





























microenvironment has classically been associated with “space clearance” for GBM via excitotoxic damage. However, an increase in glutamate activation of mGluR5 can also increase the NAS/melatonin ratio. As such, much of the plasticity within the GBM/GSC microenvironment may be driven by processes that control mitochondrial metabolism and the mitochondrial melatonergic pathway. Mutations, epigenetic alterations and changes in the diverse array of receptors and intracellular signalling pathways may mediate their effects via their interactions with higher-order processes acting to regulate the mitochondrial melatonergic pathway.

Chemo-resistance may be seen in this context, whereby resistance to temozolomide or tamoxifen that often emerge following surgery for GBM/GSC may be seen as a mechanism in which GBM/GSC manage to maintain this control over the mitochondrial melatonergic pathway across different cells of the tumour microenvironment. This does not necessarily suggest total inhibition of melatonin production in the cells of the GBM/GSC microenvironment, but it does indicate that regulation of this pathway is of crucial importance. Future research should clarify the regulation of NAS and melatonin in different cell compartments. For example, melatonin uptake into mitochondria in oocytes is mediated by the peptide transporter (PEPT) 1/2<sup>[133]</sup>, the regulation of which may be crucial to the compartmental regulation of melatonin under specific conditions. Other changes in the cell may arise as a consequence of this, and possibly correlate with GBM/GSC maintenance, but would be arising from the need to regulate the mitochondrial melatonergic pathway and not because these other changes are crucial per se. Changes in circadian genes and the impact of the circadian rhythm may also be seen in this context. Such a model highlights the hierarchical influence of patterned transcriptions driven by mitochondrial function, and the key role that alterations of the mitochondrial melatonergic pathways have in driving such patterned changes.

As 14-3-3 stabilizes AANAT, the mitochondrial melatonergic pathway can also be regulated by 14-3-3 protein levels. 14-3-3 is expressed in mitochondria, with down-regulation of 14-3-3 $\zeta$  induced by ceramide, sensitizing GBM to apoptosis, indicating a role for 14-3-3 $\zeta$  in GBM/GSC chemoresistance<sup>[65]</sup>. Heightened expression levels of 14-3-3 $\zeta$  in GBM patients are associated with a poor prognosis<sup>[64]</sup>. As to whether this is mediated by 14-3-3 $\zeta$  stabilization of AANAT, leading to enhanced activation of the mitochondrial melatonergic pathway that is biased towards an increase in NAS production and TrkB activation requires investigation. A decrease in 14-3-3 $\gamma$  inhibits the stem-like qualities of GSC<sup>[13]</sup>, whilst the downregulation of 14-3-3 $\beta$  induces senescence in GBM/GSC<sup>[134]</sup>. Clearly, different 14-3-3 isoforms are important to GBM/GSC survival, which may be closely linked to 14-3-3 regulation of the mitochondrial melatonergic pathway.

Overall, the mitochondrial melatonergic pathway may be intimately linked to the plethora of diverse data associated with GBM/GSC pathophysiology. The melatonergic pathway has been evident from the beginning of cell evolution, being present in the first bacteria to become integrated into an early cellular structure that ultimately evolved into mitochondria<sup>[135]</sup>. Consequently, the melatonergic pathway has been an integral aspect of mitochondrial and cellular evolution and therefore, with the various mitochondrial factors, such as sirtuins, and processes such as metabolism, that underpin the function and survival of GBM/GSC and the cells of the tumour microenvironment.

## FUTURE RESEARCH

Does the increased YY1 in tumours associate with heightened levels of CYP1B1, mGluR5, P2Y1, O-demethylation and CYP2C19, and thereby with the “backward” conversion of melatonin to NAS and TrkB-driven GSC, survival and proliferation?

Does YY1-miR-135b inhibition of Bmal1 modulate OXPHOS in GBM/GSC over the circadian rhythm?

What is the influence of the circadian rhythm on gene expression in GBM/GSC, and in the cells of the tumour microenvironment, including miRNAs?

Do GBM/GSC determine the nature of the tumour microenvironment via the realignment of immune cell circadian rhythms and associated regulation of mitochondrial metabolism and the mitochondrial melatonergic pathway?

Does YY1-induced miR-135b regulate TNF receptor associated factor (TRAF) 2, given both miR-135b<sup>[136]</sup> and TRAF2<sup>[137]</sup> attenuate the transcriptional activity of Bmal1?

The AhR is an important regulator of the circadian rhythm, including by forming a heteromer with Bmal1, thereby inhibiting Bmal1's influence on the circadian amplitude and metabolism<sup>[138]</sup>. AhR expression levels and activation over the circadian rhythm in GBM and GSC will therefore be important to determine.

As mGluR5 activation on microglia/macrophages can increase TrkB activation and BDNF production<sup>[139]</sup>, is this mediated via increased NAS?

As butyrate, via HDAC inhibition, modulates not only mast cell activation<sup>[34]</sup> but increases GBM and GSC apoptosis<sup>[24,25]</sup>, do inducers of gut dysbiosis/permeability, such as stress, alcohol and diet, then impact on GBM/GSC pathoetiology and pathophysiology via gut dysbiosis/permeability?

In some cells, sodium butyrate can increase AhR-induced induced CYP1<sup>[140]</sup>. This will be important to determine in GBM/GSC, especially the impact, if any, of sodium butyrate on CYP1B1-driven 'backward' conversion of melatonin to NAS over the circadian rhythm.

Do activated mast cells suppress GSC pathoetiology, as indicated by the negative correlation of allergies and GBM development<sup>[141]</sup>?

Is there a role for mast cells in GBM/GSC pathophysiology, given mast cell suppression of GBM proliferation and migration, as well as their suppression of GSC self-renewal capacity and stemness markers<sup>[142]</sup>? What is the role of mast cell melatonin production<sup>[143]</sup>?

As wild-type p53 induces AhR-driven CYP1B1, does the resulting NAS suppress the apoptotic effects of wild-type p53 in GBM/GSC?

The roles of the ceramide/S1P ratio in the regulation of mitochondrial function, metabolism and melatonergic pathway in GBM/GSC and the cells of the tumour microenvironment will be important to determine.

Leucine zipper-EF-hand containing transmembrane protein (LETM) 1 is classically associated with mitochondria ionic regulation via pore formation across the mitochondrial membranes. LETM1 is highly expressed in many different types of tumours, especially in cancer stem cells, and is associated with poor prognosis<sup>[144]</sup>. As the C-terminal of LETM1 in the mitochondrial matrix has a 14-3-3-like motif, can this motif stabilize mitochondrial AANAT<sup>[145]</sup>?

As PTEN-induced kinase (PINK) 1 phosphorylates and optimizes LETM1 function, the loss of PINK1 in GBM/GSC<sup>[146]</sup> will modulate not only mitochondrial ionic regulation but potentially impact upon the mitochondrial melatonergic pathway. This requires investigation as potentially this links previously diverse bodies of data pertaining to GBM/GSC survival and chemoresistance, especially as Ca<sup>2+</sup> dysregulation is evident in GBM mitochondria<sup>[147]</sup>.

Given *Bmal1* and SIRT1 mutually induce each other in non-neoplastic cells<sup>[129]</sup>, it requires investigation as to the interactions of *Bmal1* and sirtuins in the regulation of mitochondria and the mitochondrial melatonergic pathway. Resveratrol, a SIRT1 inducer, affords protection in GBM/GSC and has recently been proposed as an adjunctive in GBM/GSC treatment<sup>[148]</sup>, with the induction of SIRT1 having effects that include regulation of the GBM/GSC microenvironment<sup>[149]</sup>. As noted above, resveratrol also inhibits AhR, suggesting that it will have an impact on the NAS/melatonin ratio. The interactions of sirtuins with resveratrol and the melatonergic pathway will be important to determine.

## TREATMENT IMPLICATIONS

It is highly likely that melatonin per se, as well as its adjunctive use with chemotherapy, will have positive benefits on GBM/GSC patient outcomes<sup>[150]</sup>.

Ultrasound-induced local BBB disruption may allow for an increased availability of the wide array of factors that have been shown in preclinical studies to suppress GBM/GSC<sup>[151]</sup>, and may allow for an increased invasion of mast cells as well as other melatonin producing cells into the GBM/GSC microenvironment. Whether mast cells or other cells chemoattracted to the GBM/GSC microenvironment can be primed for an increase in melatonin release and/or for the inhibition of the 'backward' conversion of melatonin to NAS will be interesting to investigate.

Melatonin increases miR-149<sup>[152]</sup>, which is epigenetically suppressed in GBM<sup>[153]</sup>, with miR-149 and miR-128 increasing chemosensitivity to temozolomide<sup>[154]</sup>.

Given the efficacy of butyrate in the induction of senescence in GBM<sup>[25]</sup> and apoptosis in GSC<sup>[24]</sup> as well as increasing chemosensitivity and suppressing YY1<sup>[31]</sup>, it requires investigation as to the clinical utility of sodium butyrate in the management of GBM/GSC, including as an adjunctive to chemo- and radio-therapy.

Given that the maximal toxicity of temozolomide is optimized by the upregulation of the circadian gene, *Bmal1*<sup>[5]</sup>, it would seem likely that research to clarify the roles of circadian genes, circadian rhythm and the melatonergic pathway, will provide ways of improving clinical utility and treatment outcomes.

## CONCLUSION

Chemoresistance is common in GBM/GSC, being reflective of the plasticity of response and adaptation that is inherent in the tumour microenvironment. The above would indicate that the mitochondrial melatonergic pathway might be an important aspect, if not driver, of the plasticity in the intercellular interactions of the cells of the tumour microenvironment. The incorporation of the mitochondrial melatonergic pathway provides a conceptual frame of reference that better integrates the diverse array of previously disparate data on the biological underpinnings of GBM/GSC. This has implications for understanding the ubiquitous radio- and chemo-resistance that are evident in GBM/GSC as well as providing novel, and readily achievable, treatment targets.

## DECLARATIONS

### Authors' contributions

The author contributed solely to the article.

### Availability of data and materials

Review of current literature.

### Financial support and sponsorship

None.



### Conflicts of interest

The author declared that there are no conflicts of interest.

### Ethical approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Copyright

© The Author(s) 2020.

### REFERENCES

1. Zeng AL, Yan W, Liu YW, Wang Z, Hu Q, et al. Tumour exosomes from cells harbouring PTPRZ1-MET fusion contribute to a malignant phenotype and temozolomide chemoresistance in glioblastoma. *Oncogene* 2017;36:5369-81.
2. Sharifzad F, Ghavami S, Verdi J, Mardpour S, Mollapour Sisakht M, et al. Glioblastoma cancer stem cell biology: potential theranostic targets. *Drug Resist Updat* 2019;42:35-45.
3. Beischlag TV, Anderson G, Mazzocchi G. Glioma: tryptophan catabolite and melatoninergic pathways link microRNA, 14-3-3, chromosome 4q35, epigenetic processes and other glioma biochemical changes. *Curr Pharm Des* 2016;22:1033-48.
4. Anderson G, Reiter RJ. Glioblastoma: role of mitochondria N-acetylserotonin/melatonin ratio in mediating effects of mir-451 and aryl hydrocarbon receptor and in coordinating wider biochemical changes. *Int J Tryptophan Res* 2019;12:1178646919855942.
5. Slat EA, Sponagel J, Marpegan L, Simon T, Kfoury N, et al. Cell-intrinsic, Bmal1-dependent circadian regulation of temozolomide sensitivity in glioblastoma. *J Biol Rhythms* 2017;32:121-9.
6. Abou-Antoun TJ, Hale JS, Lathia JD, Dombrowski SM. Brain cancer stem cells in adults and children: cell biology and therapeutic implications. *Neurotherapeutics* 2017;14:372-84.
7. Anderson G, Maes M. Gut dysbiosis dysregulates central and systemic homeostasis via suboptimal mitochondrial function: assessment, treatment and classification implications. *Curr Top Med Chem* 2020;20:524-39.
8. Anderson G. Daytime orexin and night-time melatonin regulation of mitochondria melatonin: roles in circadian oscillations systemically and centrally in breast cancer symptomatology. *Melatonin Res* 2019;2:1-8.
9. Zhao K, Wang L, Li T, Zhu M, Zhang C, et al. The role of miR-451 in the switching between proliferation and migration in malignant glioma cells: AMPK signaling, mTOR modulation and Rac1 activation required. *Int J Oncol* 2017;50:1989-99.
10. Jia B, Liu W, Gu J, Wang J, Lv W, et al. MiR-7-5p suppresses stemness and enhances temozolomide sensitivity of drug-resistant glioblastoma cells by targeting Yin Yang 1. *Exp Cell Res* 2019;375:73-81.
11. Li GF, Cheng YY, Li BJ, Zhang C, Zhang XX, et al. miR-375 inhibits the proliferation and invasion of glioblastoma by regulating Wnt5a. *Neoplasia* 2019;66:350-6.
12. Anderson G. The effects of melatonin on signaling pathways and molecules involved in glioma: Melatonin and glioblastoma: pathophysiology and treatment. *Fundam Clin Pharmacol* 2020;34:189-91.
13. Im CN. Combination treatment with PPAR $\gamma$  ligand and its specific inhibitor GW9662 downregulates BIS and 14-3-3 gamma, inhibiting stem-like properties in glioblastoma cells. *Biomed Res Int* 2017;2017:5832824.
14. Yang X, Cao W, Zhou J, Zhang W, Zhang X, et al. 14-3-3 $\zeta$  positive expression is associated with a poor prognosis in patients with glioblastoma. *Neurosurgery* 2011;68:932-8; discussion 938.
15. Lawn S, Krishna N, Pisklakova A, Qu X, Fenstermacher DA, et al. Neurotrophin signaling via TrkB and TrkC receptors promotes the growth of brain tumor-initiating cells. *J Biol Chem* 2015;290:3814-24.
16. Pinet S, Bessette B, Vedrenne N, Lacroix A, Richard L, et al. TrkB-containing exosomes promote the transfer of glioblastoma aggressiveness to YKL-40-inactivated glioblastoma cells. *Oncotarget* 2016;7:50349-64.
17. Bostian AC, Maddukuri L, Reed MR, Savenka T, Hartman JH, et al. Kynurenine signaling increases DNA polymerase kappa expression and promotes genomic instability in glioblastoma cells. *Chem Res Toxicol* 2016;29:101-8.
18. Jin UH, Karki K, Cheng Y, Michelhaugh SK, Mittal S, et al. The aryl hydrocarbon receptor is a tumor suppressor-like gene in glioblastoma. *J Biol Chem* 2019;294:11342-53.
19. Riess C, Schneider B, Kehnscherper H, Gesche J, Irmscher N, et al. Activation of the kynurenine pathway in human malignancies can be suppressed by the cyclin-dependent kinase inhibitor dinaciclib. *Front Immunol* 2020;11:55.
20. Reiter RJ, Sharma R, Ma Q, Rosales-Corral S, Acuna-Castroviejo D, et al. Inhibition of mitochondrial pyruvate dehydrogenase kinase: a proposed mechanism by which melatonin causes cancer cells to overcome aerobic glycolysis, limit tumor growth and reverse insensitivity to chemotherapy. *Melatonin Res* 2019;2:105-19.
21. Markus RP, Fernandes PA, Kinker GS, da Silveira Cruz-Machado S, Marçola M. Immune-pineal axis - acute inflammatory responses coordinate melatonin synthesis by pinealocytes and phagocytes. *Br J Pharmacol* 2018;175:3239-50.
22. Anderson G. Breast cancer: occluded role of mitochondria N-acetylserotonin/melatonin ratio in co-ordinating pathophysiology. *Biochem*

- Pharmacol 2019;168:259-68.
23. Chen J, Zhao KN, Vitetta L. Effects of intestinal microbial-elaborated butyrate on oncogenic signaling pathways. *Nutrients* 2019;11:E1026.
  24. Tung B, Ma D, Wang S, Oyinlade O, Laterra J, et al. Krüppel-like factor 9 and histone deacetylase inhibitors synergistically induce cell death in glioblastoma stem-like cells. *BMC Cancer* 2018;18:1025.
  25. Nakagawa H, Sasagawa S, Itoh K. Sodium butyrate induces senescence and inhibits the invasiveness of glioblastoma cells. *Oncol Lett* 2018;15:1495-502.
  26. Baglietto L, Giles GG, English DR, Karahalios A, Hopper JL, et al. Alcohol consumption and risk of glioblastoma; evidence from the Melbourne Collaborative Cohort Study. *Int J Cancer* 2011;128:1929-34.
  27. Stärkel P, Leclercq S, de Timary P, Schnabl B. Intestinal dysbiosis and permeability: the yin and yang in alcohol dependence and alcoholic liver disease. *Clin Sci (Lond)* 2018;132:199-212.
  28. Yamawaki Y, Yoshioka N, Nozaki K, Ito H, Oda K, et al. Sodium butyrate abolishes lipopolysaccharide-induced depression-like behaviors and hippocampal microglial activation in mice. *Brain Res* 2018;1680:13-38.
  29. Catalano M, D'Alessandro G, Trettel F, Limatola C. Role of infiltrating microglia/macrophages in glioma. *Adv Exp Med Biol* 2020;1202:281-98.
  30. Pedrote MM, Motta MF, Ferretti GDS, Norberto DR, Spohr TCLS, et al. Oncogenic gain of function in glioblastoma is linked to mutant p53 amyloid oligomers. *iScience* 2020;23:100820.
  31. Wang ZT, Chen ZJ, Jiang GM, Wu YM, Liu T, et al. Histone deacetylase inhibitors suppress mutant p53 transcription via HDAC8/YY1 signals in triple negative breast cancer cells. *Cell Signal* 2016;28:506-15.
  32. Albert-Bayo M, Paracuellos I, González-Castro AM, Rodríguez-Urrutia A, Rodríguez-Lagunas MJ, et al. Intestinal mucosal mast cells: key modulators of barrier function and homeostasis. *Cells* 2019;8:E135.
  33. Vanuytsel T, van Wanrooy S, Vanheel H, Vanormelingen C, Verschuere S, et al. Psychological stress and corticotropin-releasing hormone increase intestinal permeability in humans by a mast cell-dependent mechanism. *Gut* 2014;63:1293-9.
  34. Folkerts J, Redegeld F, Folkerts G, Blokhuis B, van den Berg MPM, et al. Butyrate inhibits human mast cell activation via epigenetic regulation of FcεRI-mediated signaling. *Allergy* 2020; epub ahead of print. doi: 10.1111/all.14254
  35. Liu S, Stolz DB, Sappington PL, Macias CA, Killeen ME, et al. HMGB1 is secreted by immunostimulated enterocytes and contributes to cytomix-induced hyperpermeability of Caco-2 monolayers. *Am J Physiol Cell Physiol* 2006;290:C990-9.
  36. Li H, Li J, Zhang G, Da Q, Chen L, et al. HMGB1-induced p62 overexpression promotes snail-mediated epithelial-mesenchymal transition in glioblastoma cells via the degradation of GSK-3β. *Theranostics* 2019;9:1909-22.
  37. Han S, Wang C, Qin X, Xia J, Wu A. LPS alters the immuno-phenotype of glioma and glioma stem-like cells and induces in vivo antitumor immunity via TLR4. *J Exp Clin Cancer Res* 2017;36:83.
  38. Pozuelo-Rubio M. Proteomic and biochemical analysis of 14-3-3-binding proteins during C2-ceramide-induced apoptosis. *FEBS J* 2010;277:3321-42.
  39. Jin CJ, Engstler AJ, Sellmann C, Ziegenhardt D, Landmann M, et al. Sodium butyrate protects mice from the development of the early signs of non-alcoholic fatty liver disease: role of melatonin and lipid peroxidation. *Br J Nutr* 2016;1-12.
  40. Mazurek M, Litak J, Kamieniak P, Osuchowska I, Maciejewski R, et al. Micro RNA molecules as modulators of treatment resistance, immune checkpoints controllers and sensitive biomarkers in glioblastoma multiforme. *Int J Mol Sci* 2020;21:E1507.
  41. Wang X, Tian G, Li Z, Zheng L. The crosstalk between miRNA and mammalian circadian clock. *Curr Med Chem* 2015;22:1582-8.
  42. Reszka E, Zienolddiny S. Epigenetic basis of circadian rhythm disruption in cancer. *Methods Mol Biol* 2018;1856:173-201.
  43. Bai J, Chen WB, Zhang XY, Kang XN, Jin LJ, et al. HIF-2α regulates CD44 to promote cancer stem cell activation in triple-negative breast cancer via PI3K/AKT/mTOR signaling. *World J Stem Cells* 2020;12:87-99.
  44. Conner C, Lager TW, Guldner IH, Wu MZ, Hishida Y, et al. Cell surface GRP78 promotes stemness in normal and neoplastic cells. *Sci Rep* 2020;10:3474.
  45. Si D, Yin F, Peng J, Zhang G. High expression of CD44 predicts a poor prognosis in glioblastomas. *Cancer Manag Res* 2020;12:769-75.
  46. Yang LW, Wu XJ, Liang Y, Ye GQ, Che YC, et al. miR-155 increases stemness and decitabine resistance in triple-negative breast cancer cells by inhibiting TSPAN5. *Mol Carcinog* 2020;59:447-61.
  47. Wu W, Yu T, Wu Y, Tian W, Zhang J, et al. The miR155HG/miR-185/ANXA2 loop contributes to glioblastoma growth and progression. *J Exp Clin Cancer Res* 2019;38:133.
  48. Tang H, Liu Q, Liu X, Ye F, Xie X, et al. Plasma miR-185 as a predictive biomarker for prognosis of malignant glioma. *J Cancer Res Ther* 2015;11:630-4.
  49. Onishi M, Ichikawa T, Kurozumi K, Inoue S, Maruo T, et al. Annexin A2 regulates angiogenesis and invasion phenotypes of malignant glioma. *Brain Tumor Pathol* 2015;32:184-94.
  50. Christensen MV, Høgdall CK, Jochumsen KM, Høgdall EVS. Annexin A2 and cancer: a systematic review. *Int J Oncol* 2018;52:5-18.
  51. Chen C, Ling MY, Lin FH, Xu L, Lv ZM. Melatonin appears to protect against steroidogenic collapse in both mice fed with high-fat diet and H2O2-treated TM3 cells. *Andrologia* 2019;51:e13323.
  52. Akbarzadeh M, Movassaghpour AA, Ghanbari H, Kheirandish M, Fathi Maroufi N, et al. The potential therapeutic effect of melatonin on human ovarian cancer by inhibition of invasion and migration of cancer stem cells. *Sci Rep* 2017;7:17062.
  53. Gonçalves Ndo N, Colombo J, Lopes JR, Gelaleti GB, Moschetta MG, et al. Effect of melatonin in epithelial mesenchymal transition markers and invasive properties of breast cancer stem cells of canine and human cell lines. *PLoS One* 2016;11:e0150407.
  54. Casado J, Iñigo-Chaves A, Jiménez-Ruiz SM, Ríos-Arrabal S, Carazo-Gallego Á, et al. AA-NAT, MT1 and MT2 correlates with cancer

- stem-like cell markers in colorectal cancer: study of the influence of stage and p53 status of tumors. *Int J Mol Sci* 2017;18:E1251.
55. Zheng X, Pang B, Gu G, Gao T, Zhang R, et al. Melatonin inhibits glioblastoma stem-like cells through suppression of EZH2-NOTCH1 signaling axis. *Int J Biol Sci* 2017;13:245-53.
  56. Sung GJ, Kim SH, Kwak S, Park SH, Song JH, et al. Inhibition of TFEB oligomerization by co-treatment of melatonin with vorinostat promotes the therapeutic sensitivity in glioblastoma and glioma stem cells. *J Pineal Res* 2019;66:e12556.
  57. Chen X, Hao A, Li X, Du Z, Li H, et al. Melatonin inhibits tumorigenicity of glioblastoma stem-like cells via the AKT-EZH2-STAT3 signaling axis. *J Pineal Res* 2016;61:208-17.
  58. Martín V, Sanchez-Sanchez AM, Puente-Moncada N, Gomez-Lobo M, Alvarez-Vega MA, et al. Involvement of autophagy in melatonin-induced cytotoxicity in glioma-initiating cells. *J Pineal Res* 2014;57:308-16.
  59. Ogawa D, Ansari K, Nowicki MO, Salińska E, Bronisz A, et al. MicroRNA-451 Inhibits migration of glioblastoma while making it more susceptible to conventional therapy. *Noncoding RNA* 2019;5:E25.
  60. Gal H, Pandi G, Kanner AA, Ram Z, Lithwick-Yanai G, et al. MIR-451 and Imatinib mesylate inhibit tumor growth of Glioblastoma stem cells. *Biochem Biophys Res Commun* 2008;376:86-90.
  61. Bernard M, Voisin P. Photoreceptor-specific expression, light-dependent localization, and transcriptional targets of the zinc-finger protein Yin Yang 1 in the chicken retina. *J Neurochem* 2008;105:595-604.
  62. Lu ZJ, Liu SY, Yao YQ, Zhou YJ, Zhang S, et al. The effect of miR-7 on behavior and global protein expression in glioma cell lines. *Electrophoresis* 2011;32:3612-20.
  63. Yao GY, Zhu Q, Xia J, Chen FJ, Huang M, et al. Ischemic postconditioning confers cerebroprotection by stabilizing VDACS after brain ischemia. *Cell Death Dis* 2018;9:1033.
  64. Lee YS, Lee JK, Bae Y, Lee BS, Kim E, et al. Suppression of 14-3-3γ-mediated surface expression of ANO1 inhibits cancer progression of glioblastoma cells. *Sci Rep* 2016;6:26413.
  65. Hashemi M, Zali A, Hashemi J, Oraee-Yazdani S, Akbari A. Down-regulation of 14-3-3 zeta sensitizes human glioblastoma cells to apoptosis induction. *Apoptosis* 2018;23:616-25.
  66. Sarvagalla S, Kolapalli SP, Vallabhapurapu S. The two sides of YY1 in cancer: a friend and a foe. *Front Oncol* 2019;9:1230.
  67. Sinkevicius KW, Kriegel C, Bellaria KJ, Lee J, Lau AN, et al. Neurotrophin receptor TrkB promotes lung adenocarcinoma metastasis. *Proc Natl Acad Sci U S A* 2014;111:10299-304.
  68. Jiang W, Zhao S, Shen J, Guo L, Sun Y, et al. The MiR-135b-BMAL1-YY1 loop disturbs pancreatic clockwork to promote tumorigenesis and chemoresistance. *Cell Death Dis* 2018;9:149.
  69. Li J, Song J, Guo F. miR-186 reverses cisplatin resistance and inhibits the formation of the glioblastoma-initiating cell phenotype by degrading Yin Yang 1 in glioblastoma. *Int J Mol Med* 2019;43:517-24.
  70. Perekatt AO, Valdez MJ, Davila M, Hoffman A, Bonder EM, et al. YY1 is indispensable for Lgr5+ intestinal stem cell renewal. *Proc Natl Acad Sci U S A* 2014;111:7695-700.
  71. Ji K, Zheng J, Lv J, Xu J, Ji X, et al. Skeletal muscle increases FGF21 expression in mitochondrial disorders to compensate for energy metabolic insufficiency by activating the mTOR-YY1-PGC1α pathway. *Free Radic Biol Med* 2015;84:161-70.
  72. de Groot JF, Fuller G, Kumar AJ, Piao Y, Eterovic K, et al. Tumor invasion after treatment of glioblastoma with bevacizumab: radiographic and pathologic correlation in humans and mice. *Neuro Oncol* 2010;12:233-42.
  73. Nandi S, Liang G, Sindhava V, Angireddy R, Basu A, et al. YY1 control of mitochondrial-related genes does not account for regulation of immunoglobulin class switch recombination in mice. *Eur J Immunol* 2020; epub ahead of print. doi: 10.1002/eji.201948385
  74. Park A, Lee J, Mun S, Kim DJ, Cha BH, et al. Identification of transcription factor YY1 as a regulator of a prostate cancer-specific pathway using proteomic analysis. *J Cancer* 2017;8:2303-11.
  75. de Assis LVM, Kinker GS, Moraes MN, Markus RP, Fernandes PA, et al. Expression of the circadian clock gene bmal1 positively correlates with antitumor immunity and patient survival in metastatic melanoma. *Front Oncol* 2018;8:185.
  76. Takenaka MC, Gabriely G, Rothhammer V, Mascanfroni ID, Wheeler MA, et al. Control of tumor-associated macrophages and T cells in glioblastoma via AHR and CD39. *Nat Neurosci* 2019;22:729-40.
  77. Muxel SM, Pires-Lapa MA, Monteiro AW, Cecon E, Tamura EK, et al. NF-κB drives the synthesis of melatonin in RAW 264.7 macrophages by inducing the transcription of the arylalkylamine-N-acetyltransferase (AA-NAT) gene. *PLoS One* 2012;7:e52010.
  78. Pires-Afonso Y, Niclou SP, Michelucci A. Revealing and Harnessing Tumour-Associated Microglia/Macrophage Heterogeneity in Glioblastoma. *Int J Mol Sci* 2020;21:E689.
  79. Leite DM, Zvar Baskovic B, Civita P, Neto C, Gumbleton M, et al. A human co-culture cell model incorporating microglia supports glioblastoma growth and migration, and confers resistance to cytotoxics. *FASEB J* 2020;34:1710-27.
  80. Buonfiglioli A, Efe IE, Guneykaya D, Ivanov A, Huang Y, et al. let-7 MicroRNAs regulate microglial function and suppress glioma growth through toll-like receptor 7. *Cell Rep* 2019;29:3460-71.e7.
  81. Huang M, Zhang D, Wu JY, Xing K, Yeo E, et al. Wnt-mediated endothelial transformation into mesenchymal stem cell-like cells induces chemoresistance in glioblastoma. *Sci Transl Med* 2020;12:eaay7522.
  82. Luchetti F, Canonico B, Bartolini D, Arcangeletti M, Ciffolilli S, et al. Melatonin regulates mesenchymal stem cell differentiation: a review. *J Pineal Res* 2014;56:382-97.
  83. Yuan Y, Ye HQ, Ren QC. Upregulation of the BDNF/TrkB pathway promotes epithelial-mesenchymal transition, as well as the migration and invasion of cervical cancer. *Int J Oncol* 2018;52:461-72.
  84. Tajbakhsh A, Mokhtari-Zaer A, Rezaee M, Afzaljavan F, Rivandi M, et al. Therapeutic potentials of BDNF/TrkB in breast cancer; current status and perspectives. *J Cell Biochem* 2017;118:2502-15.

85. Lee JH, Lee JE, Kahng JY, Kim SH, Park JS, et al. Human glioblastoma arises from subventricular zone cells with low-level driver mutations. *Nature* 2018;560:243-7.
86. Brandao M, Simon T, Critchley G, Giamas G. Astrocytes, the rising stars of the glioblastoma microenvironment. *Glia* 2019;67:779-90.
87. Henrik Heiland D, Ravi VM, Behringer SP, Frenking JH, Wurm J, et al. Tumor-associated reactive astrocytes aid the evolution of immunosuppressive environment in glioblastoma. *Nat Commun* 2019;10:2541.
88. Kast RE, Hill QA, Wion D, Mellstedt H, Focosi D, et al. Glioblastoma-synthesized G-CSF and GM-CSF contribute to growth and immunosuppression: potential therapeutic benefit from dapsone, fenofibrate, and ribavirin. *Tumour Biol* 2017;39:1010428317699797.
89. Nehser M, Dark J, Schweitzer D, Campbell M, Zwicker J, et al. System Xc<sup>-</sup> antiporter inhibitors: azo-linked amino-naphthyl-sulfonate analogues of sulfasalazine. *Neurochem Res* 2019; epub ahead of print. doi: 10.1007/s11064-019-02901-6
90. Mega A, Hartmark Nilsen M, Leiss LW, Tobin NP, Miletic H, et al. Astrocytes enhance glioblastoma growth. *Glia* 2020;68:316-27.
91. Anderson G, Maes M. Local melatonin regulates inflammation resolution: a common factor in neurodegenerative, psychiatric and systemic inflammatory disorders. *CNS Neurol Disord Drug Targets* 2014;13:817-27.
92. Hara S, Nakashima S, Kiyono T, Sawada M, Yoshimura S, et al. p53-Independent ceramide formation in human glioma cells during gamma-radiation-induced apoptosis. *Cell Death Differ* 2004;11:853-61.
93. Doan NB, Nguyen HS, Al-Gizawiy MM, Mueller WM, Sabbadini RA, et al. Acid ceramidase confers radioresistance to glioblastoma cells. *Oncol Rep* 2017;38:1932-40.
94. Doan NB, Alhajala H, Al-Gizawiy MM, Mueller WM, Rand SD, et al. Acid ceramidase and its inhibitors: a de novo drug target and a new class of drugs for killing glioblastoma cancer stem cells with high efficiency. *Oncotarget* 2017;8:112662-74.
95. White-Gilbertson S, Lu P, Jones CM, Chiodini S, Hurley D, et al. Tamoxifen is a candidate first-in-class inhibitor of acid ceramidase that reduces amitotic division in polyploid giant cancer cells-Unrecognized players in tumorigenesis. *Cancer Med* 2020; epub ahead of print. doi: 10.1002/cam4.2960
96. Qu C, Ma J, Zhang Y, Han C, Huang L, et al. Estrogen receptor variant ER- $\alpha$ 36 promotes tamoxifen agonist activity in glioblastoma cells. *Cancer Sci* 2019;110:221-34.
97. Wan S, Jiang J, Zheng C, Wang N, Zhai X, et al. Estrogen nuclear receptors affect cell migration by altering sublocalization of AQP2 in glioma cell lines. *Cell Death Discov* 2018;4:49.
98. Pinacho-Garcia LM, Valdez RA, Navarrete A, Cabeza M, Segovia J, et al. The effect of finasteride and dutasteride on the synthesis of neurosteroids by glioblastoma cells. *Steroids* 2020;155:108556.
99. Dubrovka A, Hartung A, Bouchez LC, Walker JR, Reddy VA, et al. CXCR4 activation maintains a stem cell population in tamoxifen-resistant breast cancer cells through AhR signalling. *Br J Cancer* 2012;107:43-52.
100. van Schaik RH, Kok M, Sweep FC, van Vliet M, van Fessem M, et al. The CYP2C19\*2 genotype predicts tamoxifen treatment outcome in advanced breast cancer patients. *Pharmacogenomics* 2011;12:1137-46.
101. He W, Liu R, Yang SH, Yuan F. Chemotherapeutic effect of tamoxifen on temozolomide-resistant gliomas. *Anticancer Drugs* 2015;26:293-300.
102. Anderson G, Maes M. Interactions of tryptophan and its catabolites with melatonin and the alpha 7 nicotinic receptor in central nervous system and psychiatric disorders: role of the aryl hydrocarbon receptor and direct mitochondria regulation. *Int J Tryptophan Res* 2017;10:1178646917691738.
103. Mahajan-Thakur S, Bien-Möller S, Marx S, Schroeder H, Rauch BH. Sphingosine 1-phosphate (S1P) signaling in glioblastoma multiforme-a systematic review. *Int J Mol Sci* 2017;18:E2448.
104. Quint K, Stiel N, Neureiter D, Schlicker HU, Nimsky C, et al. The role of sphingosine kinase isoforms and receptors S1P1, S1P2, S1P3, and S1P5 in primary, secondary, and recurrent glioblastomas. *Tumour Biol* 2014;35:8979-89.
105. Anderson G, Maes M. Reconceptualizing adult neurogenesis: role for sphingosine-1-phosphate and fibroblast growth factor-1 in coordinating astrocyte-neuronal precursor interactions. *CNS Neurol Disord Drug Targets* 2014;13:126-36.
106. Pirmoradi L, Seyfizadeh N, Ghavami S, Zeki AA, Shojaei S. Targeting cholesterol metabolism in glioblastoma: a new therapeutic approach in cancer therapy. *J Investig Med* 2019;67:715-9.
107. Romani R, Manni G, Donati C, Pirisinu I, Bernacchioni C, et al. S1P promotes migration, differentiation and immune regulatory activity in amniotic-fluid-derived stem cells. *Eur J Pharmacol* 2018;833:173-82.
108. Mendoza A, Fang V, Chen C, Serasinghe M, Verma A, et al. Lymphatic endothelial S1P promotes mitochondrial function and survival in naive T cells. *Nature* 2017;546:158-61.
109. Niba ET, Nagaya H, Kanno T, Tsuchiya A, Gotoh A, et al. Crosstalk between PI3 kinase/PDK1/Akt/Rac1 and Ras/Raf/MEK/ERK pathways downstream PDGF receptor. *Cell Physiol Biochem* 2013;31:905-13.
110. Al-Koussa H, Atat OE, Jaafar L, Tashjian H, El-Sibai M. The role of Rho GTPases in motility and invasion of glioblastoma cells. *Anal Cell Pathol (Amst)* 2020;2020:9274016.
111. Wang YC, Tsai CF, Chuang HL, Chang YC, Chen HS, et al. Benzyl butyl phthalate promotes breast cancer stem cell expansion via SPHK1/S1P/S1PR3 signaling. *Oncotarget* 2016;7:29563-76.
112. Wang HC, Wong TH, Wang LT, Su HH, Yu HY, et al. Aryl hydrocarbon receptor signaling promotes ORMDL3-dependent generation of sphingosine-1-phosphate by inhibiting sphingosine-1-phosphate lyase. *Cell Mol Immunol* 2019;16:783-90.
113. Fugio LB, Coeli-Lacchini FB, Leopoldino AM. Sphingolipids and mitochondrial dynamic. *Cells* 2020;9:E581.
114. Wu W, Wu Y, Mayer K, von Rosenstiel C, Schecker J, et al. Lipid peroxidation plays an important role in chemotherapeutic effects of temozolomide and the development of therapy resistance in human glioblastoma. *Transl Oncol* 2020;13:100748.
115. Wang W, He S, Zhang R, Peng J, Guo D, et al. ALDH1A1 maintains the cancer stem-like cells properties of esophageal squamous cell

- carcinoma by activating the AKT signal pathway and interacting with  $\beta$ -catenin. *Biomed Pharmacother* 2020;125:109940.
116. Gui S, Xie X, O'Neill WQ, Chatfield-Reed K, Yu JG, et al. p53 functional states are associated with distinct aldehyde dehydrogenase transcriptomic signatures. *Sci Rep* 2020;10:1097.
  117. Chen Z, Wang HW, Wang S, Fan L, Feng S, et al. USP9X deubiquitinates ALDH1A3 and maintains mesenchymal identity in glioblastoma stem cells. *J Clin Invest* 2019;129:2043-55.
  118. Cheng P, Wang J, Waghmare I, Sartini S, Coviello V, et al. FOXD1-ALDH1A3 signaling is a determinant for the self-renewal and tumorigenicity of mesenchymal glioma stem cells. *Cancer Res* 2016;76:7219-30.
  119. Wu W, Schecker J, Würstle S, Schneider F, Schönfelder M, et al. Aldehyde dehydrogenase 1A3 (ALDH1A3) is regulated by autophagy in human glioblastoma cells. *Cancer Lett* 2018;417:112-23.
  120. Suwala AK, Koch K, Rios DH, Aretz P, Uhlmann C, et al. Inhibition of Wnt/beta-catenin signaling downregulates expression of aldehyde dehydrogenase isoform 3A1 (ALDH3A1) to reduce resistance against temozolomide in glioblastoma in vitro. *Oncotarget* 2018;9:22703-16.
  121. Sullivan KE, Rojas K, Cerione RA, Nakano I, Wilson KF. The stem cell/cancer stem cell marker ALDH1A3 regulates the expression of the survival factor tissue transglutaminase, in mesenchymal glioma stem cells. *Oncotarget* 2017;8:22325-43.
  122. Kawakami R, Mashima T, Kawata N, Kumagai K, Migita T, et al. ALDH1A3-mTOR axis as a therapeutic target for anticancer drug-tolerant persister cells in gastric cancer. *Cancer Sci* 2020;111:962-73.
  123. Park J, Shim JK, Kang JH, Choi J, Chang JH, et al. Regulation of bioenergetics through dual inhibition of aldehyde dehydrogenase and mitochondrial complex I suppresses glioblastoma tumorspheres. *Neuro Oncol* 2018;20:954-65.
  124. Yamazaki H, Nishiguchi K, Miyamoto R, Nakanishi S. Circadian rhythms in the activities of brain and liver aldehyde dehydrogenase isozymes in mice. *Life Sci* 1986;38:515-20.
  125. Matsunaga N, Ogino T, Hara Y, Tanaka T, Koyanagi S, et al. Optimized dosing schedule based on circadian dynamics of mouse breast cancer stem cells improves the antitumor effects of aldehyde dehydrogenase inhibitor. *Cancer Res* 2018;78:3698-708.
  126. Li A, Lin X, Tan X, Yin B, Han W, et al. Circadian gene Clock contributes to cell proliferation and migration of glioma and is directly regulated by tumor-suppressive miR-124. *FEBS Lett* 2013;587:2455-60.
  127. Dong Z, Zhang G, Qu M, Gimple RC, Wu Q, et al. Targeting glioblastoma stem cells through disruption of the circadian clock. *Cancer Discov* 2019;9:1556-73.
  128. Beker MC, Caglayan B, Caglayan AB, Kelestemur T, Yalcin E, et al. Interaction of melatonin and Bmal1 in the regulation of PI3K/AKT pathway components and cellular survival. *Sci Rep* 2019;9:19082.
  129. Wang Y, Lv D, Liu W, Li S, Chen J, et al. Disruption of the circadian clock alters antioxidative defense via the SIRT1-BMAL1 pathway in 6-OHDA-induced models of Parkinson's disease. *Oxid Med Cell Longev* 2018;2018:4854732.
  130. Agnihotri S, Zadeh G. Metabolic reprogramming in glioblastoma: the influence of cancer metabolism on epigenetics and unanswered questions. *Neuro Oncol* 2016;18:160-72.
  131. Kuramoto K, Yamamoto M, Suzuki S, Sanomachi T, Togashi K, et al. Verteporfin inhibits oxidative phosphorylation and induces cell death specifically in glioma stem cells. *FEBS J* 2019; epub ahead of print. doi: 10.1111/febs.15187
  132. Yoo DY, Nam SM, Kim W, Lee CH, Won MH, et al. N-acetylserotonin increases cell proliferation and differentiating neuroblasts with tertiary dendrites through upregulation of brain-derived neurotrophic factor in the mouse dentate gyrus. *J Vet Med Sci* 2011;73:1411-6.
  133. Huo X, Wang C, Yu Z, Peng Y, Wang S, et al. Human transporters, PEPT1/2, facilitate melatonin transportation into mitochondria of cancer cells: an implication of the therapeutic potential. *J Pineal Res* 2017;62.
  134. Seo SB, Lee JJ, Yun HH, Im CN, Kim YS, et al. 14-3-3 $\beta$  depletion drives a senescence program in glioblastoma cells through the ERK/SKP2/p27 pathway. *Mol Neurobiol* 2018;55:1259-70.
  135. Tan DX, Manchester LC, Liu X, Rosales-Corral SA, Acuna-Castroviejo D, et al. Mitochondria and chloroplasts as the original sites of melatonin synthesis: a hypothesis related to melatonin's primary function and evolution in eukaryotes. *J Pineal Res* 2013;54:127-38.
  136. Zhang W, Sun Y, Liu L, Li Z. Prognostic significance of TNFR-associated factor 1 and 2 (TRAF1 and TRAF2) in glioblastoma. *Med Sci Monit* 2017;23:4506-12.
  137. Chen S, Yang J, Yang L, Zhang Y, Zhou L, et al. Ubiquitin ligase TRAF2 attenuates the transcriptional activity of the core clock protein BMAL1 and affects the maximal Per1 mRNA level of the circadian clock in cells. *FEBS J* 2018;285:2987-3001.
  138. Jaeger C, Khazaal AQ, Xu C, Sun M, Krager SL, et al. Aryl hydrocarbon receptor deficiency alters circadian and metabolic rhythmicity. *J Biol Rhythms* 2017;32:109-20.
  139. Ye X, Yu L, Zuo D, Zhang L, Zu J, et al. Activated mGluR5 protects BV2 cells against OGD/R induced cytotoxicity by modulating BDNF-TrkB pathway. *Neurosci Lett* 2017;654:70-9.
  140. Zapletal O, Tylichová Z, Neča J, Kohoutek J, Machala M, et al. Butyrate alters expression of cytochrome P450 1A1 and metabolism of benzo[a]pyrene via its histone deacetylase activity in colon epithelial cell models. *Arch Toxicol* 2017;91:2135-50.
  141. Costanza M, Finocchiaro G. Allergic signs in glioma pathology: current knowledge and future perspectives. *Cancers (Basel)* 2019;11:E404.
  142. Attarha S, Roy A, Westermarck B, Tchougounova E. Mast cells modulate proliferation, migration and stemness of glioma cells through downregulation of GSK3 $\beta$  expression and inhibition of STAT3 activation. *Cell Signal* 2017;37:81-92.
  143. Maldonado MD, Mora-Santos M, Naji L, Carrascosa-Salmoral MP, Naranjo MC, et al. Evidence of melatonin synthesis and release by mast cells. Possible modulatory role on inflammation. *Pharmacol Res* 2010;62:282-7.
  144. Piao L, Feng Y, Yang Z, Qi W, Li H, et al. LETM1 is a potential cancer stem-like cell marker and predicts poor prognosis in colorectal adenocarcinoma. *Pathol Res Pract* 2019;215:152437.
  145. Lupo D, Vollmer C, Deckers M, Mick DU, Tews I, et al. Mdm38 is a 14-3-3-like receptor and associates with the protein synthesis

- machinery at the inner mitochondrial membrane. *Traffic* 2011;12:1457-66.
146. Agnihotri S, Golbourn B, Huang X, Remke M, Younger S, et al. PINK1 is a negative regulator of growth and the warburg effect in glioblastoma. *Cancer Res* 2016;76:4708-19.
  147. Li X, Spelat R, Bartolini A, Cesselli D, Ius T, et al. Mechanisms of malignancy in glioblastoma cells are linked to MCU upregulation and higher intracellular calcium level. *J Cell Sci* 2020;133:jcs237503.
  148. Dionigi L, Ragonese F, Monarca L, Covino S, de Luca A, et al. Focus on the use of resveratrol as adjuvant in glioblastoma therapy. *Curr Pharm Des* 2020; epub ahead of print. doi: 10.2174/1381612826666200401085634
  149. Lai SW, Liu YS, Lu DY, Tsai CF. Melatonin modulates the microenvironment of glioblastoma multiforme by targeting sirtuin 1. *Nutrients* 2019;11:E1343.
  150. Moretti E, Favero G, Rodella LF, Rezzani R. Melatonin's antineoplastic potential against glioblastoma. *Cells* 2020;9:E599.
  151. Beccaria K, Canney M, Bouchoux G, Desseaux C, Grill J, et al. Ultrasound-induced blood-brain barrier disruption for the treatment of gliomas and other primary CNS tumors. *Cancer Lett* 2020;479:13-22.
  152. Han YS, Lee JH, Lee SH. Melatonin suppresses ischemia-induced fibrosis by regulating miR-149. *Biochem Biophys Res Commun* 2020;525:354-9.
  153. Ghasemi A, Fallah S, Ansari M. MicroRNA-149 is epigenetically silenced tumor-suppressive microRNA, involved in cell proliferation and downregulation of AKT1 and cyclinD1 in human glioblastoma multiforme. *Biochem Cell Biol* 2016;94:569-76.
  154. She X, Yu Z, Cui Y, Lei Q, Wang Z, et al. miR-128 and miR-149 enhance the chemosensitivity of temozolomide by Rap1B-mediated cytoskeletal remodeling in glioblastoma. *Oncol Rep* 2014;32:957-64.