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## Supplementary Material

**Different fitness to arabinogalactan side chains is supported by diversified specificity of glycoside hydrolase family 39 enzymes in the genus *Bifidobacterium***

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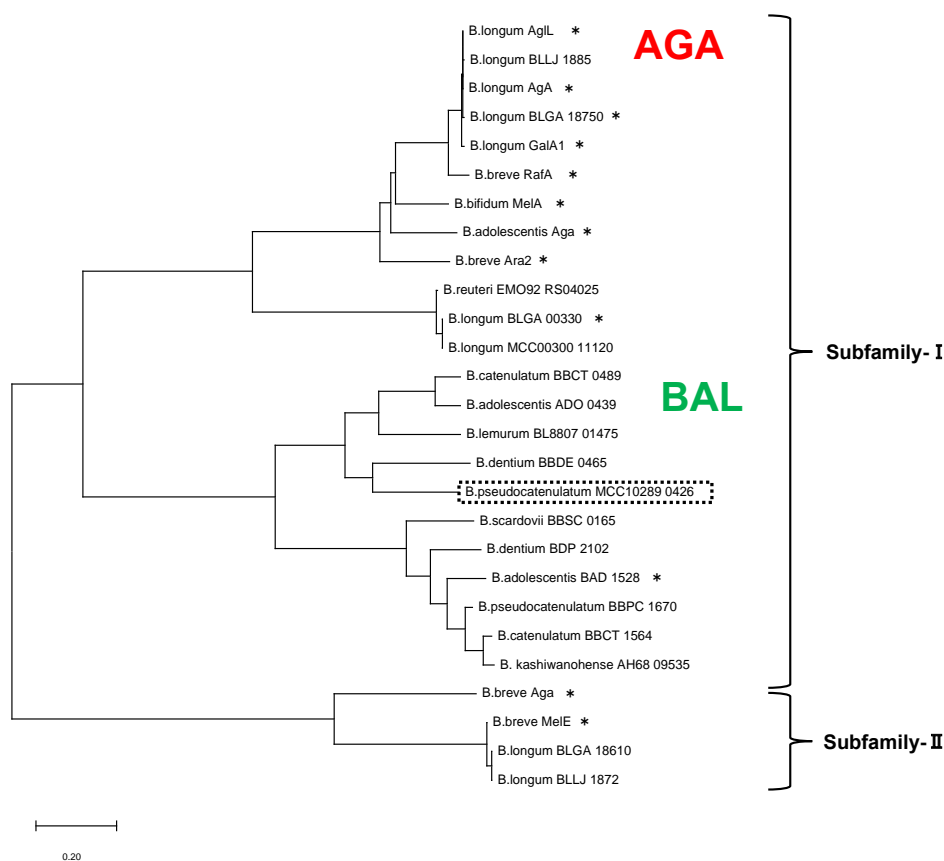
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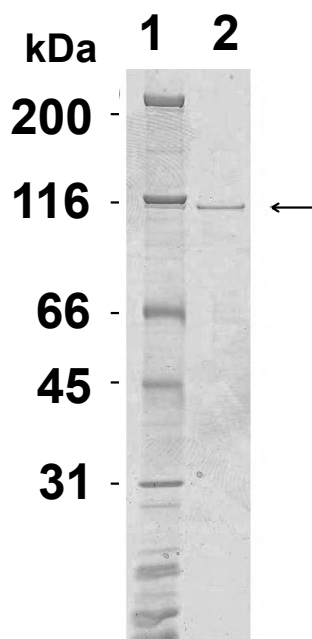
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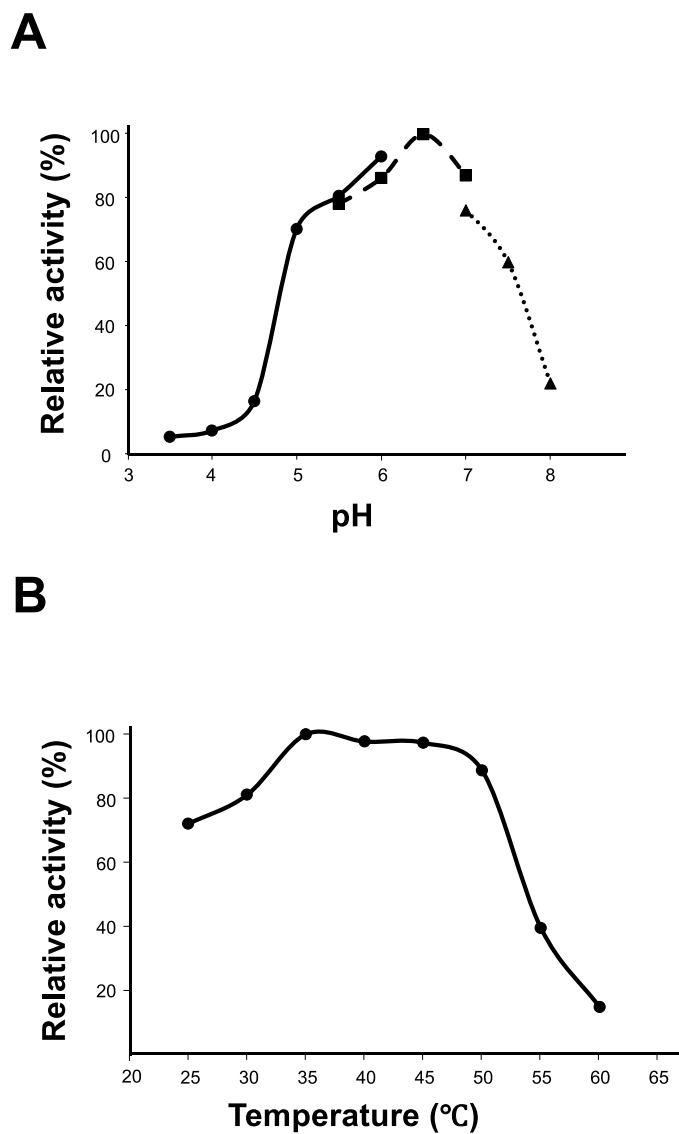
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 18 **Supplementary Figure 1.** The phylogenetic relationships of bifidobacterial GH36  $\alpha$ -  
 19 galactosidases and  $\beta$ -L-arabinopyranosidases. The phylogenetic tree of *B.*  
 20 *pseudocatenulatum* MCC10289\_0426 with homologous proteins from bifidobacteria  
 21 was constructed by the neighbor-joining method using the aligned sequences; for the  
 22 construction, the program Clustal W was implemented in the MEGA11 software. *B.*  
 23 *pseudocatenulatum*\_MCC10289\_0426 is indicated by a dashed-line box. The protein  
 24 names or locus tags are shown alongside *Bifidobacterium* strains as follows: *B.*  
 25 *adolescentis* Aga (GenBank ID: AAD30994.2), *B. bifidum* MelA (ABD96085.1), *B.*  
 26 *breve* Aga (AAK96217.2), *B. breve* Aga2 (ABB76662.1), *B. longum* GalA1  
 27 (ACD98928.1), *B. longum* AgA (AAN25312.1), *B. longum* AgIL (AAG02023.1), *B.*  
 28 *breve* RafA (ABE96531.1), *B. breve* MeIE (ABE96518.1), *B. longum* BLLJ\_1872  
 29 (BAJ67536.1), *B. longum* BLLJ\_1885 (BAJ67549.1), *B. longum* BlAga3  
 30 (BBV22622.1), *B. longum* BlAga1 (BBV24464.1), and *B. longum* BlAga2  
 31 (BBV24450.1), *B. longum* MCC00300\_11120 (WP\_077381863.1), *B. reuteri*  
 32 EMO92\_RS04025 (WP\_150335335.1), *B. catenulatum* BBCT\_0489 (BAR01457.1),  
 33 *B. adolescentis* BADO\_0439 (AII75864.1), *B. lemurum* BL8807\_01475

34 (QOL34630.1), *B. dentium* BBDE\_0465 (BAQ26459.1), *B. adolescentis* BAD\_1528  
35 (BBD14080.1), *B. pseudocatenulatum* BBPC\_1670 (BAR04348.1), *B. catenulatum*  
36 BBCT\_1564 (BAR02532.1), *B. kashiwanohense* AH68\_09535 (AIZ15229.1), *B.*  
37 *dentium* BDP\_2102 (ADB10663.1), and *B. scardovii* BBSC\_0165 (BAQ30245.1).  
38 Asterisk indicates characterized enzymes.

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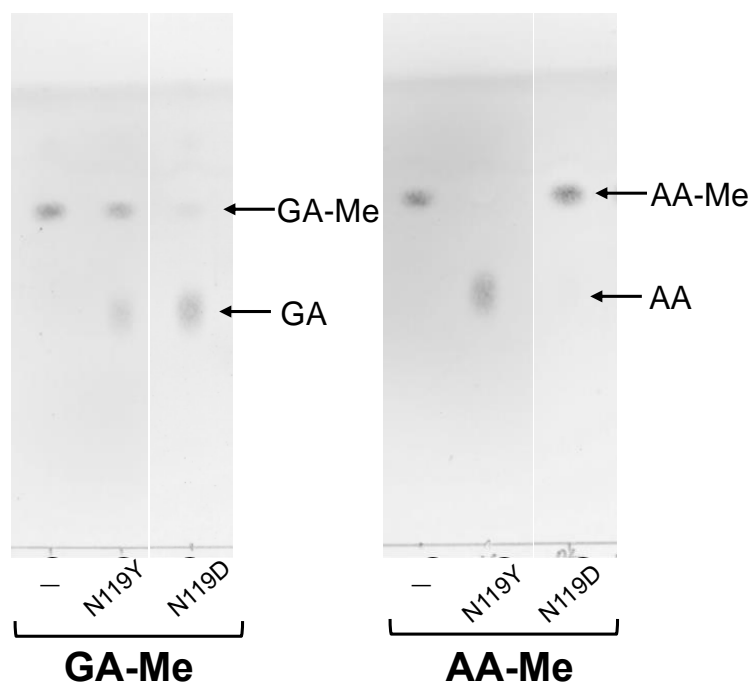


40 **Supplementary Figure 2.** SDS-PAGE analysis of recombinant AAFase. Purified  
41 AAFase was electrophoresed on a 5–20% gradient polyacrylamide gel and stained with  
42 Coomassie Brilliant Blue R-250. Lane 1, molecular size marker; lane 2, purified  
43 AAFase. Arrow indicates target protein at expected molecular size.



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45 **Supplementary Figure 3.** Optimal pH and temperature of AAFase. (A) pH dependence  
46 of AAFase activity in various buffers at 45 °C for 20 min. Sodium acetate buffer (closed  
47 circle and solid line), MES buffer (closed square and dashed line), and HEPES buffer  
48 (closed triangle and dotted line) were used. Enzyme activities are expressed as a  
49 percentage of the activity in MES buffer at pH 6.5. (B) The temperature dependence of  
50 AAFase activity at pH 6.5 for 20 min. The enzymatic activities are expressed as the  
51 percentage of the activity at 35 °C.

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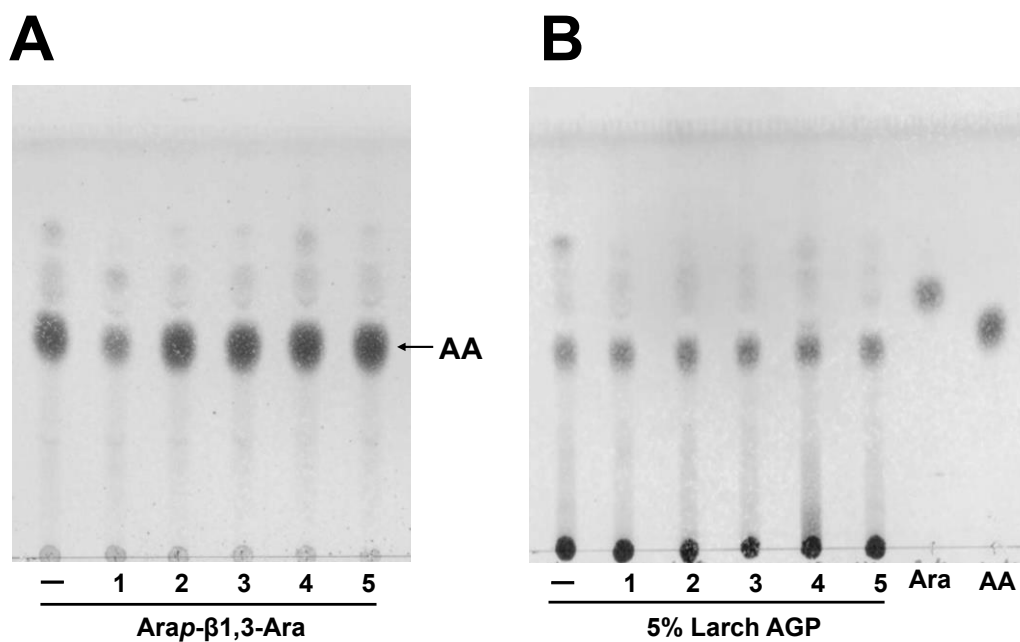


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55 **Supplementary Figure 4.** TLC analysis of the enzymatic activity of Gafase N119Y and Gafase  
56 N119D. GA-Me and AA-Me are the abbreviation of  $\alpha$ -D-Gal-(1 $\rightarrow$ 3)- $\alpha$ -L-Araf-OMe and  $\beta$ -L-Arap-  
57 (1 $\rightarrow$ 3)- $\alpha$ -L-Araf-OMe, respectively. 1 mM GA-Me and AA-Me were incubated with absence (-) or  
58 presence (+) of the two types of mutants in 50 mM sodium acetate buffer (pH 5.0) at 37 °C for 18 h. As  
59 enzymes, the elution fraction containing imidazole after His-tag affinity purification were used in this  
60 study, because desalted Gafase N119D got lost enzymatic activity. The amount of enzyme was matched  
61 when the same enzyme was used in order to see the activity on different substrates, although the quantity  
62 of N119Y and N119D were different. The reaction mixtures were analyzed by TLC, as described in main  
63 text.

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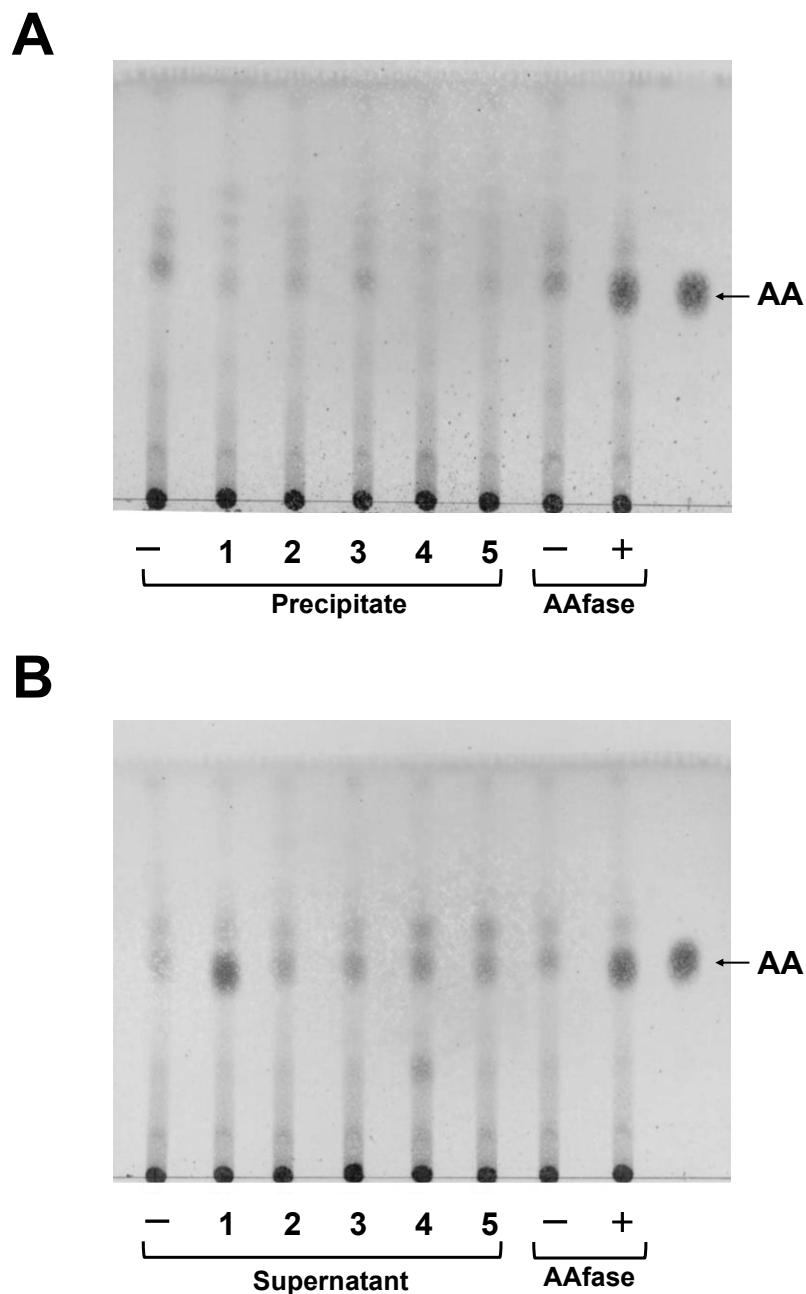
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67 **Supplementary Figure 5.** TLC analysis of the culture supernatants after incubation for68 41 hours in  $\beta$ -L-Arap-(1 $\rightarrow$ 3)-L-Ara (A) and and 48 hours in larch AGP (B). 1, *B.*69 *pseudocatenulatum* MCC10289; 2, *B. pseudocatenulatum* MCC10285; 3, *B.*70 *pseudocatenulatum* MCC10311; 4, *B. kashiwanohense* MCC10250; 5, *B.*71 *pseudocatenulatum* JCM1200<sup>T</sup>. As a control, the medium without any bacteria (–) was72 used. AA is the abbreviation for  $\beta$ -L-Arap-(1 $\rightarrow$ 3)-L-Ara.

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76 **Supplementary Figure 6.** TLC analysis of the bacterial enzymatic activities with 2.5%

77 larch AGP. Larch AGP was incubated with bacterial cell fractions (A) and culture

78 supernatant (B) after cultivation of *B. pseudocatenulatum* MCC10289 (1), *B.*

79 *pseudocatenulatum* MCC10285 (2), *B. pseudocatenulatum* MCC10311 (3), *B.*

80 *kashiwanohense* MCC10250 (4), and *B. pseudocatenulatum* JCM1200T (5) grown in

81 larch AGP after 18 h incubation. As a control, the medium without any bacteria (-) was

82 used for enzymatic activity assay. As a control, larch AGP was incubated in the

83 absence (–) or presence (+) of AAFase.

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