk of fibrosis, might

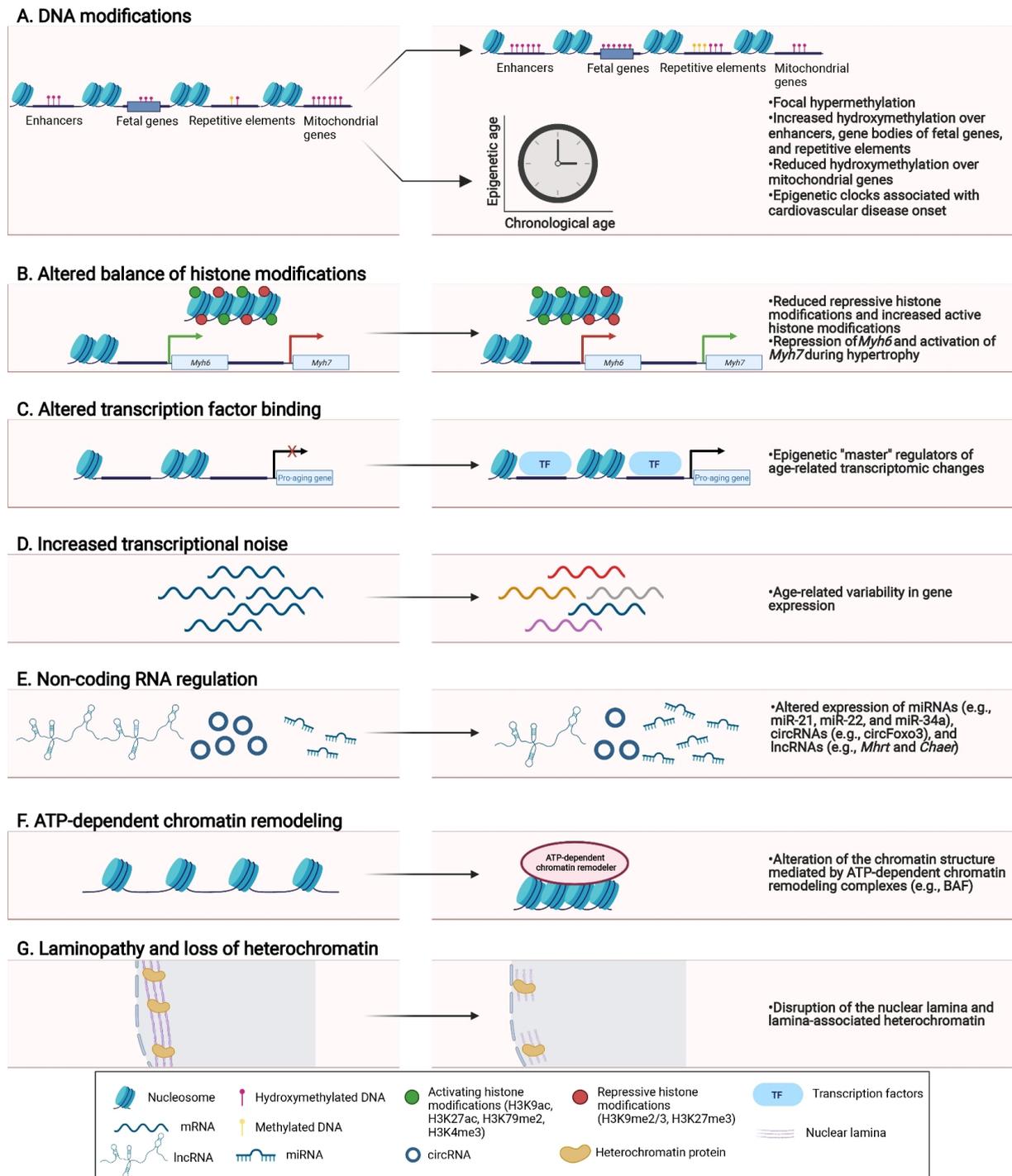


Figure 2. Epigenetic mechanisms involved in cardiovascular aging and disease. Several epigenetic changes are documented in cardiovascular aging and disease, including (A) DNA modifications (5-methylcytosine is also used in epigenetic clocks and associated with cardiovascular disease onset); (B) altered balance of active and repressive histone marks; (C) alterations to transcription factor binding; (D) transcriptional changes; (E) altered expression of non-coding RNAs; (F) chromatin remodeling; and (G) laminopathy and loss of heterochromatin.

also show benefits but have not been directly tested. Potential SASP modulators include glucocorticoids^[161], rapamycin^[162], metformin^[163] and CR/CR mimetics^[164]. Although the exact mechanisms underlying age

reversal are lacking, these molecules have a profound effect on the epigenome (reviewed in^[87,88,165]). Some direct effects of epigenome remodeling are exemplified by enzymes such as MLL1^[166] and BRD4^[167], which are critical regulators of *SASP* genes. Additional synthetic therapeutics that could potentially modulate senescence or *SASP* include locked nucleic acids, anti-miRs, and other antisense oligonucleotides that block non-coding RNA activity and/or target them for degradation^[168]. Overall, given the predominance of senescent cell function in CVD, targeting them is a viable option to treat age-related cardiac dysfunction.

Another targetable cell type in CVD is the quiescent cardiomyocyte and fibroblast populations that comprise most adult heart tissue. Unlike senescent cells, quiescent cells are responsive to growth factors and apoptotic signals, making them more pliable for modulation. While neonatal cardiomyocytes are capable of proliferation and regeneration, this function declines rapidly in adults^[169]. The cardiac stem cell theory was recently annulled following extensive fate-mapping data that clearly showed that non-myocytes could not produce new cardiomyocytes in the adult during homeostasis or following infarction^[170]. Thus, an endogenous stem cell-centric therapy in CVD is contentious. However, the exogenous supply of cardiac progenitors produced from induced pluripotent stem cells could be explored but need careful testing. Another avenue to improve regeneration of adult cardiomyocytes is by inducing controlled proliferation, for example, by cyclical expression of Yamanaka factors. In a premature aging model carrying a *Lmna* mutation, cyclic induction of these pluripotency factors partially rescued the degeneration of VSMCs in the aortic arch compared to untreated mice, as indicated by an increase in the nuclei number. At the functional level, electrocardiographic analysis showed that there was also a partial rescue of bradycardia in the treated mice compared to controls. Yamanaka factors induce reprogramming by changing the histone modification landscape, specifically restoring H3K9me3 and H4K20me3 to youthful levels^[171].

Pathological, activated cardiac fibroblasts (as opposed to quiescent fibroblasts) are induced following cardiac injury and can cause excessive fibrosis. These activated fibroblasts have been shown to have a unique gene expression signature, prominently the upregulation of fibroblast activation protein, which was targeted to eliminate them by chimeric antigen receptor (CAR)-T cell therapy^[172] selectively. We propose that a similar survey of the transcriptome and epigenome can discover neoantigens on aged tissues, which can then be exploited for immunotherapy in CVD.

Of note, most of the studies discussed in this review use mouse models of heart failure or cardiac disease to interrogate epigenetic features. However, the experimental rodent is usually an adult (2-4 months old) with a very different epigenomic landscape than older animals, who present the most risk for disease. These models thus may accurately capture acute pathological changes while completely missing the contribution to disease of any long-term chronic effects such as systemic inflammation or global epigenetic changes. Conversely, studies that focused on interventions that extend lifespan rarely measured whether the cardiovascular function was improved^[173]. Collectively, the field must embrace naturally aged mice models and impose the inclusion of age as a biological variable to gain deeper insight into the etiology of age-related cardiac dysfunction and disease.

CONCLUDING REMARKS

In the studies that have considered both the perspective of aging and cardiovascular health, we have accumulated important insights into the epigenetic mechanisms of cardiovascular aging that we describe in this review. Genomic regions that are targeted during aging include repeat elements and lamina-bound heterochromatin, developmental gene promoters, polycomb targets, and stress response genes. These regions also show prominent changes in DNA modifications. The histone code itself is unaltered, but specific master TFs co-opt epigenetic enzymes and chromatin architectural proteins to promote a disease

phenotype. Concordant with changes in DNA and histone modifications, the coding, and non-coding transcriptome is also significantly altered, impacting cardiac function and vascular physiology. Further studies will illuminate more precise roles of epigenetic factors that can ultimately be exploited to design novel therapeutics.

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Authors' contributions

Wrote the manuscript: Herman AB, Occean JR, Sen P
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REFERENCES

1. Fleg JL, Aronow WS, Frishman WH. Cardiovascular drug therapy in the elderly: benefits and challenges. *Nat Rev Cardiol* 2011;8:13-28. [DOI](#) [PubMed](#)
2. Heidenreich PA, Trogon JG, Khavjou OA, et al; American Heart Association Advocacy Coordinating Committee; Stroke Council; Council on Cardiovascular Radiology and Intervention; Council on Clinical Cardiology; Council on Epidemiology and Prevention; Council on Arteriosclerosis, Thrombosis and Vascular Biology; Council on Cardiopulmonary, Critical Care, Perioperative and Resuscitation; Council on Cardiovascular Nursing; Council on the Kidney in Cardiovascular Disease; Council on Cardiovascular Surgery and Anesthesia; Interdisciplinary Council on Quality of Care and Outcomes Research. Forecasting the future of cardiovascular disease in the United States: a policy statement from the American Heart Association. *Circulation* 2011;123:933-44. [DOI](#) [PubMed](#)
3. Foreman KJ, Marquez N, Dolgert A, et al. Forecasting life expectancy, years of life lost, and all-cause and cause-specific mortality for 250 causes of death: reference and alternative scenarios for 2016-40 for 195 countries and territories. *Lancet* 2018;392:2052-90. [DOI](#) [PubMed](#) [PMC](#)
4. Savji N, Rockman CB, Skolnick AH, et al. Association between advanced age and vascular disease in different arterial territories: a population database of over 3.6 million subjects. *J Am Coll Cardiol* 2013;61:1736-43. [DOI](#) [PubMed](#)
5. Lakatta EG, Levy D. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: Part I: aging arteries: a "set up" for vascular disease. *Circulation* 2003;107:139-46. [DOI](#) [PubMed](#)
6. Lakatta EG, Levy D. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: Part II: the aging heart in health: links to heart disease. *Circulation* 2003;107:346-54. [DOI](#) [PubMed](#)
7. Barton M, Husmann M, Meyer MR. Accelerated vascular aging as a paradigm for hypertensive vascular disease: prevention and

- therapy. *Can J Cardiol* 2016;32:680-6.e4. DOI PubMed
8. Fadini GP, Ceolotto G, Pagnin E, de Kreutzenberg S, Avogaro A. At the crossroads of longevity and metabolism: the metabolic syndrome and lifespan determinant pathways. *Aging Cell* 2011;10:10-7. DOI PubMed
 9. Niccoli T, Partridge L. Ageing as a risk factor for disease. *Curr Biol* 2012;22:R741-52. DOI PubMed
 10. Aviv A. Chronology versus biology: telomeres, essential hypertension, and vascular aging. *Hypertension* 2002;40:229-32. DOI PubMed
 11. Shakeri H, Lemmens K, Gevaert AB, De Meyer GRY, Segers VFM. Cellular senescence links aging and diabetes in cardiovascular disease. *Am J Physiol Heart Circ Physiol* 2018;315:H448-62. DOI PubMed
 12. Mccay CM, Maynard LA, Sperling G, Barnes LL. Retarded growth, life span, ultimate body size and age changes in the Albino rat after feeding diets restricted in calories. *J Nutr* 1939;18:1-13. DOI
 13. Mattison JA, Colman RJ, Beasley TM, et al. Caloric restriction improves health and survival of rhesus monkeys. *Nat Commun* 2017;8:14063. DOI PubMed PMC
 14. Campisi J, Kapahi P, Lithgow GJ, Melov S, Newman JC, Verdin E. From discoveries in ageing research to therapeutics for healthy ageing. *Nature* 2019;571:183-92. DOI PubMed PMC
 15. McHugh D, Gil J. Senescence and aging: causes, consequences, and therapeutic avenues. *J Cell Biol* 2018;217:65-77. DOI PubMed PMC
 16. Hayflick L. The limited in vitro lifetime of human diploid cell strains. *Exp Cell Res* 1965;37:614-36. DOI PubMed
 17. Coppé JP, Patil CK, Rodier F, et al. Senescence-associated secretory phenotypes reveal cell-nonautonomous functions of oncogenic RAS and the p53 tumor suppressor. *PLoS Biol* 2008;6:2853-68. DOI PubMed PMC
 18. Niedernhofer LJ, Robbins PD. Senotherapeutics for healthy ageing. *Nat Rev Drug Discov* 2018;17:377. DOI PubMed
 19. Childs BG, Gluscevic M, Baker DJ, et al. Senescent cells: an emerging target for diseases of ageing. *Nat Rev Drug Discov* 2017;16:718-35. DOI PubMed PMC
 20. Zhu Y, Tchkonina T, Pirtskhalava T, et al. The Achilles' heel of senescent cells: from transcriptome to senolytic drugs. *Aging Cell* 2015;14:644-58. DOI PubMed PMC
 21. Ellison-Hughes GM. First evidence that senolytics are effective at decreasing senescent cells in humans. *EBioMedicine* 2020;56:102473. DOI PubMed PMC
 22. Deursen JM. Senolytic therapies for healthy longevity. *Science* 2019;364:636-7. DOI PubMed PMC
 23. Kang C. Senolytics and Senostatics: A two-pronged approach to target cellular senescence for delaying aging and age-related diseases. *Mol Cells* 2019;42:821-7. DOI PubMed PMC
 24. Kirkland JL, Tchkonina T. Senolytic drugs: from discovery to translation. *J Intern Med* 2020;5:518-36. DOI PubMed PMC
 25. Lujambio A. To clear, or not to clear (senescent cells)? *Bioessays* 2016;38 Suppl 1:S56-64. DOI PubMed
 26. Robbins PD, Jurk D, Khosla S, et al. Senolytic drugs: reducing senescent cell viability to extend health span. *Annu Rev Pharmacol Toxicol* 2021;61:779-803. DOI PubMed PMC
 27. Demaria M, Ohtani N, Youssef SA, et al. An essential role for senescent cells in optimal wound healing through secretion of PDGF-AA. *Dev Cell* 2014;31:722-33. DOI PubMed PMC
 28. Matthews C, Gorenne I, Scott S, et al. Vascular smooth muscle cells undergo telomere-based senescence in human atherosclerosis: effects of telomerase and oxidative stress. *Circ Res* 2006;99:156-64. DOI PubMed
 29. Chen HZ, Wang F, Gao P, et al. Age-associated Sirtuin 1 reduction in vascular smooth muscle links vascular senescence and inflammation to abdominal aortic aneurysm. *Circ Res* 2016;119:1076-88. DOI PubMed PMC
 30. Sawaki D, Czibik G, Pini M, et al. Visceral adipose tissue drives cardiac aging through modulation of fibroblast senescence by osteopontin production. *Circulation* 2018;138:809-22. DOI PubMed
 31. Gevaert AB, Shakeri H, Leloup AJ, et al. Endothelial senescence contributes to heart failure with preserved ejection fraction in an aging mouse model. *Circ Heart Fail* 2017;10:e003806. DOI PubMed
 32. Boe AE, Eren M, Murphy SB, et al. Plasminogen activator inhibitor-1 antagonist TM5441 attenuates N^o-nitro-L-arginine methyl ester-induced hypertension and vascular senescence. *Circulation* 2013;128:2318-24. DOI PubMed PMC
 33. Childs BG, Baker DJ, Wijshake T, Conover CA, Campisi J, van Deursen JM. Senescent intimal foam cells are deleterious at all stages of atherosclerosis. *Science* 2016;354:472-7. DOI PubMed PMC
 34. Roos CM, Zhang B, Palmer AK, et al. Chronic senolytic treatment alleviates established vasomotor dysfunction in aged or atherosclerotic mice. *Aging Cell* 2016;15:973-7. DOI PubMed PMC
 35. Mosterd A, Hoes AW. Clinical epidemiology of heart failure. *Heart* 2007;93:1137-46. DOI PubMed PMC
 36. Benjamin EJ, Muntner P, Alonso A, et al; American Heart Association Council on Epidemiology and Prevention Statistics Committee and Stroke Statistics Subcommittee. Heart disease and stroke statistics-2019 update: a report from the American heart association. *Circulation* 2019;139:e56-e528. DOI PubMed
 37. Ock S, Lee WS, Ahn J, et al. Deletion of IGF-1 receptors in cardiomyocytes attenuates cardiac aging in male mice. *Endocrinology* 2016;157:336-45. DOI PubMed PMC
 38. Tang X, Li PH, Chen HZ. Cardiomyocyte senescence and cellular communications within myocardial microenvironments. *Front Endocrinol (Lausanne)* 2020;11:280. DOI PubMed PMC
 39. Torella D, Rota M, Nurzynska D, et al. Cardiac stem cell and myocyte aging, heart failure, and insulin-like growth factor-1 overexpression. *Circ Res* 2004;94:514-24. DOI PubMed
 40. Spallarossa P, Altieri P, Aloï C, et al. Doxorubicin induces senescence or apoptosis in rat neonatal cardiomyocytes by regulating the expression levels of the telomere binding factors 1 and 2. *Am J Physiol Heart Circ Physiol* 2009;297:H2169-81. DOI PubMed

41. Sharifi-Sanjani M, Oyster NM, Tichy ED, et al. Cardiomyocyte-specific telomere shortening is a distinct signature of heart failure in humans. *J Am Heart Assoc* 2017;6:e005086. DOI PubMed PMC
42. Anderson R, Lagnado A, Maggiorani D, et al. Length-independent telomere damage drives post-mitotic cardiomyocyte senescence. *EMBO J* 2019;38:e100492. DOI PubMed PMC
43. Cui S, Xue L, Yang F, et al. Postinfarction hearts are protected by premature senescent cardiomyocytes via GATA 4-dependent CCN1 secretion. *J Am Heart Assoc* 2018;7:e009111. DOI PubMed PMC
44. Xie F, Wu D, Huang SF, et al. The endoplasmic reticulum stress-autophagy pathway is involved in apelin-13-induced cardiomyocyte hypertrophy in vitro. *Acta Pharmacol Sin* 2017;38:1589-600. DOI PubMed PMC
45. Forman DE, Cittadini A, Azhar G, Douglas PS, Wei JY. Cardiac morphology and function in senescent rats: gender-related differences. *J Am Coll Cardiol* 1997;30:1872-7. DOI PubMed
46. Walaszczyk A, Dookun E, Redgrave R, et al. Pharmacological clearance of senescent cells improves survival and recovery in aged mice following acute myocardial infarction. *Aging Cell* 2019;18:e12945. DOI PubMed PMC
47. Chimenti C, Kajstura J, Torella D, et al. Senescence and death of primitive cells and myocytes lead to premature cardiac aging and heart failure. *Circ Res* 2003;93:604-13. DOI PubMed
48. Maejima Y, Adachi S, Ito H, Hirao K, Isobe M. Induction of premature senescence in cardiomyocytes by doxorubicin as a novel mechanism of myocardial damage. *Aging Cell* 2008;7:125-36. DOI PubMed
49. Mitry MA, Laurent D, Keith BL, et al. Accelerated cardiomyocyte senescence contributes to late-onset doxorubicin-induced cardiotoxicity. *Am J Physiol Cell Physiol* 2020;318:C380-91. DOI PubMed PMC
50. Civitarese RA, Kapus A, McCulloch CA, Connelly KA. Role of integrins in mediating cardiac fibroblast-cardiomyocyte cross talk: a dynamic relationship in cardiac biology and pathophysiology. *Basic Res Cardiol* 2017;112:6. DOI PubMed
51. Saucerman JJ, Tan PM, Buchholz KS, McCulloch AD, Omens JH. Mechanical regulation of gene expression in cardiac myocytes and fibroblasts. *Nat Rev Cardiol* 2019;16:361-78. DOI PubMed PMC
52. Yoshida Y, Shimizu I, Katsuomi G, et al. p53-Induced inflammation exacerbates cardiac dysfunction during pressure overload. *J Mol Cell Cardiol* 2015;85:183-98. DOI PubMed
53. Biernacka A, Frangogiannis NG. Aging and cardiac fibrosis. *Aging Dis* 2011;2:158-73. PubMed PMC
54. Russo I, Frangogiannis NG. Diabetes-associated cardiac fibrosis: Cellular effectors, molecular mechanisms and therapeutic opportunities. *J Mol Cell Cardiol* 2016;90:84-93. DOI PubMed PMC
55. Olivetti G, Melissari M, Capasso JM, Anversa P. Cardiomyopathy of the aging human heart. Myocyte loss and reactive cellular hypertrophy. *Circ Res* 1991;68:1560-8. DOI PubMed
56. Porrello ER, Mahmoud AI, Simpson E, et al. Transient regenerative potential of the neonatal mouse heart. *Science* 2011;331:1078-80. DOI PubMed PMC
57. Senyo SE, Steinhauser ML, Pizzimenti CL, et al. Mammalian heart renewal by pre-existing cardiomyocytes. *Nature* 2013;493:433-6. DOI PubMed PMC
58. Bergmann O, Bhardwaj RD, Bernard S, et al. Evidence for cardiomyocyte renewal in humans. *Science* 2009;324:98-102. DOI PubMed PMC
59. Bergmann O, Zdunek S, Felker A, et al. Dynamics of cell generation and turnover in the human heart. *Cell* 2015;161:1566-75. DOI PubMed
60. Wencker D, Chandra M, Nguyen K, et al. A mechanistic role for cardiac myocyte apoptosis in heart failure. *J Clin Invest* 2003;111:1497-504. DOI PubMed PMC
61. Eschenhagen T, Bolli R, Braun T, et al. Cardiomyocyte regeneration: a consensus statement. *Circulation* 2017;136:680-6. DOI PubMed PMC
62. Ali SR, Hippenmeyer S, Saadat LV, Luo L, Weissman IL, Ardehali R. Existing cardiomyocytes generate cardiomyocytes at a low rate after birth in mice. *Proc Natl Acad Sci USA* 2014;111:8850-5. DOI PubMed PMC
63. Hsieh PC, Segers VF, Davis ME, et al. Evidence from a genetic fate-mapping study that stem cells refresh adult mammalian cardiomyocytes after injury. *Nat Med* 2007;13:970-4. DOI PubMed PMC
64. Safar ME. Systolic hypertension in the elderly: arterial wall mechanical properties and the renin-angiotensin-aldosterone system. *J Hypertens* 2005;23:673-81. DOI PubMed
65. Strait JB, Lakatta EG. Aging-associated cardiovascular changes and their relationship to heart failure. *Heart Fail Clin* 2012;8:143-64. DOI PubMed PMC
66. Spina M, Garbisa S, Hinnie J, Hunter JC, Serafini-Fracassini A. Age-related changes in composition and mechanical properties of the tunica media of the upper thoracic human aorta. *Arteriosclerosis* 1983;3:64-76. DOI PubMed
67. Harvey A, Montezano AC, Touyz RM. Vascular biology of ageing-Implications in hypertension. *J Mol Cell Cardiol* 2015;83:112-21. DOI PubMed PMC
68. Morgan RG, Ives SJ, Lesniewski LA, et al. Age-related telomere uncapping is associated with cellular senescence and inflammation independent of telomere shortening in human arteries. *Am J Physiol Heart Circ Physiol* 2013;305:H251-8. DOI PubMed PMC
69. Marchand A, Atassi F, Gaaya A, et al. The Wnt/beta-catenin pathway is activated during advanced arterial aging in humans. *Aging Cell* 2011;10:220-32. DOI PubMed
70. Minamino T, Miyauchi H, Yoshida T, Ishida Y, Yoshida H, Komuro I. Endothelial cell senescence in human atherosclerosis: role of telomere in endothelial dysfunction. *Circulation* 2002;105:1541-4. DOI PubMed
71. O'Brien ER, Alpers CE, Stewart DK, et al. Proliferation in primary and restenotic coronary atherectomy tissue. Implications for

- antiproliferative therapy. *Circ Res* 1993;73:223-31. DOI PubMed
72. Bennett MR, Evan GI, Schwartz SM. Apoptosis of human vascular smooth muscle cells derived from normal vessels and coronary atherosclerotic plaques. *J Clin Invest* 1995;95:2266-74. DOI PubMed PMC
 73. O'Sullivan M, Scott SD, McCarthy N, et al. Differential cyclin E expression in human in-stent stenosis smooth muscle cells identifies targets for selective anti-restenosis therapy. *Cardiovasc Res* 2003;60:673-83. DOI PubMed
 74. Bennett MR, Macdonald K, Chan SW, Boyle JJ, Weissberg PL. Cooperative interactions between RB and p53 regulate cell proliferation, cell senescence, and apoptosis in human vascular smooth muscle cells from atherosclerotic plaques. *Circ Res* 1998;82:704-12. DOI PubMed
 75. Miao SB, Xie XL, Yin YJ, et al. Accumulation of smooth muscle 22a protein accelerates senescence of vascular smooth muscle cells via stabilization of p53 in vitro and in vivo. *Arterioscler Thromb Vasc Biol* 2017;37:1849-59. DOI PubMed
 76. Gardner SE, Humphry M, Bennett MR, Clarke MC. Senescent vascular smooth muscle cells drive inflammation through an interleukin-1 α -dependent senescence-associated secretory phenotype. *Arterioscler Thromb Vasc Biol* 2015;35:1963-74. DOI PubMed PMC
 77. Uryga AK, Grootaert MOJ, Garrido AM, et al. Telomere damage promotes vascular smooth muscle cell senescence and immune cell recruitment after vessel injury. *Commun Biol* 2021;4:611. DOI PubMed PMC
 78. Warboys CM, de Luca A, Amini N, et al. Disturbed flow promotes endothelial senescence via a p53-dependent pathway. *Arterioscler Thromb Vasc Biol* 2014;34:985-95. DOI PubMed
 79. Ungvari Z, Kaley G, de Cabo R, Sonntag WE, Csizsar A. Mechanisms of vascular aging: new perspectives. *J Gerontol A Biol Sci Med Sci* 2010;65:1028-41. DOI PubMed PMC
 80. Chilton W, O'Brien B, Charchar F. Telomeres, aging and exercise: guilty by association? *Int J Mol Sci* 2017;18:2573. DOI PubMed PMC
 81. Cawthon RM, Smith KR, O'Brien E, Sivatchenko A, Kerber RA. Association between telomere length in blood and mortality in people aged 60 years or older. *Lancet* 2003;361:393-5. DOI PubMed
 82. Benetos A, Toupance S, Gautier S, et al. Short leukocyte telomere length precedes clinical expression of atherosclerosis: the blood-and-muscle model. *Circ Res* 2018;122:616-23. DOI PubMed PMC
 83. Haycock PC, Heydon EE, Kaptoge S, Butterworth AS, Thompson A, Willeit P. Leucocyte telomere length and risk of cardiovascular disease: systematic review and meta-analysis. *BMJ* 2014;349:g4227. DOI PubMed PMC
 84. Nakajima T, Schulte S, Warrington KJ, et al. T-cell-mediated lysis of endothelial cells in acute coronary syndromes. *Circulation* 2002;105:570-5. DOI PubMed
 85. Calvert PA, Liew TV, Gorenne I, et al. Leukocyte telomere length is associated with high-risk plaques on virtual histology intravascular ultrasound and increased proinflammatory activity. *Arterioscler Thromb Vasc Biol* 2011;31:2157-64. DOI PubMed
 86. Cudejko C, Wouters K, Fuentes L, et al. p16INK4a deficiency promotes IL-4-induced polarization and inhibits proinflammatory signaling in macrophages. *Blood* 2011;118:2556-66. DOI PubMed PMC
 87. Sen P, Shah PP, Nativo R, Berger SL. Epigenetic mechanisms of longevity and aging. *Cell* 2016;166:822-39. DOI PubMed PMC
 88. Yang N, Sen P. The senescent cell epigenome. *Aging (Albany NY)* 2018;10:3590-609. DOI PubMed PMC
 89. Ellulu MS, Patimah I, Khaza'i H, Rahmat A, Abed Y. Obesity and inflammation: the linking mechanism and the complications. *Arch Med Sci* 2017;13:851-63. DOI PubMed PMC
 90. Cruzen C, Colman RJ. Effects of caloric restriction on cardiovascular aging in non-human primates and humans. *Clin Geriatr Med* 2009;25:733-43, ix. DOI PubMed PMC
 91. Colman RJ, Anderson RM, Johnson SC, et al. Caloric restriction delays disease onset and mortality in rhesus monkeys. *Science* 2009;325:201-4. DOI PubMed PMC
 92. Walford RL, Harris SB, Gunion MW. The calorically restricted low-fat nutrient-dense diet in Biosphere 2 significantly lowers blood glucose, total leukocyte count, cholesterol, and blood pressure in humans. *Proc Natl Acad Sci U S A* 1992;89:11533-7. DOI PubMed PMC
 93. Heckman GA, McKelvie RS. Cardiovascular aging and exercise in healthy older adults. *Clin J Sport Med* 2008;18:479-85. DOI PubMed
 94. Pimentel AE, Gentile CL, Tanaka H, Seals DR, Gates PE. Greater rate of decline in maximal aerobic capacity with age in endurance-trained than in sedentary men. *J Appl Physiol (1985)* 2003;94:2406-13. DOI PubMed
 95. Goldspink DF. Ageing and activity: their effects on the functional reserve capacities of the heart and vascular smooth and skeletal muscles. *Ergonomics* 2005;48:1334-51. DOI PubMed
 96. Schulman SP, Fleg JL, Goldberg AP, et al. Continuum of cardiovascular performance across a broad range of fitness levels in healthy older men. *Circulation* 1996;94:359-67. DOI PubMed
 97. Ding N, Sang Y, Chen J, et al. Cigarette smoking, smoking cessation, and long-term risk of 3 major atherosclerotic diseases. *J Am Coll Cardiol* 2019;74:498-507. DOI PubMed PMC
 98. Zeilinger S, Kühnel B, Klopp N, et al. Tobacco smoking leads to extensive genome-wide changes in DNA methylation. *PLoS One* 2013;8:e63812. DOI PubMed PMC
 99. Lu AT, Quach A, Wilson JG, et al. DNA methylation GrimAge strongly predicts lifespan and healthspan. *Aging (Albany NY)* 2019;11:303-27. DOI PubMed PMC
 100. Pope CA 3rd, Burnett RT, Thurston GD, et al. Cardiovascular mortality and long-term exposure to particulate air pollution: epidemiological evidence of general pathophysiological pathways of disease. *Circulation* 2004;109:71-7. DOI PubMed
 101. Brook RD, Franklin B, Cascio W, et al; Expert Panel on Population and Prevention Science of the American Heart Association. Air

- pollution and cardiovascular disease: a statement for healthcare professionals from the Expert Panel on Population and Prevention Science of the American Heart Association. *Circulation* 2004;109:2655-71. DOI PubMed
102. Levine ME, Lu AT, Quach A, et al. An epigenetic biomarker of aging for lifespan and healthspan. *Aging (Albany NY)* 2018;10:573-91. DOI PubMed PMC
103. Ammous F, Zhao W, Ratliff SM, et al. Epigenetic age acceleration is associated with cardiometabolic risk factors and clinical cardiovascular disease risk scores in African Americans. *Clin Epigenetics* 2021;13:55. DOI PubMed PMC
104. Horvath S. DNA methylation age of human tissues and cell types. *Genome Biol* 2013;14:R115. DOI PubMed PMC
105. Gilsbach R, Preissl S, Grüning BA, et al. Dynamic DNA methylation orchestrates cardiomyocyte development, maturation and disease. *Nat Commun* 2014;5:5288. DOI PubMed PMC
106. Westerman K, Sebastiani P, Jacques P, Liu S, DeMeo D, Ordovas JM. DNA methylation modules associate with incident cardiovascular disease and cumulative risk factor exposure. *Clin Epigenetics* 2019;11:142. DOI PubMed PMC
107. Greco CM, Kunderfranco P, Rubino M, et al. DNA hydroxymethylation controls cardiomyocyte gene expression in development and hypertrophy. *Nat Commun* 2016;7:12418. DOI PubMed PMC
108. Zaina S, Heyn H, Carmona FJ, et al. DNA methylation map of human atherosclerosis. *Circ Cardiovasc Genet* 2014;7:692-700. DOI PubMed
109. Jiang YZ, Manduchi E, Stoeckert CJ Jr, Davies PF. Arterial endothelial methylome: differential DNA methylation in atherosusceptible disturbed flow regions in vivo. *BMC Genomics* 2015;16:506. DOI PubMed PMC
110. Yuan T, Jiao Y, de Jong S, Ophoff RA, Beck S, Teschendorff AE. An integrative multi-scale analysis of the dynamic DNA methylation landscape in aging. *PLoS Genet* 2015;11:e1004996. DOI PubMed PMC
111. Ehrlich M. DNA methylation in cancer: too much, but also too little. *Oncogene* 2002;21:5400-13. DOI PubMed
112. De Cecco M, Ito T, Petrashen AP, et al. L1 drives IFN in senescent cells and promotes age-associated inflammation. *Nature* 2019;566:73-8. DOI PubMed PMC
113. Booth MJ, Ost TW, Beraldi D, et al. Oxidative bisulfite sequencing of 5-methylcytosine and 5-hydroxymethylcytosine. *Nat Protoc* 2013;8:1841-51. DOI PubMed PMC
114. Fyodorov DV, Zhou BR, Skoultchi AI, Bai Y. Emerging roles of linker histones in regulating chromatin structure and function. *Nat Rev Mol Cell Biol* 2018;19:192-206. DOI PubMed PMC
115. Bannister AJ, Kouzarides T. Regulation of chromatin by histone modifications. *Cell Res* 2011;21:381-95. DOI PubMed PMC
116. Papait R, Cattaneo P, Kunderfranco P, et al. Genome-wide analysis of histone marks identifying an epigenetic signature of promoters and enhancers underlying cardiac hypertrophy. *Proc Natl Acad Sci U S A* 2013;110:20164-9. DOI PubMed PMC
117. Benayoun BA, Pollina EA, Ucar D, et al. H3K4me3 breadth is linked to cell identity and transcriptional consistency. *Cell* 2014;158:673-88. DOI PubMed PMC
118. Thienpont B, Aronsen JM, Robinson EL, et al. The H3K9 dimethyltransferases EHMT1/2 protect against pathological cardiac hypertrophy. *J Clin Invest* 2017;127:335-48. DOI PubMed PMC
119. Zhang QJ, Chen HZ, Wang L, Liu DP, Hill JA, Liu ZP. The histone trimethyllysine demethylase JMJD2A promotes cardiac hypertrophy in response to hypertrophic stimuli in mice. *J Clin Invest* 2011;121:2447-56. DOI PubMed PMC
120. Haberland M, Montgomery RL, Olson EN. The many roles of histone deacetylases in development and physiology: implications for disease and therapy. *Nat Rev Genet* 2009;10:32-42. DOI PubMed PMC
121. Montgomery RL, Davis CA, Potthoff MJ, et al. Histone deacetylases 1 and 2 redundantly regulate cardiac morphogenesis, growth, and contractility. *Genes Dev* 2007;21:1790-802. DOI PubMed PMC
122. Montgomery RL, Potthoff MJ, Haberland M, et al. Maintenance of cardiac energy metabolism by histone deacetylase 3 in mice. *J Clin Invest* 2008;118:3588-97. DOI PubMed PMC
123. Imai S, Guarente L. NAD⁺ and sirtuins in aging and disease. *Trends Cell Biol* 2014;24:464-71. DOI PubMed PMC
124. Potente M, Ghaeni L, Baldessari D, et al. SIRT1 controls endothelial angiogenic functions during vascular growth. *Genes Dev* 2007;21:2644-58. DOI PubMed PMC
125. Vassallo PF, Simoncini S, Ligi I, et al. Accelerated senescence of cord blood endothelial progenitor cells in premature neonates is driven by SIRT1 decreased expression. *Blood* 2014;123:2116-26. DOI PubMed
126. Zheng Z, Chen H, Li J, et al. Sirtuin 1-mediated cellular metabolic memory of high glucose via the LKB1/AMPK/ROS pathway and therapeutic effects of metformin. *Diabetes* 2012;61:217-28. DOI PubMed PMC
127. Sleiman M, Jha P, Houtkooper R, Williams RW, Wang X, Auwerx J. The gene-regulatory footprint of aging highlights conserved central regulators. *Cell Rep* 2020;32:108203. DOI PubMed PMC
128. Bahar R, Hartmann CH, Rodriguez KA, et al. Increased cell-to-cell variation in gene expression in ageing mouse heart. *Nature* 2006;441:1011-4. DOI PubMed
129. Muris Consortium. A single-cell transcriptomic atlas characterizes ageing tissues in the mouse. *Nature* 2020;583:590-5. DOI
130. Eddy SR. Non-coding RNA genes and the modern RNA world. *Nat Rev Genet* 2001;2:919-29. DOI PubMed
131. Rossi M, Gorospe M. Noncoding RNAs controlling telomere homeostasis in senescence and aging. *Trends Mol Med* 2020;26:422-33. DOI PubMed PMC
132. Grillari J, Grillari-Voglauer R. Novel modulators of senescence, aging, and longevity: small non-coding RNAs enter the stage. *Exp Gerontol* 2010;45:302-11. DOI PubMed
133. Thum T, Gross C, Fiedler J, et al. MicroRNA-21 contributes to myocardial disease by stimulating MAP kinase signalling in fibroblasts. *Nature* 2008;456:980-4. DOI PubMed
134. Jazbutyte V, Fiedler J, Kneitz S, et al. MicroRNA-22 increases senescence and activates cardiac fibroblasts in the aging heart. *Age*

- (*Dordr*) 2013;35:747-62. DOI PubMed PMC
135. Boon RA, Iekushi K, Lechner S, et al. MicroRNA-34a regulates cardiac ageing and function. *Nature* 2013;495:107-10. DOI PubMed
 136. Ito T, Yagi S, Yamakuchi M. MicroRNA-34a regulation of endothelial senescence. *Biochem Biophys Res Commun* 2010;398:735-40. DOI PubMed
 137. Badi I, Burba I, Ruggeri C, et al. MicroRNA-34a Induces Vascular Smooth Muscle Cells Senescence by SIRT1 Downregulation and Promotes the Expression of Age-Associated Pro-inflammatory Secretory Factors. *J Gerontol A Biol Sci Med Sci* 2015;70:1304-11. DOI PubMed
 138. Menghini R, Casagrande V, Cardellini M, et al. MicroRNA 217 modulates endothelial cell senescence via silent information regulator 1. *Circulation* 2009;120:1524-32. DOI PubMed
 139. Boon RA, Seeger T, Heydt S, et al. MicroRNA-29 in aortic dilation: implications for aneurysm formation. *Circ Res* 2011;109:1115-9. DOI PubMed
 140. de Lucia C, Komici K, Borghetti G, et al. microRNA in cardiovascular aging and age-related cardiovascular diseases. *Front Med (Lausanne)* 2017;4:74. DOI PubMed PMC
 141. Verduci L, Tarcitano E, Strano S, Yarden Y, Blandino G. CircRNAs: role in human diseases and potential use as biomarkers. *Cell Death Dis* 2021;12:468. DOI PubMed PMC
 142. Altesha MA, Ni T, Khan A, Liu K, Zheng X. Circular RNA in cardiovascular disease. *J Cell Physiol* 2019;234:5588-600. DOI PubMed
 143. Du WW, Yang W, Chen Y, et al. Foxo3 circular RNA promotes cardiac senescence by modulating multiple factors associated with stress and senescence responses. *Eur Heart J* 2017;38:1402-12. DOI PubMed
 144. Holdt LM, Stahring A, Sass K, et al. Circular non-coding RNA ANRIL modulates ribosomal RNA maturation and atherosclerosis in humans. *Nat Commun* 2016;7:12429. DOI PubMed PMC
 145. Burd CE, Jeck WR, Liu Y, Sanoff HK, Wang Z, Sharpless NE. Expression of linear and novel circular forms of an INK4/ARF-associated non-coding RNA correlates with atherosclerosis risk. *PLoS Genet* 2010;6:e1001233. DOI PubMed PMC
 146. Lozano-Vidal N, Bink DI, Boon RA. Long noncoding RNA in cardiac aging and disease. *J Mol Cell Biol* 2019;11:860-7. DOI PubMed PMC
 147. Han P, Li W, Lin CH, et al. A long noncoding RNA protects the heart from pathological hypertrophy. *Nature* 2014;514:102-6. DOI PubMed PMC
 148. Wang Z, Zhang XJ, Ji YX, et al. The long noncoding RNA Chaer defines an epigenetic checkpoint in cardiac hypertrophy. *Nat Med* 2016;10:1131-9. DOI PubMed PMC
 149. Boon RA, Hofmann P, Michalik KM, et al. Long noncoding RNA Meg3 controls endothelial cell aging and function: implications for regenerative angiogenesis. *J Am Coll Cardiol* 2016;68:2589-91. DOI PubMed
 150. Chen J, Zou Q, Lv D, et al. Comprehensive transcriptional landscape of porcine cardiac and skeletal muscles reveals differences of aging. *Oncotarget* 2018;9:1524-41. DOI PubMed PMC
 151. Hang CT, Yang J, Han P, et al. Chromatin regulation by Brg1 underlies heart muscle development and disease. *Nature* 2010;466:62-7. DOI PubMed PMC
 152. Han P, Li W, Yang J, et al. Epigenetic response to environmental stress: assembly of BRG1-G9a/GLP-DNMT3 repressive chromatin complex on Myh6 promoter in pathologically stressed hearts. *Biochim Biophys Acta* 2016;1863:1772-81. DOI PubMed PMC
 153. Pierre R, Kadoch C. Mammalian SWI/SNF complexes in cancer: emerging therapeutic opportunities. *Curr Opin Genet Dev* 2017;42:56-67. DOI PubMed PMC
 154. Centore RC, Sandoval GJ, Soares LMM, Kadoch C, Chan HM. Mammalian SWI/SNF chromatin remodeling complexes: emerging mechanisms and therapeutic strategies. *Trends Genet* 2020;36:936-50. DOI PubMed
 155. Olive M, Harten I, Mitchell R, et al. Cardiovascular pathology in Hutchinson-Gilford progeria: correlation with the vascular pathology of aging. *Arterioscler Thromb Vasc Biol* 2010;30:2301-9. DOI PubMed PMC
 156. Scaffidi P, Misteli T. Reversal of the cellular phenotype in the premature aging disease Hutchinson-Gilford progeria syndrome. *Nat Med* 2005;11:440-5. DOI PubMed PMC
 157. Shumaker DK, Dechat T, Kohlmaier A, et al. Mutant nuclear lamin A leads to progressive alterations of epigenetic control in premature aging. *Proc Natl Acad Sci U S A* 2006;103:8703-8. DOI PubMed PMC
 158. Baker DJ, Childs BG, Durik M, et al. Naturally occurring p16(Ink4a)-positive cells shorten healthy lifespan. *Nature* 2016;530:184-9. DOI PubMed PMC
 159. Kirkland JL, Tchkonja T, Zhu Y, Niedernhofer LJ, Robbins PD. The Clinical Potential of Senolytic Drugs. *J Am Geriatr Soc* 2017;65:2297-301. DOI PubMed PMC
 160. Tchkonja T, Kirkland JL. Aging, cell senescence, and chronic disease: emerging therapeutic strategies. *JAMA* 2018;320:1319-20. DOI PubMed
 161. Laberge RM, Zhou L, Sarantos MR, et al. Glucocorticoids suppress selected components of the senescence-associated secretory phenotype. *Aging Cell* 2012;11:569-78. DOI PubMed PMC
 162. Wang R, Yu Z, Sunchu B, et al. Rapamycin inhibits the secretory phenotype of senescent cells by a Nrf2-independent mechanism. *Aging Cell* 2017;16:564-74. DOI PubMed PMC
 163. Moiseeva O, Deschênes-Simard X, St-Germain E, et al. Metformin inhibits the senescence-associated secretory phenotype by interfering with IKK/NF- κ B activation. *Aging Cell* 2013;12:489-98. DOI PubMed
 164. Chen W, Wang X, Wei G, et al. Single-cell transcriptome analysis reveals six subpopulations reflecting distinct cellular fates in

- senescent mouse embryonic fibroblasts. *Front Genet* 2020;11:867. DOI PubMed PMC
165. Shi C, Wang L, Sen P. The eroding chromatin landscape of aging stem cells. *Transl Med Aging* 2020;4:121-31. DOI PubMed PMC
166. Capell BC, Drake AM, Zhu J, et al. MLL1 is essential for the senescence-associated secretory phenotype. *Genes Dev* 2016;30:321-36. DOI PubMed PMC
167. Tasdemir N, Banito A, Roe JS, et al. BRD4 Connects Enhancer Remodeling to Senescence Immune Surveillance. *Cancer Discov* 2016;6:612-29. DOI PubMed PMC
168. Phillips MI, Costales J, Lee RJ, Oliveira E, Burns AB. Antisense therapy for cardiovascular diseases. *Curr Pharm Des* 2015;21:4417-26. DOI PubMed
169. Yun MH. Changes in regenerative capacity through lifespan. *Int J Mol Sci* 2015;16:25392-432. DOI PubMed PMC
170. Li Y, He L, Huang X, et al. Genetic lineage tracing of nonmyocyte population by dual recombinases. *Circulation* 2018;138:793-805. DOI PubMed
171. Ocampo A, Reddy P, Martinez-Redondo P, et al. In vivo amelioration of age-associated hallmarks by partial reprogramming. *Cell* 2016;167:1719-33.e12. DOI PubMed PMC
172. Aghajanian H, Kimura T, Rurik JG, et al. Targeting cardiac fibrosis with engineered T cells. *Nature* 2019;573:430-3. DOI PubMed PMC
173. North BJ, Sinclair DA. The intersection between aging and cardiovascular disease. *Circ Res* 2012;110:1097-108. DOI PubMed PMC
174. Wang Z, Zang C, Rosenfeld JA, et al. Combinatorial patterns of histone acetylations and methylations in the human genome. *Nat Genet* 2008;40:897-903. DOI PubMed PMC
175. Bonn S, Zinzen RP, Girardot C, et al. Tissue-specific analysis of chromatin state identifies temporal signatures of enhancer activity during embryonic development. *Nat Genet* 2012;44:148-56. DOI PubMed
176. Schübeler D, MacAlpine DM, Scalzo D, et al. The histone modification pattern of active genes revealed through genome-wide chromatin analysis of a higher eukaryote. *Genes Dev* 2004;18:1263-71. DOI PubMed PMC
177. Poleshko A, Shah PP, Gupta M, et al. Genome-nuclear lamina interactions regulate cardiac stem cell lineage restriction. *Cell* 2017;171:573-87.e14. DOI PubMed PMC
178. Hahn MA, Wu X, Li AX, Hahn T, Pfeifer GP. Relationship between gene body DNA methylation and intragenic H3K9me3 and H3K36me3 chromatin marks. *PLoS One* 2011;6:e18844. DOI PubMed PMC
179. Wiles ET, Selker EU. H3K27 methylation: a promiscuous repressive chromatin mark. *Curr Opin Genet Dev* 2017;43:31-7. DOI PubMed PMC
180. Dambacher S, Hahn M, Schotta G. The compact view on heterochromatin. *Cell Cycle* 2013;12:2925-6. DOI PubMed PMC